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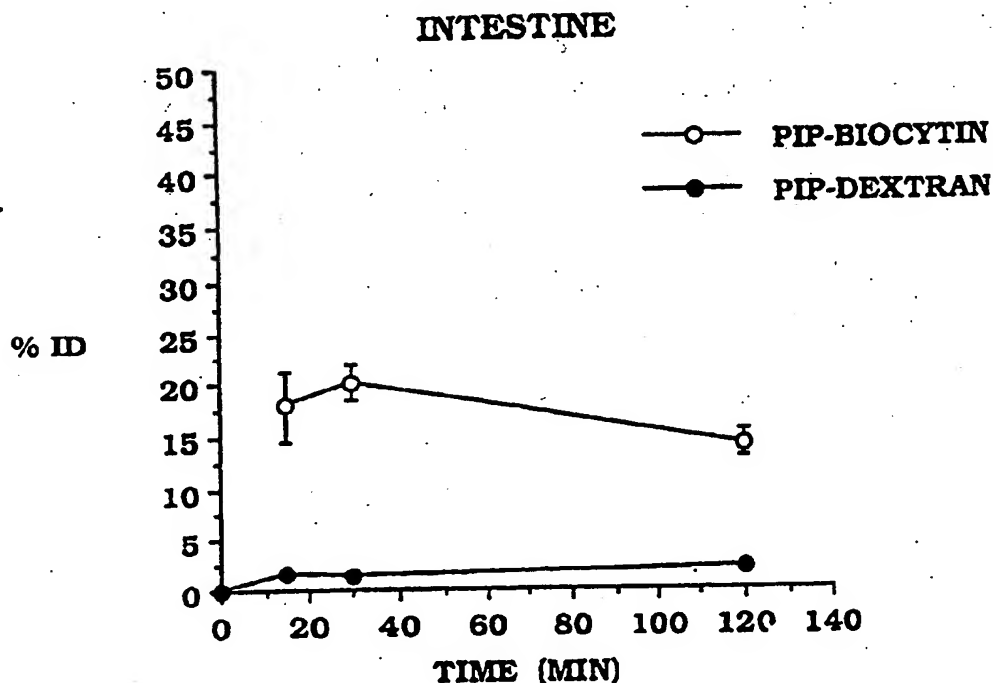
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(54) Title: DIRECTED BIODISTRIBUTION OF SMALL MOLECULES



(57) Abstract

The present invention provides methods for directing the biodistribution of molecules that are not generally specifically excreted via the renal pathway to renal excretion. The methods employ conjugates or complexes containing a directed biodistribution molecule (DBM) and one or more bound molecules, wherein the biodistribution of the conjugate or complex is directed to renal excretion *in vivo* by the DBM component thereof.

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DIRECTED BIODISTRIBUTION OF SMALL MOLECULES

10 Technical Field of the Invention

 The present invention is directed to directed
biodistribution molecules (DBMs) that direct the
biodistribution of administered small molecules to
improve the performance thereof in diagnostic or
15 therapeutic protocols. More specifically, the present
invention is directed to DBMs that mask or override the
biodistribution properties of small molecules, such that
the small molecule excretion mimics DBM excretion.
Directed biodistribution to renal excretion as well as
20 DBM/small molecule conjugates or complexes designed for
renal excretion are discussed.

Background of the Invention

 Radiolabeled antibodies are exemplary targeted
25 moieties that are used in a variety of diagnostic and
therapeutic medical procedures. The increased
specificity of monoclonal antibodies, compared to
polyclonal antibodies, makes them more useful for
delivering diagnostic or therapeutic agents such as
30 radioisotopes to desired target sites in vivo. A
monoclonal antibody specific for a desired type of
target cells (e.g., tumor cells) may be used to deliver
a therapeutic radionuclide-antibody conjugate to target
cells, thereby causing the eradication of the undesired
35 target cells. Alternatively, a monoclonal antibody
having a diagnostically effective radionuclide attached
thereto may be administered, whereupon the radiolabeled
antibody localizes to target tissue. Conventional
diagnostic procedures then may be used to detect the

presence of target sites within the patient.

One method for radiolabeling proteins such as antibodies involves attachment of radionuclide metal chelates thereto. Chelates having a variety of chemical structures have been developed for this purpose. The usefulness of such chelates is dependent upon a number of factors including the stability of radionuclide binding within the chelate and the reactivity of the chelate with the desired protein. The efficiency of the radiolabeling process producing the radionuclide metal chelate is also important. Another consideration is the biodistribution of the radiolabeled antibody and catabolites thereof in vivo.

Image contrast achievable in diagnostic applications of this technology is limited by the ratio of radioactivity localized to the tumor and radioactivity associated with normal tissue including blood. A limitation in the use of radioimmunotherapy in the treatment of cancer is dose limiting toxicity to normal organs including the radiosensitive bone marrow. Biodistribution studies in mice have shown that compounds, such as radiolabeled small molecules, are rapidly cleared from the blood, taken up by the liver and excreted into the intestines. Such hepatobiliary excretion poses additional problems with respect to conventionally labeled antibodies, because antibody-chelate-radionuclide conjugates are metabolically degraded to form stable chelate-lysine adducts that show significant intestinal accumulation.

Certain peptides (e.g., somatostatin and derivatives thereof) have been employed as targeting agents to direct active agents to protein receptor-rich tumor targets. Hepatobiliary clearance of these radiolabeled peptides is also problematic. Also, radiolabeled ligand (e.g., biotin) derivatives used in pretargeting protocols have also exhibited undesirable hepatobiliary excretion.

Summary of the Invention

The present invention provides methods for directing the biodistribution of molecules that are not generally specifically excreted via the renal pathway to renal excretion. The methods employ conjugates or complexes as described below.

The present invention provides conjugates or complexes containing a directed biodistribution molecule (DBM) and one or more bound molecules, wherein the biodistribution of the conjugate or complex is directed to renal excretion in vivo by the DBM component thereof. Small molecules, including targeting moieties, such as oligonucleotides and peptides, drugs, ligands, anti-ligands and chelates, are characterized by improved biodistribution characteristics when conjugated to DBMs in accordance with the present invention. Preferably, conjugates or complexes of the present invention include a diagnostic or therapeutic active agent. Optionally, such conjugates or complexes exhibiting improved biodistribution in accordance with the present invention may include larger components, such as an antibody, an antibody fragment, an analog or derivative thereof, a bispecific antibody, a peptide, an oligopeptide, a moiety that binds to a natural receptor, enzymes, or like moieties. Molecules incorporating combinations of the aforementioned components (e.g., radionuclide-chelate-ligand, radionuclide-chelate-anti-ligand, antibody-chelate-radionuclide, antibody-ligand-drug, antibody-anti-ligand-drug, antibody-ligand-chelate-radionuclide, antibody-anti-ligand-chelate-radionuclide and the like) are useful in the practice of the present invention. The improved biodistribution properties of DBM-containing complexes or conjugates of the present invention are imparted by covalent binding, non-covalent complexing or otherwise associating a small molecule component (including one or more small molecules) to or with a DBM component exhibiting such improved

characteristics.

One DBM property that facilitates the favorable biodistribution thereof is size. In one embodiment of the present invention, one or more small molecules are bound to or associated with a large (e.g., polymeric) DBM, wherein the DBM masks the small molecule such that the biodistribution of the small molecule substantially corresponds to that of the DBM, while maintaining any binding properties of the small molecule(s) necessary for proper targeting. More specifically, the biodistribution properties of the small molecule are blocked or overridden by those of the DBM. For instance, sites on the small molecule recognized, for example, by liver cells are rendered sterically unavailable for liver cell binding or otherwise fail to be recognized by liver cells as a result of the DBM structure or DBM-small molecule(s) conjugate or complex configuration.

Another benefit imparted by size is the ability to bind a plurality of active agents. Increasing the concentration of active agent at the target enhances the efficacy of the active agent and the protocol in which it is used. Polymeric and other intermediate or high molecular weight DBMs, having a plurality of available functional groups for direct or indirect active agent binding or exhibiting the capability of derivitization to acquire such functional groups, are exemplary multiple active agent binding DBMs. Preferred DBMs having this characteristic, which also impart renal clearance properties to associated small molecules, include dextran, amino-ethyl dextran, carboxymethyl cellulose, hydroxyethyl cellulose, 2-hydroxyethyl methacrylate polymers, hydroxy-propylmethacrylamide polymers (HPMA), carboxymethyl dextrans, polyethyleneglycol (PEG), polyaspartate, polyglutamate and succinylated polylysine. A more preferred DBM incorporates dextran, such preferred molecules including

biotin-dextran, lysine fixable (BDLF) and other aminodextran-based molecules. Preferred DBM-small molecule conjugates useful in the practice of the present invention are directly or indirectly (i.e., through a chelate) radiolabeled biotin-dextran, somatostatin- or somatostatin derivative-dextran as well as antibody- or antibody fragment-dextran.

Another property of DBMs that influences the biodistribution thereof is charge. Consequently, conjugates or complexes of the present invention exhibiting improved biodistribution characteristics include small molecule(s) bound to, complexed with or associated with a highly polar molecule having desirable renal clearance biodistribution characteristics. This desirable property is imparted to the small molecule as a consequence of conjugation or complexation of the small molecule to the DBM. Preferred DBM-small molecule conjugates of this aspect of the present invention include diethylene triamine penta-acetic acid (DTPA) or derivatives or analogs thereof bound to a targeting moiety, such as an antibody, antibody fragment, oligonucleotide or other small proteinaceous molecule, a ligand, an anti-ligand, a stable chelate such as an N_2S_2 mercaptodiamide molecule, or a combination thereof. More preferred DBM-small molecule conjugates or complexes of this aspect of the present invention are radiolabeled DTPA derivative-antibody or -antibody fragment, DTPA derivative-somatostatin or -somatostatin derivative and DTPA derivative-biotin.

DBM-small molecule complexes or conjugates of the present invention are useful in the delivery of a variety of therapeutic and diagnostic active agents to target cell populations using targeted, direct labeled protocols (e.g., administration of targeting moiety-DBM-active agent or active agent-targeting moiety-DBM-conjugate or complex) and pretargeting approaches (e.g., administration of targeting moiety-anti-ligand followed

by administration of ligand-DBM-active agent or active agent-ligand-DBM conjugate or complex). Such targeted, direct-labeled diagnostic and therapeutic protocols and diagnostic and therapeutic pretargeting protocols are also contemplated by the present invention. Drugs, diagnostic radionuclides and therapeutic radionuclides are preferred active agents for delivery in accordance with the present invention.

An additional aspect of the present invention provides renal excretion of active agent-containing conjugate or complex metabolites. More specifically, the present invention provides chelate-DBM-proteinaceous targeting moiety conjugates or complexes which are metabolized to form amino acid adducts of the chelate-DBM portion of the conjugate or complex. These adducts are excreted by a renal pathway as a result of the DBM component thereof.

Brief Description of the Drawings

Figure 1A depicts the percent injected dose of radiolabeled PIP-biocytn and biotin-dextran in the intestine over time.

Figure 1B depicts the percent injected dose of radiolabeled PIP-biocytn and biotin-dextran in the bladder over time.

Figure 1C depicts the percent injected dose of radiolabeled PIP-biocytn and biotin-dextran in the liver over time.

Figure 1D depicts the percent injected dose of radiolabeled PIP-biocytn and biotin-dextran in the kidney over time.

Figure 1E depicts the percent injected dose of radiolabeled PIP-biocytn and biotin-dextran in the blood over time.

Detailed Description of Preferred Embodiments

Prior to describing preferred embodiments of the

present invention, definitions of terms thought to be useful in the understanding the invention are set forth.

Chelate: Bifunctional chelates that are capable of (i) coordinately binding a metal or radiometal and (ii) covalently attaching to a DBM, targeting moiety, ligand or anti-ligand. Preferred chelates are N_xS_y chelates.

N_xS_y Chelates: Bifunctional chelators that are capable of (i) coordinately binding a metal or radiometal and (ii) covalently attaching to a DBM, targeting moiety, ligand or anti-ligand. Particularly preferred N_xS_y chelates have N_2S_2 and N_3S cores. Exemplary N_xS_y chelates are described in Fritzberg et al., Proc. Natl. Acad. Sci. USA 85:4024-29, 1988; in Weber et al., Bioconj. Chem. 1:431-37, 1990; and in the references cited therein, for instance.

Directed Biodistribution Molecule (DBM): A moiety that is excreted through a renal pathway when administered to a mammalian recipient and that is capable of covalently or non-covalently binding to one or more antibodies or antibody fragments as well as drugs, peptides, chelates, ligands, anti-ligands or other small molecules and imposing a renal route of excretion upon the associated molecule(s). DBMs include polar molecules having a molecular weight ranging from about 3 kD to about 70 kD) or highly polar or ionic moieties having a molecular weight below about 2 kD.

Targeting moiety: A molecule that binds to a defined population of cells. The targeting moiety may bind a receptor, an oligonucleotide, an enzymatic substrate, an antigenic determinant, or other binding site present on or in the target cell population. Antibodies and peptides are used throughout the specification as prototypical examples of targeting moieties. Tumor is used as a prototypical example of a target in describing the present invention.

Ligand/anti-ligand pair: A complementary/anti-complementary set of molecules that demonstrate specific

binding, generally of relatively high affinity. Exemplary ligand/anti-ligand pairs include hapten/antibody, lectin/carbohydrate, ligand/receptor, and biotin/avidin or biotin/streptavidin. Biotin/streptavidin is used throughout the specification as a prototypical example of a ligand/anti-ligand pair.

Anti-ligand: A moiety demonstrating high affinity, and preferably, multivalent binding of the complementary ligand. Preferably, the anti-ligand is large enough to avoid rapid renal clearance, and contains sufficient multivalency to accomplish crosslinking and aggregation of targeting moiety-ligand conjugates. Univalent anti-ligands are also contemplated by the present invention. Streptavidin is used as a prototypical anti-ligand.

Streptavidin: This term includes streptavidin and avidin as well as derivatives and analogs thereof that are capable of high affinity, multivalent or univalent binding of biotin.

Ligand: A relatively small, soluble molecule that exhibits rapid serum, blood and/or whole body clearance when administered intravenously in a mammal. Biotin is used as the prototypical ligand.

Active Agent: A diagnostic or therapeutic agent, including radionuclides, drugs, and the like. Radionuclide therapeutic agents are used as prototypical active agents.

Small Molecule: A molecule having a molecular weight below about 2 kD, including one or more of the following components: a catabolite derived from targeting moieties, such as an antibody or an antibody fragment; an oligonucleotide; a peptide; a drug, a ligand, an anti-ligand, a chelate or the like. Small molecules of the present invention preferably include a diagnostic or a therapeutic active agent when administered to a mammal. For the purposes of this description, chelate, antibody metabolite, somatostatin (and derivatives thereof), biotin, radiolabeled antibody metabolite,

radiolabeled somatostatin (and somatostatin derivatives) and radiolabeled biotin are prototypical small molecules useful in the practice of the present invention.

Complex: A multi-component molecule incorporating two or more functionally different moieties that are not covalently bound. Complexes of the present invention are associated through non-covalent binding, such as ionic bonding, van der Waals force-based coupling, hydrogen bonding and the like. Exemplary complexes of the present invention incorporate one or more small molecules and a DBM. Other complexes additionally include an active agent or an antibody or an antibody fragment targeting moiety.

Conjugate: A multi-component molecule incorporating two or more functionally different moieties that are covalently bound. Exemplary conjugates of the present invention incorporate one or more small molecules and a DBM. Other conjugates additionally include an active agent or an antibody or an antibody fragment targeting moiety. For the purposes of the following description, the term "conjugate" shall include "complexes."

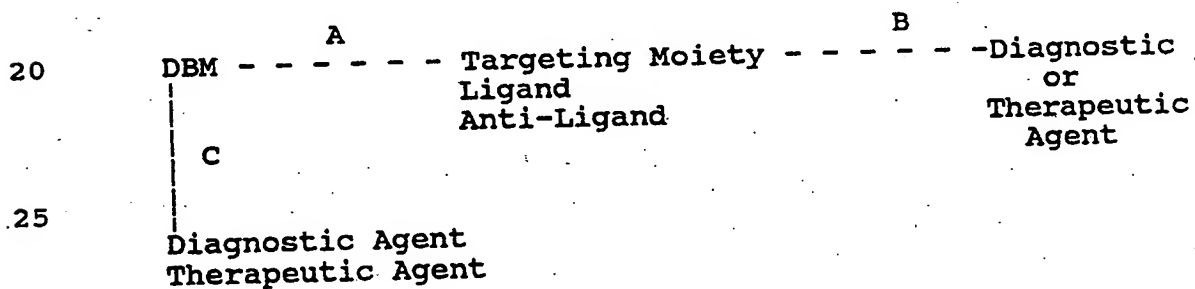
Targeted, Direct Labeled Protocol: Administration of a conjugate or a complex containing a targeting moiety and an active agent, such that the targeting moiety directs the localization of the conjugate or complex to a target cell population, and the active agent exerts a diagnostic or a therapeutic effect. DBMs of the present invention are covalently or non-covalently bound to either or both of these components and direct the biodistribution of the conjugates/complexes or active agent-containing metabolites thereof to renal excretion.

Pretargeting: Target site localization of a targeting moiety that is conjugated with one member of a ligand/anti-ligand pair; after a time period sufficient for optimal target-to-non-target accumulation of this targeting moiety conjugate, active agent

10

conjugated to the opposite member of the ligand/anti-ligand pair is administered and is bound (directly or indirectly) to the targeting moiety conjugate at the target site (two-step pretargeting). Three-step pretargeting (*i.e.*, administration of targeting moiety-ligand; anti-ligand administration; and administration of active agent-ligand) and other related methods described herein are also encompassed.

The present invention is directed to DBM-small molecule conjugates characterized by renal excretion. Preferably, the conjugates of the present invention deliver a diagnostic or therapeutic agent to a target cell population determined by the binding specificity of a targeting moiety, ligand or anti-ligand conjugate component. Consequently, preferred embodiments of the invention can be represented as follows:



where either bond "B" or bond "C" links the active agent to the conjugate.

DBM-small molecule conjugates of the present invention are useful in a variety of therapeutic and diagnostic protocols. Preferred protocols may be generally characterized as targeted, direct-labeled and pretargeting approaches. Typically, a targeted, direct-labeled approach involves administering to a mammalian recipient an agent selected from those schematically represented below, with molecules of the first two structures preferred.

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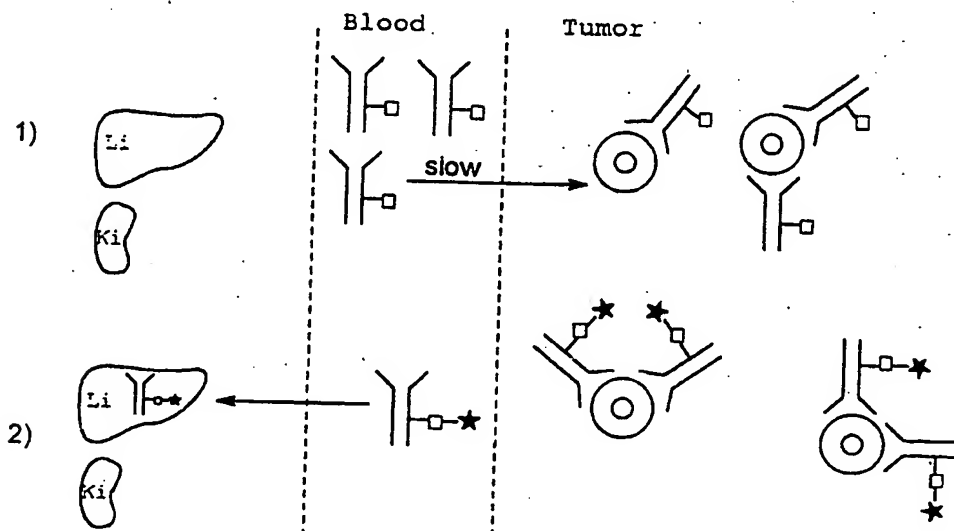
Targeting Moiety - - - - DBM - - - - Active Agent
or

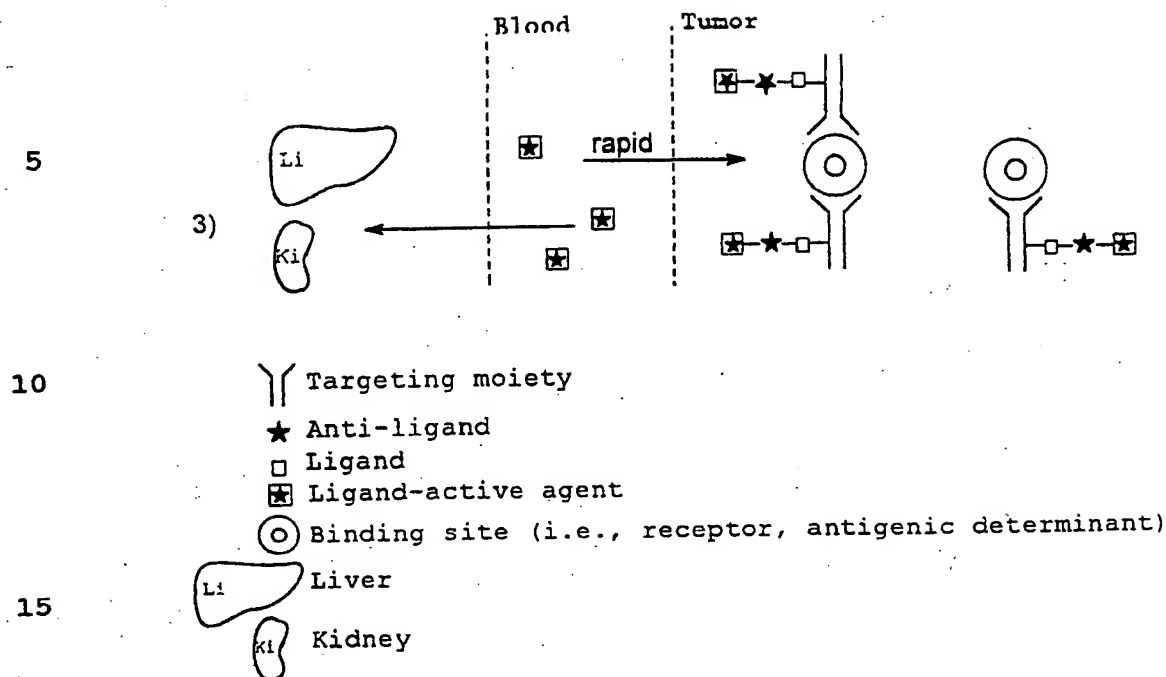
Active Agent - - - - Targeting Moiety - - - - DBM
or

5 Targeting Moiety - - - - Active Agent - - - - DBM

In this manner, active agent is delivered to the target cell population to which the targeting moiety localizes, while non-targeted conjugate as well as the metabolic byproducts thereof bearing the active agent are excreted, with the excretion proceeding primarily through the renal pathway. Undesirable hepatobiliary excretion of active agent are decreased in the practice of the present invention.

15 Pretargeting encompasses two protocols, termed the three-step and the two-step. In the three-step protocol shown schematically below, targeting moiety-ligand is administered and permitted to localize to target.





20 Anti-ligand is then administered to act as a clearing agent and facilitate and direct the excretion of circulating targeting moiety-ligand. The anti-ligand also binds to target-associated targeting moiety-ligand. Next, a conjugate of the present invention is administered, preferably selected from the following

25 schematic structures:

DBM - - - - Ligand - - - - Active Agent

or

Active Agent - - - - DBM - - - - Ligand

30 or

Ligand - - - - Active Agent - - - - DBM

35 The former two structures are preferred in the practice of the present invention. The DBM/ligand/active agent conjugate either binds to target-associated targeting moiety-ligand-anti-ligand or is preferably rapidly excreted, with the excretion proceeding primarily

through the renal pathway. Consequently, the target-non-target ratio of active agent is improved, and undesirable hepatobiliary excretion and intestinal uptake of the active agent are substantially decreased.

5 Two-step pretargeting involves administration of targeting moiety-anti-ligand. After permitting the administered agent to localize to target, a conjugate of the present invention is administered, preferably selected from the aforementioned ligand-DBM-active agent or active agent-ligand-DBM conjugates of the present invention. Preferably, as a "step 1.5," a clearing agent is administered to remove circulating targeting moiety-anti-ligand without binding to target-associated targeting moiety-anti-ligand. In this manner, the target-non-target ratio of the active agent is increased, and undesirable hepatobiliary excretion and intestinal uptake of the active agent are substantially decreased.

20 The "targeting moiety" of the present invention binds to a defined target cell population, such as tumor cells. Preferred targeting moieties useful in this regard include antibody and antibody fragments, peptides, and hormones. Proteins corresponding to known cell surface receptors (including low density lipoproteins, transferrin and insulin), melanocyte stimulating hormone, somatostatin, somatostatin derivatives, such as octreotide and MK-678 (Merck), fibrinolytic enzymes, HER2 ligand, and biological response modifiers (including interleukin, interferon, erythropoietin and colony-stimulating factor) are also preferred targeting moieties. Oligonucleotides, e.g., antisense oligonucleotides that are complementary to portions of target cellular nucleic acids (DNA or RNA), are also useful as targeting moieties in the practice of the present invention. Oligonucleotides binding to cell surfaces are also useful. Analogs of the above-listed targeting moieties that retain the capacity to bind to

a defined target cell population may also be used within the claimed invention. In addition, synthetic targeting moieties having such binding specificity may be designed.

5 Functional equivalents of the aforementioned molecules are also useful as targeting moieties of the present invention. One targeting moiety functional equivalent is a "mimetic" compound, an organic chemical construct designed to mimic the proper configuration and/or orientation for targeting moiety-target cell binding. Another targeting moiety functional equivalent is a short polypeptide designated as a "minimal" polypeptide, constructed using computer-assisted molecular modeling and mutants having altered binding affinity, which minimal polypeptides exhibit the binding affinity of the larger targeting moiety.

10 Preferred targeting moieties of the present invention are antibodies (polyclonal or monoclonal), peptides, oligonucleotides or the like. Polyclonal antibodies useful in the practice of the present invention are polyclonal (Vial and Callahan, Univ. Mich. Med. Bull., 20: 284-6, 1956), affinity-purified polyclonal or fragments thereof (Chao et al., Res. Comm. in Chem. Path. & Pharm., 9: 749-61, 1974).

25 Monoclonal antibodies useful in the practice of the present invention include whole antibody and fragments thereof. Such monoclonal antibodies and fragments are producible in accordance with conventional techniques, such as hybridoma synthesis, recombinant DNA techniques and protein synthesis. Useful monoclonal antibodies and fragments may be derived from any species (including humans) or may be formed as chimeric proteins which employ sequences from more than one species. See, generally, Kohler and Milstein, Nature, 256: 495-97, 30 1975; Eur. J. Immunol., 6: 511-19, 1976.

35 Human monoclonal antibodies or "humanized" murine antibody are also useful as targeting moieties in

accordance with the present invention. For example, murine monoclonal antibody may be "humanized" by genetically recombining the nucleotide sequence encoding the murine Fv region (i.e., containing the antigen binding sites) or the complementarity determining regions thereof with the nucleotide sequence encoding a human constant domain region and an Fc region, e.g., in a manner similar to that disclosed in European Patent Application No. 0,411,893 A2. Some murine residues may also be retained within the human variable region framework domains to ensure proper target site binding characteristics. Humanized targeting moieties are recognized to decrease the immunoreactivity of the antibody or polypeptide in the host recipient, permitting, in some cases, an increase in the half-life and facilitating a reduction in the possibility of adverse immune reactions and facilitating multiple administration protocols. Retention of murine CDR regions and framework regions is preferably undertaken when "humanizing" antibodies to be used in the practice of the present invention.

Types of active agents (diagnostic or therapeutic) useful herein include drugs and radionuclides. Preferred drugs suitable for use herein include conventional chemotherapeutics, such as vinblastine, doxorubicin, bleomycin, methotrexate, 5-fluorouracil, 6-thioguanine, cytarabine, cyclophosphamide and cisplatinum, as well as other conventional chemotherapeutics as described in Cancer: Principles and Practice of Oncology, 2d ed., V.T. DeVita, Jr., S. Hellman, S.A. Rosenberg, J.B. Lippincott Co., Philadelphia, PA, 1985, Chapter 14. A preferred drug within the present invention is a trichothecene, because a plurality of drug molecules are deliverable to the target for each targeting moiety employed.

Trichothecenes are drugs produced by soil fungi of the class *Fungi imperfecti* or isolated from *Baccharus*

megapotamica (Bamburg, J.R. Proc. Molec. Subcell. Biol. 8:41-110, 1983; Jarvis & Mazzola, Acc. Chem. Res. 15:338-395, 1982). They appear to be the most toxic molecules that contain only carbon, hydrogen and oxygen (Tamm, C. Fortschr. Chem. Org. Naturst. 31:61-117, 1974). They are all reported to act at the level of the ribosome as inhibitors of protein synthesis at the initiation, elongation, or termination phases.

There are two broad classes of trichothecenes: those that have only a central sesquiterpenoid structure and those that have an additional macrocyclic ring (simple and macrocyclic trichothecenes, respectively). The simple trichothecenes may be subdivided into three groups (i.e., Group A, B, and C) as described in U.S. Patent Nos. 4,744,981 and 4,906,452 (incorporated herein by reference). Representative examples of Group A simple trichothecenes include: Scirpene, Roridin C, dihydrotrichothecene, Scirpen-4, 8-diol, Verrucarol, Scirpentriol, T-2 tetraol, pentahydroxyscirpene, 4-deacetylneosolaniol, trichodermin, deacetylcalonecetrin, calonecetrin, diacetylverrucarol, 4-monoacetoxyscirpenol, 4,15-diacetoxyscirpenol, 7-hydroxydiacetoxyscirpenol, 8-hydroxydiacetoxyscirpenol (Neosolaniol), 7,8-dihydroxydiacetoxyscirpenol, 7-hydroxy-8-acetyldiacetoxyscirpenol, 8-acetylneosolaniol, NT-1, NT-2, HT-2, T-2, and acetyl T-2 toxin. Representative examples of Group B simple trichothecenes include: Trichothecolone, Trichothecin, deoxynivalenol, 3-acetyldeoxynivalenol, 5-acetyldeoxynivalenol, 3,15-diacetyldeoxynivalenol, Nivalenol, 4-acetylnivalenol (Fusarenon-X), 4,15-diacetylnivalenol, 4,7,15-triacetylnivalenol, and tetra-acetylnivalenol. Representative examples of Group C simple trichothecenes include: Crotocol and Crotocin. Representative macrocyclic trichothecenes include Verrucarin A, Verrucarin B, Verrucarin J

(Satratoxin C), Roridin A, Roridin D, Roridin E (Satratoxin D), Roridin H, Satratoxin F, Satratoxin G, Satratoxin H, Vertisporin, Mytoxin A, Mytoxin C, Mytoxin B, Myrotoxin A, Myrotoxin B, Myrotoxin C, Myrotoxin D, Roritoxin A, Roritoxin B, and Roritoxin D. In addition, the general "trichothecene" sesquiterpenoid ring structure is also present in compounds termed "baccharins" isolated from the higher plant *Baccharis megapotamica*, and these are described in the literature, for instance as disclosed by Jarvis et al. (Chemistry of Alleopathy, ACS Symposium Series No. 268: ed. A.C. Thompson, 1984, pp. 149-159). Other highly toxic and relatively low molecular weight toxin molecules, such as palytoxin, are also preferred for use in the practice of the present invention.

Experimental drugs, such as mercaptopurine, N-methylformamide, 2-amino-1,3,4-thiadiazole, melphalan, hexamethylmelamine, gallium nitrate, 3% thymidine, dichloromethotrexate, mitoguazone, suramin, bromodeoxyuridine, iododeoxyuridine, semustine, 1-(2-chloroethyl)-3-(2,6-dioxo-3-piperidyl)-1-nitrosourea, N,N'-hexamethylene-bis-acetamide, azacitidine, dibromodulcitol, Erwinia asparaginase, ifosfamide, 2-mercaptoethane sulfonate, teniposide, taxol, 3-deazauridine, soluble Baker's antifol, homoharringtonine, cyclocytidine, acivicin, ICRF-187, spiromustine, levamisole, chlorozotocin, aziridinyll benzoquinone, spirogermanium, aclarubicin, pentostatin, PALA, carboplatin, amsacrine, caracemide, iproplatin, misonidazole, dihydro-5-azacytidine, 4'-deoxydoxorubicin, menogaril, triciribine phosphate, fazarabine, tiazofurin, teroxirone, ethiofos, N-(2-hydroxyethyl)-2-nitro-1H-imidazole-1-acetamide, mitoxantrone, acodazole, amonafide, fludarabine phosphate, pibenzimol, didemnin B, merbarone, dihydrolenperone, flavone-8-acetic acid, oxantrazole, ipomeanol, trimetrexate, deoxyspergualin, echinomycin,

and dideoxycytidine (see NCI Investigational Drugs, Pharmaceutical Data 1987, NIH Publication No. 88-2141, Revised November 1987) are also preferred.

5 Other anti-tumor agents, e.g., agents active against proliferating cells, are administrable in accordance with the present invention. Exemplary anti-tumor agents include cytokines, such as IL-2, tumor necrosis factor or the like, lectin inflammatory response promoters (selectins), such as L-selectin, E-selectin, P-selectin
10 or the like, and like molecules.

Preferred photosensitizing agents for use in the present invention are porphyrin derivatives with a strong absorption band between 600 and 700 nm (red light). Chemical modification of porphyrin compounds is
15 undertaken to enhance performance of those compounds in photodynamic therapy protocols. Phthalocyanines, synthetic porphyrins when chelated with aluminum or zinc (e.g., chloroaluminum sulfonated phthalocyanines), are effective to destroy target cells. Photofrin II, an ether/ester derivative of porphyrin, is presently the
20 most commonly employed photosensitizing agent in photodynamic therapy. Other exemplary photosensitizing agents are chlorins (e.g., chlorin e6, tin chlorin e6, bacteriochlorin A, bacteriochlorophyllin a, mono- and
25 di-L-aspartyl chlorin e6, and the like); porphyrin diethers (e.g., di-isobutyl ethers and di-hexyl ethers); purpurins (e.g., NT2), benzoporphyrin derivatives (porphines, such as isomers of 5, 10, 15, 20-tetra(hydroxyphenyl)-porphyrin; and sulfonated
30 derivatives of tetraphenylporphine, such as TPPS₂ (a derivative of TPPS₄ with two rather than four sulfonate groups) and TPPS₄ (5,10,15,20,-tetra(4-sulfonato-phenyl)-21H,23H,porphine)); and the like. Generally, these exemplary photosensitizing agents have carboxylate
35 groups available for conjugation.

Prodrugs may also be employed in the practice of the present invention, with the prodrugs being substantially

physiologically inactive until exposure to certain enzymes. An exemplary prodrug useful in the practice of the present invention N-(4'-hydroxyphenyl-acetyl)palytoxin (penicillin G amidase enzyme). See, for example, Bignami et al., Cancer Research, 52: 5759-5764, 1992. Another example of a suitable prodrug is a phosphate form of a drug, such as phenol mustard phosphate, etoposide phosphate, mitomycin phosphate, doxorubicin phosphate and the like (alkaline phosphatase enzyme). See, for example, Wallace et al., Bioconj. Chem., 2: 349-352, 1991. Another example is 5-fluorocytosine (5FC) (cytosine deaminase enzyme), described in Senter et al., Bioconj. Chem., 2: 447-451, 1991. An additional example is beta-lactam prodrugs (beta-lactamase enzyme). See, for example, Meyer et al., Bioconj. Chem., 3: 42-48, 1992.

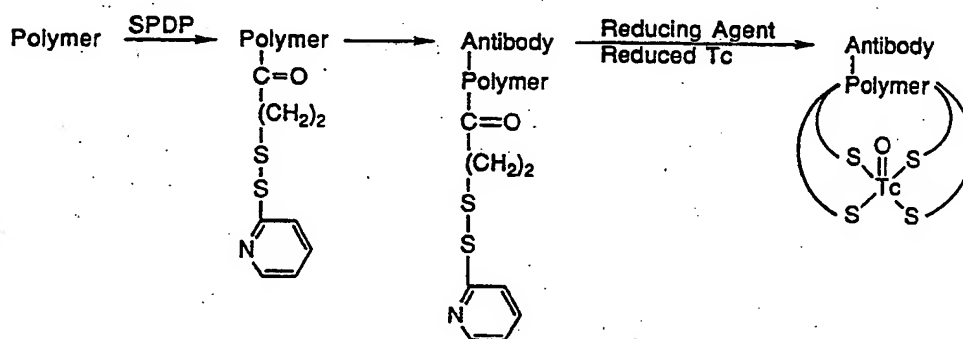
Radionuclides useful within the present invention include gamma-emitters, positron-emitters, Auger electron-emitters, X-ray emitters and fluorescence-emitters, with beta- or alpha-emitters preferred for therapeutic use. Radionuclides are well-known in the art and include ^{123}I , ^{124}I , ^{125}I , ^{130}I , ^{131}I , ^{133}I , ^{135}I , ^{47}Sc , ^{72}As , ^{72}Se , ^{90}Y , ^{88}Y , ^{97}Ru , ^{100}Pd , $^{101\text{m}}\text{Rh}$, ^{119}Sb , ^{128}Ba , ^{197}Hg , ^{211}At , ^{212}Bi , ^{153}Sm , ^{169}Eu , ^{212}Pb , ^{109}Pd , ^{111}In , ^{67}Ga , ^{68}Ga , ^{67}Cu , ^{75}Br , ^{76}Br , ^{77}Br , $^{99\text{m}}\text{Tc}$, ^{32}P , ^{11}C , ^{13}N , ^{15}O and ^{18}F . Preferred therapeutic radionuclides include ^{188}Re , ^{186}Re , ^{203}Pb , ^{212}Pb , ^{212}Bi , ^{109}Pd , ^{64}Cu , ^{67}Cu , ^{90}Y , ^{125}I , ^{131}I , ^{77}Br , ^{211}At , ^{97}Ru , ^{105}Rh , ^{198}Au and ^{199}Au or ^{177}Lu .

The radionuclides useful in the practice of the present invention are covalently bound to or are complexed with a chelate capable of stably associating with the radionuclide and with the DBM or other conjugate component. In this manner, the radionuclide active agent is directly or indirectly associated with the DBM which directs the biodistribution of the radionuclide. Exemplary chelates are N_xS_y chelates as previously discussed; para-iodo-phenyl (PIP)-based

chelates described, for example, in Wilbur, et al., J. Nucl. Med., 30: 216-226, 1989; and the like.

Radionuclide complexation is generally conducted in accordance with standard techniques therefor (e.g., pre-formed or post-formed approaches). Also, chelate binding to DBMs, ligands, anti-ligands or other conjugate components is accomplished by standard coupling procedures for formation of amides, esters, ethers and the like. Chelate-DBM conjugation is accomplished through an active ester chelate derivative, wherein exemplary active esters are N-hydroxysuccinimide, tetrafluorophenyl and the like). Michael addition reactions employing, for example, a maleimide group, can also be used for this purpose.

One aspect of the present invention is a conjugation technique characterized by minimized labeling time and defined chemistry for radiolabel attachment. The inventive synthesis is schematically shown below.



This post-formed approach of the present invention involves derivitization of a polymeric species (e.g., dextran, polylysine or the like), with a thiolating agent to incorporate thiol functional groups into the polymer structure. For example, exemplary thiolating agents include N-succinimidyl 3-(2-pyridyldithio) propionate (SPDP), iminothiolane, N-succinimidyl-S

acetylthioacetate (SATA), dimercaptosuccinyl derivatives and the like. The thiolating agent-derivitized polymer is then covalently attached to antibody, another targeting moiety, ligand, anti-ligand or the like. For example, antibody is conjugated to the derivitized polymers using a water soluble carbodiimide. Any other convenient conjugation procedure can be employed at this stage. Under reducing conditions, the thiol moieties generated by the SPDP or similar derivitization of the polymer react with a reduced radionuclide, such as technetium, rhenium or the like, resulting in rapid chelation of the radionuclide. For such metals, the polymer is derivitized with at least four thiolating agents for each metal atom to be bound thereto. This technique is addressed in Example I(C) hereof. This synthesis offers the following advantages:

- Protein (proteinaceous targeting moiety, ligand or anti-ligand) integrity is maintained, because the radionuclide labeling does not occur directly thereon;
- Reactive thiols can be generated on the polymeric DBM only;
- Chelation is likely to proceed through an S_4 complex with no participation from functionalities present on the targeting moiety (especially proteinaceous targeting moieties), ligand or anti-ligand;
- The tentacle-like configuration of the reactive thiols located on the polymer DBM facilitate proper orientation and maximum stability for the resultant radionuclide-chelate complex; and
- Metabolites of the product conjugate containing radionuclide are preferably excreted via the desirable renal pathway.

An alternative "tentacle chelate" formation process involves thiol functional group generation using a thiolating agent in the presence of reduced radionuclide. A reducing agent capable of concurrent

reduction of radionuclide and thiolating agent in the presence of protein is required to carry out this alternative method. For technetium radionuclide and SPDP thiolating agent, useful reducing agents include stannous ion, dithionite, sulfites, cuprous ion, chromous ion, phosphines and the like. Other examples of chelate-containing conjugates of the present invention and syntheses therefor are set forth below.

Another chelate molecule useful in the practice of the present invention is one known as DOTA for +3 metals. 1,4,7,10-tetraazacyclododecane-N,N',N'',N'''-tetra acetic acid (DOTA)-biotin conjugates have been reported to have desirable in vivo biodistribution and to clear primarily by renal excretion. See, for example, Renn et al., "Large Scale Synthesis of Bifunctional Chelating Agent Q-(p-nitrobenzyl)-1,4,7,10-tetraazacyclododecane-N,N',N'',N'''-tetra acetic acid, and the Determination of its Enantiomeric Purity by Chiral Chromatography," Bioconj. Chem., 3: 563-9, 1992.

Because DOTA strongly binds Y-90 and other radionuclides, it has been proposed for use in radioimmunotherapy. For therapy, it is very important that the stable DOTA bound radionuclide be stably attached to other conjugate components, such as a ligand or an anti-ligand. Release of the radionuclide from the DOTA chelate or cleavage of the biotin and DOTA conjugate components in serum or at non-target sites renders the conjugate unsuitable for use in therapy.

For illustrative purposes, DOTA-biotin conjugates are described in the Examples that follow. Preferably, DOTA and biotin are joined by an N-methyl-glycine linker moiety. In this preferred embodiment, DOTA provides stable complexation of Y-90 or like radionuclide and the N-methyl-glycine spacer provides a stable linkage between the chelated radionuclide and the biotin ligand. DOTA may also be conjugated to other ligands or to anti-ligands in the practice of the present invention.

Ligands suitable for use within the present invention include biotin, haptens, lectins, epitopes and analogs and derivatives thereof. Useful complementary anti-ligands include avidin (for biotin), carbohydrates (for lectins) and antibody, fragments or analogs thereof, including mimetics (for haptens and epitopes). Preferred ligand-anti-ligands binding pairs associate with each other at an affinity of at least about $k_D \geq 10^9$ M.

DBMs of the present invention exhibit renal excretion as well as the ability to direct the excretion of targeting moieties or small molecules bound thereto to renal excretion, while maintaining binding site recognition and binding by the targeting moiety. In addition, preferred DBMs exhibit one or more of the following characteristics: hydrophilicity, polar structure, neutral or anionic charge, non-toxicity to mammalian recipients, a lack of molecular units recognizable by organ receptors, stability in serum, tumor bioavailability and the like. More preferred DBMs are so specifically excreted by the kidneys that when small molecules are bound to the DBM, the conjugated DBM and active agent-containing metabolites thereof are still specifically excreted by the kidneys.

One type of DBM molecule, a large (*i.e.*, of intermediate to high molecular weight ranging from about 5,000 Dal. to about 70,000 Dal.), preferably polymeric (*i.e.*, incorporating from about 30 to about 500 monomeric units) DBM, "masks" the biodistribution properties of the targeting moiety or small molecule conjugate components and prevents liver cell recognition of such moieties necessary for hepatic uptake and biliary excretion thereof. These DBMs of the present invention also preferably exhibit polar structure. Exemplary of such DBMs are dextran; dextran derivatives including carboxymethyl dextran, aminodextran, 3-mercapto-2-hydroxypropyl dextran and the like;

hyaluronic acid, inulin, carboxymethyl cellulose; hydroxy-propylmethacrylamide (HPMA) polymers; succinylated polylysine; polyaspartate; polyglutamate; polyethyleneglycol (PEG); and the like.

5 The disposition characteristics of macromolecules have been investigated. See, for example, Takakura, et al., Pharm. Res., 7:339, 1990. For example, dextrans having a molecular radii of less than 20 angstroms have been reported to cross the glomerulus without measurable
10 restriction by Hurwitz, et al., Eur. J. Cancer, 14:1213, 1978. Antibody-drug binding using dextran has been documented. See, Hurwitz et al. cited above. Also, the water solubility of polymeric macromolecules has been
15 exploited to provide simplified protocols for the preparation of soluble, multivalent macromolecular conjugates. See U.S. Patent No. 5,026,785.

For use in the present invention, dextran DBMs preferably range between about 5 and about 15 kD in size, although larger moieties may also be used. These
20 dextran DBMs are capable of binding multiple and varied small molecules and larger targeting moieties such as antibodies and antibody fragments. More specifically, dextran DBMs useful in pretargeting applications of the present invention may bind one or more ligands or anti-
25 ligands as well as one or more active agents. That is, dextran DBMs of the present invention useful in pretargeting applications thereof may bind, for example, from about 1 to about 3 biotin molecules as well as from about 1 to about 5 chelating groups. For
30 targeted, direct labeled methods of the present invention, DBMs are bound to one or more targeting moieties (e.g., antibodies) and one or more active agents (e.g., chelating agents having a radionuclide complexed therein).

35 When larger (from about 40 to about 70 kD) polymers are employed as DBMs in accordance with the present invention, the rate of clearance of the conjugate from

a recipient is slowed, but the renal pathway for excretion is maintained. In this manner, small molecules or larger targeting moieties bound to the DBM exhibit increased bioavailability as a result of the increased circulation time thereof. Such bound molecules remain directed to renal excretion, however.

Exemplary dextran DBM-containing conjugates of the present invention are dextran-biotin conjugates. Chelate-biotin-dextran conjugates of the present invention are formed, for example, from oxidized dextran by conjugating biocytin hydrazide thereto followed by reaction with a chelate active ester, such as a N-hydroxy succinimidyl ester, a tetrafluorophenyl ester and the like. Alternatively, chelate-biotin-dextran can be formed by reacting dextran hydrazide with the carboxy terminus of a chelate-biocytin conjugate. Preferably, the carboxy terminus has been derivitized to form an active ester, such as an N-hydroxysuccinimidyl ester or the like. Examples of these reaction schemes are discussed in Examples II-IV hereof.

A pre-formed Re-labeling approach useful in the practice of the present invention involves rhenium labeling of biotin-dextran conjugate through acylation with a rhenium complex active ester, such as a tetrafluorophenyl ester as discussed in Example IV(B). A post-formed Re-labeling approach may also be used in accordance with the present invention. As set forth in Example IV(A), the chelating group is conjugated to dextran-biotin conjugate prior to Re chelate labeling in the presence of SnCl_2 and citric acid.

The molar ratio of biotin/dextran in biotin-dextran conjugates of the present invention can be altered by a practitioner in the art to form molecules having different structures by modifying the offering ratio of the ligand with respect to the polymer. For example, an increased offering ratio of biocytin hydrazide to dextran results in conjugates characterized by an

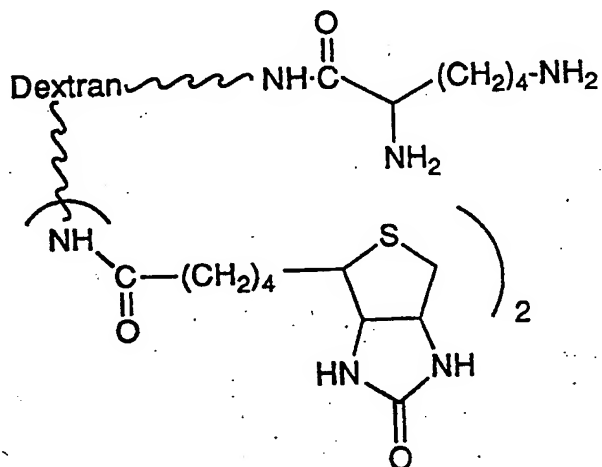
increased number of bound biotin molecules. Tumor targeting can therefore be optimized with respect to the parameter of molar ratio.

Biotin-dextran, lysine fixable (BDLF, available from Sigma Chemical Company, St. Louis, Missouri) is a preferred DBM-ligand of the present invention. BDLF is characterizable as follows:

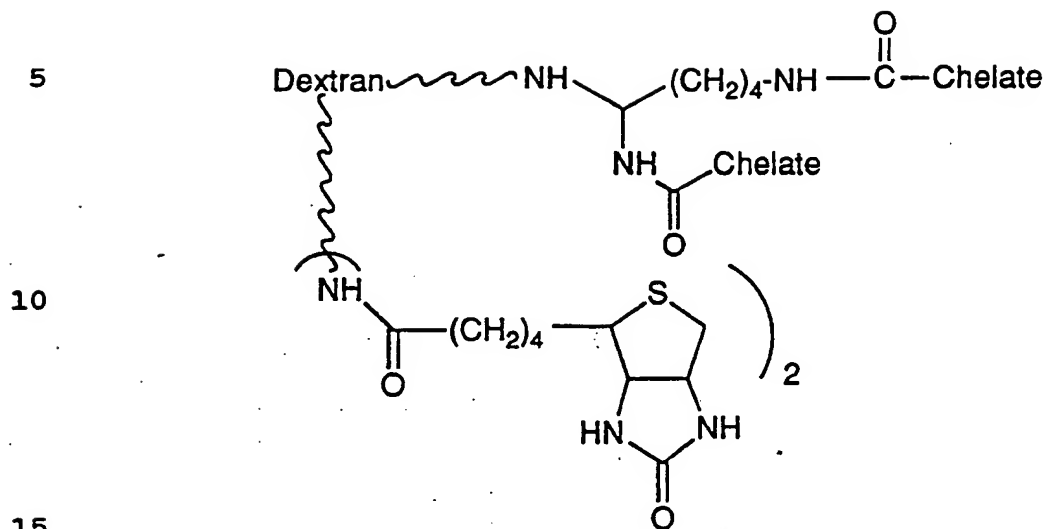
- biotin and lysine carboxyl groups are attached by amide linkages to amino groups of aminodextran;

- biotin attached to dextran through an amine; and

- 2 moles biotin/dextran (10,000 dalton molecular weight) or 20 moles biotin/dextran (70,000 dalton molecular weight). Based upon available information, the structure of BDLF (10,000 dalton) is believed to be the following:



Iodine labeling of BDLF is conducted through acylation with a chelate-active ester, such as N-hydroxysuccinimidyl p-iodobenzoate, as discussed in Example III below. Additional procedures for preparing BDLF-containing conjugates of the present invention are discussed in Example IV below. The BDLF structure depicted above accommodates the binding of a small molecule or active agent, e.g., at the lysine epsilon amino group. Two small molecules or active agents, e.g., radionuclide chelates, may be bound to a derivitized BDLF, an example of such a dual derivitization being shown below:



20 A chelate-dextran-antibody conjugate can be prepared from dextran hydrazide, for example, by reaction of the polymer with a derivitized chelate having an aldehyde functional group and with an oxidized antibody or an antibody-glutaraldehyde conjugate. In addition, antibody-chelate-dextran moieties of the present invention are preparable from carboxymethyl dextran by reacting the polymer with a chelate bearing a first functional group reactive with the modified dextran (e.g., an alcohol to form an ester with the carboxyl moiety of carboxymethyl dextran, an amine to form an amide with such carboxyl moieties or the like) and a second functional group reactive with an antibody or a derivitized antibody (e.g., a maleimide group to react with a free sulfhydryl moiety on the antibody, an aldehyde group to react with an amine moiety located on the antibody, a protected carboxyl group to be deprotected and activated to react with an antibody amino group or the like). Exemplification of these

protocols is shown in Example V hereof.

Chelate-somatostatin derivative (e.g., octreotide)-dextran conjugates are prepared from carboxymethyl dextran, for example, by conjugation of D-Phe-octreotide to the polymer at the amino terminus of the peptide, while the chelate is bound through an amino group to the peptide at the carboxy terminus. The octreotide-polymer bond is formed, for example, by reaction of the peptide terminal amino group with carbodiimide or a pre-activated polymer carboxyl (e.g., an active ester). Also, the chelate-peptide bond is formed, for example, by reaction of the peptide terminal carboxyl group and a chelate amino group, using either carbodiimide or active ester coupling chemistry.

Alternatively, dextran hydrazide is conjugated to the carboxy terminus of D-Phe-octreotide, and a chelate active ester is linked to the amino terminus. Exemplary active esters employable for this purpose are N-hydroxysuccinimide, tetrafluorophenyl ester and the like. Also, the chelate may be bound at the carboxy terminus of the peptide (through a chelate functional group, such as an amino group or the like), and the chelate-peptide is then conjugated to oxidized dextran. Moreover, the chelate-peptide bond may be formed, for example, by reaction of the peptide terminal amino group with an aldehyde functional group of oxidized dextran. In addition, reaction of aminodextran with the carboxy terminus of the peptide can be employed following reaction of a chelate active ester at the peptide amino terminus. These reaction schemes are the subject of Example VI hereof.

As in all three component embodiments of the present invention, dextran/octreotide/chelate conjugates are amenable to three configurations as follows:

(1) Octreotide - - - - DBM - - - - Chelate
or

(2) DBM - - - - Octreotide - - - - Chelate

or

(3) DBM - - - - Chelate - - - - Octreotide

5 In configuration (1), modification of octreotide is minimized, because the somatostatin analog is required to bind to the DBM only. In configuration (2), chelate-octreotide-glucose is retained upon conjugate metabolism, thereby preserving the targeting moiety and
10 active agent as a single entity.

In addition, chelating agents can be bound to dextran DBMs characterized by short term serum stable linkages disposed between monomeric, dimeric, trimeric or other convenient unit thereof. Such conjugates are also
15 preferably large (exhibiting a molecular weight ranging from about 40 to about 70 kD) These conjugates of the present invention are used to enhance the absolute amount of an active agent (e.g., radionuclide, drug, anti-tumor agent or the like) or other small molecule
20 (e.g., ligand, anti-ligand, peptide or the like) or larger targeting moiety (e.g., antibody or antibody fragment) localizing to target sites. One advantage of such polymer DBMs is that the molecules affixed thereto will exhibit an increased circulation time as well as
25 decreased liver uptake. More specifically, the circulation time of the bound molecules is dictated by the maintenance of polymeric structural integrity in vivo. Depolymerization releases bound molecule-DBM monomer, dimer, trimer or like moieties that are
30 themselves rapidly cleared from the recipient's circulation, preferably via the renal pathway.

Depolymerization may be controlled in any convenient manner therefor, including, for example, the following methods:

35 - Use of linkages between dextran units containing chemical groups that are stable in serum for a period of time sufficient to provide an appropriate circulation

time to the small molecules bound thereto (e.g., 1 to 3 hours for active agent in both the pretargeting and targeted, direct labeled protocols); or

- 5 - Use of linkages between dextran units that are enzymatically cleaved upon administration of enzyme after the passage of an appropriate amount of bound molecule circulation time.

10 In the first approach, groups such as esters, acetals, disulfides, thioacetals or the like are employed. For example, an ester linkage, having a serum stability of 1-3 hours such as phenyl or activated phenyl or phthalyl (e.g., chloro-substituted, fluoro-substituted, multi-halogen-substituted, nitro-substituted or the like), may be employed to attach
15 dextran DBM units. Such a linkage is stable in serum for a time sufficient to facilitate localization of the molecules bound to the DBM to the target site.

20 The second approach includes the use of dextran units susceptible to cleavage by an administrable enzyme that is not found in large amounts in human serum. Exemplary enzymes useful in the practice of this aspect of the present invention include dextranase, alpha-amylase, pullulanase (a bacterial alpha-1,6-polysaccharidase) and the like. The enzymes are administered by any
25 convenient route in any convenient dosage form therefor. Such enzymes are optionally conjugated or formed as fusion proteins with long circulating proteins, including albumin, immunoglobulins or portions thereof, and the like. In this manner, the administered enzymes
30 remain in circulation for a time sufficient to effect depolymerization of the polymer DBM.

35 In a modified version of this technique, a dextran DBM is employed, which has both targeting moiety and active agent bound thereto in a manner permitting targeting moiety and active agent to be released as a single molecule as depolymerization progresses. For example, dextran units are linkable to both targeting

moiety and a radionuclide chelate via a stable linkage. Target binding of these conjugates of the present invention maintains both radionuclide and targeting moiety at the target with both components being released upon DBM depolymerization. For pretargeting approaches, ligand or anti-ligand may be bound to the DBM in a manner to be released separately from or as integrally with an active agent-containing molecule.

Conjugates including internalizing targeting moieties bound to short term serum stable polymers bearing multiple active agents are also contemplated by the present invention. In this manner, multiple active agents can be simultaneously targeted, thereby overcoming active agent delivery problems such as low receptor number or low active agent specific activity. For some methods of the present invention employing internalizing targeting moieties, the linkages (e.g., disulfides, esters, ethers, thioethers, hydrazides and the like) between the dextran units are selected to be cleavable in an intracellular environment (e.g., the acidic environment of a lysosome and the like), or upon target localization of subsequently administered cleaving enzymes are also directed by an internalizing targeting moiety.

Intracellularly cleavable linkers are used when the active agent requires release from the internalized conjugate to exert its diagnostic or therapeutic benefit. Exemplary active agents of this type are drugs and the like. For active agents that do not require intracellular cleavage to exert its diagnostic or therapeutic benefit, the linkage between the DBM and the active agent may be stable to intracellular conditions. Exemplary active agents of this type are radionuclides and the like.

Radiolabeled ligand (e.g., biotin) is an exemplary small molecule capable of directed biodistribution in accordance with the present invention. Other molecules

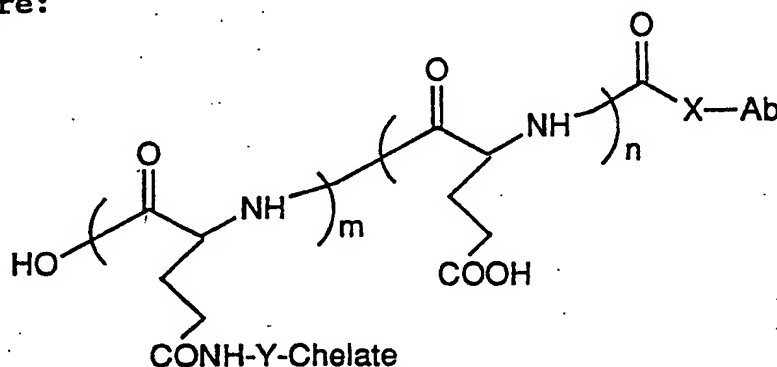
which may show directed biodistribution (*i.e.*, small molecules or larger targeting moieties) consist of or contain drugs, ligands such as biotin, anti-ligands such as avidin and streptavidin, targeting moieties, such as antibodies and peptides specific for target cell receptors (*e.g.*, somatostatin, derivatives thereof such as octreotide and MK-678 (Merck), melanocyte stimulating hormone and the like), epidermal growth factor, chemotactic peptides, cytokines, such as interleukins including interleukins such as IL-2, IL-4, IL-6, IL-12, tumor necrosis factor (TNF), interferons such as gamma-interferon, growth factors, such as GM-CSF, M-CSF and the like, photosensitizing agents, prodrugs, c-kit ligand or stem cell growth factor and the like.

Small molecules useful in the practice of the present invention exhibit one or more of the following characteristics: a molecular weight below about 2 kD, lipophilicity, hepatobiliary excretion, ability to bind (directly or indirectly) to DBMs of the present invention and the like. Useful targeting moieties selectively localize to the target cells of interest and are capable of binding (directly or indirectly) to DBMs to form conjugates of the present invention. In addition, useful small molecules and larger targeting moieties preferably exhibit or are derivitizable to exhibit one or more of the following functional groups: aldehyde, maleimide, amine, thiol, alcohol, carboxyl and the like.

A problem encountered in effectively delivering active agent to target cells is created by the recipient's catabolism or metabolism. Catabolic byproducts of administered conjugates (*e.g.*, stable chelate-lysine adducts of antibody-chelate-radionuclide conjugates, such as Tc-99m or Re-186 mercaptoacetylglycylglycyl-gamma-aminobutyryl lysine, Tc-99m or Re-186 4,5-dimercaptoacetamido-pentanoyl lysine and the like) exhibit undesirable biodistribution

profiles in that such adducts are subject to hepatobiliary excretion. DBMs of the present invention also preferably direct the excretion of such metabolites to the renal pathway.

Polyglutamate is another example of a DBM useful in the practice of the present invention. Polyglutamate DBMs are structurally capable of delivering multiple active agents per targeting moiety and directing excretion of such active agents to the renal pathway. Exemplary polyglutamate conjugates include chelate-polyglutamate-antibody (Ab) molecules of the following structure:



where X is a stable or conditionally unstable (e.g., acid labile, enzyme cleavable, base labile or the like) linkage. For example, X can be -NH- (amide, stable), -NH₂CH₂CONH- (alkoximine, acid labile) or -O(CH₂)_nCONH-; n is 1-2 (ester, enzyme cleavable, base labile). Y is a spacer incorporated for synthetic convenience, which can constitute for example, an alkyl or amino acid amide moiety. This moiety contains m carboxylates attached to chelate amino groups (preferably through amino terminus attachment to the antibody-linker to reduce cross-linking) and n free carboxylates. The values of n and m are manipulable by one of ordinary skill in the art to deliver a diagnostic or therapeutic dose of radioactivity to target cells, while maintaining the desired biodistribution profile.

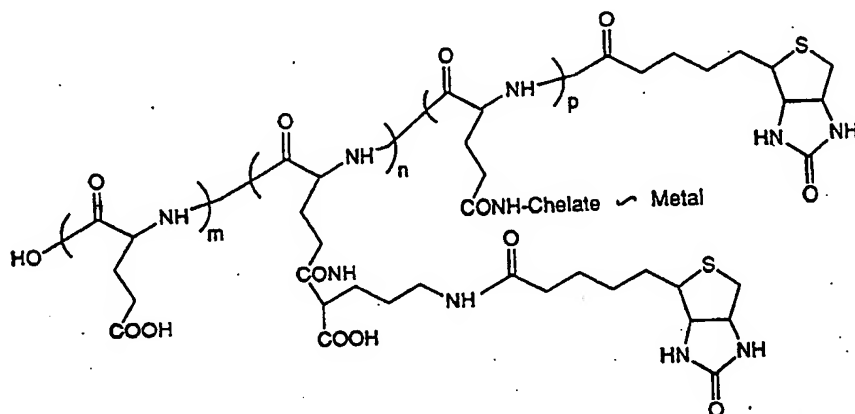
As can be ascertained from an examination of the chelate-DBM-antibody conjugate structure, the number of

radionuclides bound to the conjugate ranges between about 0.01x and about 0.1x (where x is the number of units in the polyglutamate polymer, generally ranging between from about 7 and about 50). By delivering more radiometal atoms per antibody targeting molecule, the specific activity of the administered conjugate is enhanced. This feature is shared by the other polymeric DBMs of the present invention.

In addition, the antibody-DBM linkage may be rendered stable (e.g., simple amide linkage), acid labile (e.g., alkoximine linkage) or enzyme cleavable, base labile (e.g., ester linkage). The advantage of using conditionally cleavable linkers is an improvement in target/non-target localization of the active agent. More specifically, use of linkers that are cleaved by enzymes, which are present in non-target tissues but reduced in amount or absent in target tissue, can increase target cell retention of internalized active agent (e.g., targeted by an internalizing targeting moiety) relative to non-target cell retention, because the polymeric DBMs are unlikely to be excreted once internalized into the target cells. For example, ester linkages are susceptible to esterases in the liver and kidney as well as to hydrolysis. Also, alkoximine linkers are susceptible to hydrolysis in the liver and in lysosomes within tumor cells. Such linkers are useful, for example, in delivering therapeutic radionuclides to target cells, because such active agents do not require internalization for efficacy, provided that the linker is stable at the target cell surface or protected from target cell degradation.

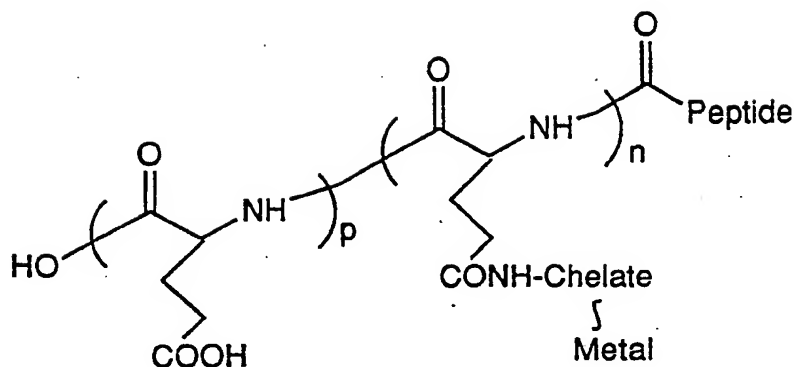
Both non-target associated conjugate and active agent-containing metabolites thereof are directed to renal excretion by the polyglutamate DBM. Synthesis of conjugates containing the exemplary stable and conditionally unstable polyglutamyl-antibody linkages are set forth in Example IX hereof.

Other examples of polyglutamate DBM conjugates include DBM-ligand molecules, where, for example, biotin-N-hydroxysuccinimide (NHS) active ester is conjugated to polyglutamyl polymer to form the compound depicted below.



An exemplary synthesis of this molecule is set forth in Example X. Other active esters, such as tetrafluorophenyl or the like may also be employed. This moiety contains p monomers attached to chelate functional groups, n + 1 monomers attached to ligand functional groups and m underivatized monomers. The values of n, m and p are manipulable by one of ordinary skill in the art to achieve a therapeutic or diagnostic objective, while maintaining the desired biodistribution profile.

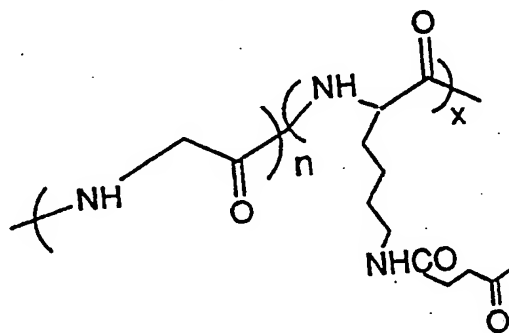
Peptide-polyglutamyl conjugates are prepared, for example, by reacting the activated carboxy terminus of the peptide (e.g., an active ester form of the carboxylic acid group, such as a tetrafluorophenyl ester, a N-hydroxysuccinimide ester or the like) to polyglutamate to form a compound of the structure shown below.



This moiety contains n monomers attached to chelate functional groups (e.g., amines) and p underivatized monomers. The values of n and p are manipulable by one of ordinary skill in the art to achieve a therapeutic or diagnostic objective, while maintaining the desired biodistribution profile.

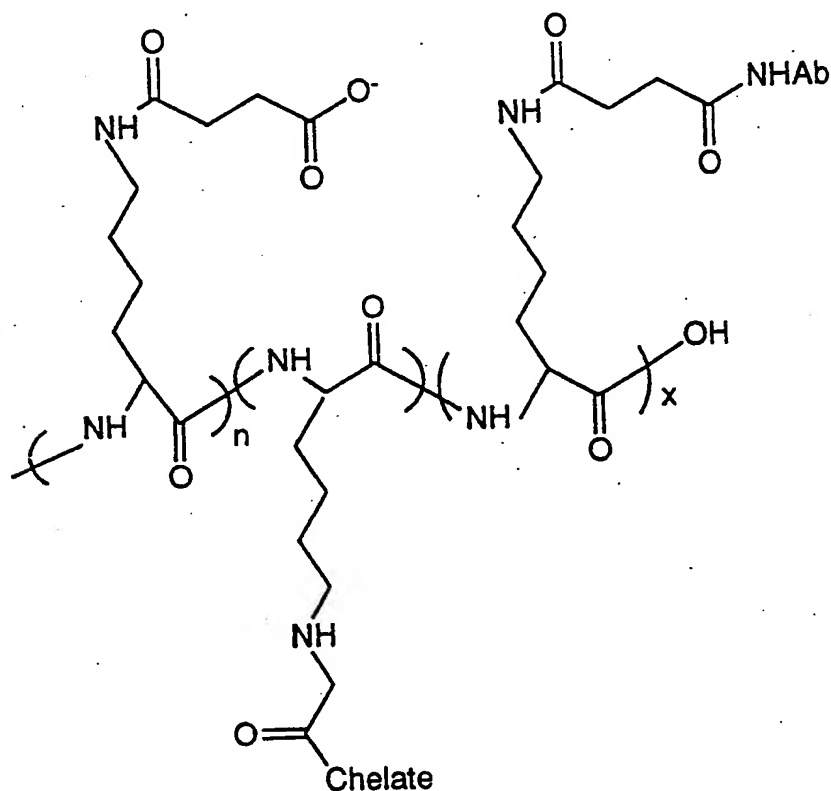
Chelate conjugation to the exposed acid moieties of the polyglutamate can be conducted for the DBM-biotin and DBM-peptide molecules in substantially the same manner as for the antibody-DBM discussed in Example IX. In addition, multiple chemotherapeutic drugs or other active agents containing, for example, amino or alcohol groups, can be conjugated to the exposed carboxy groups using conventional chemistry therefor.

Another exemplary DBM of the present invention is succinylated polylysine having the following structure:



where n ranges from about 25 to about 500, and x ranges

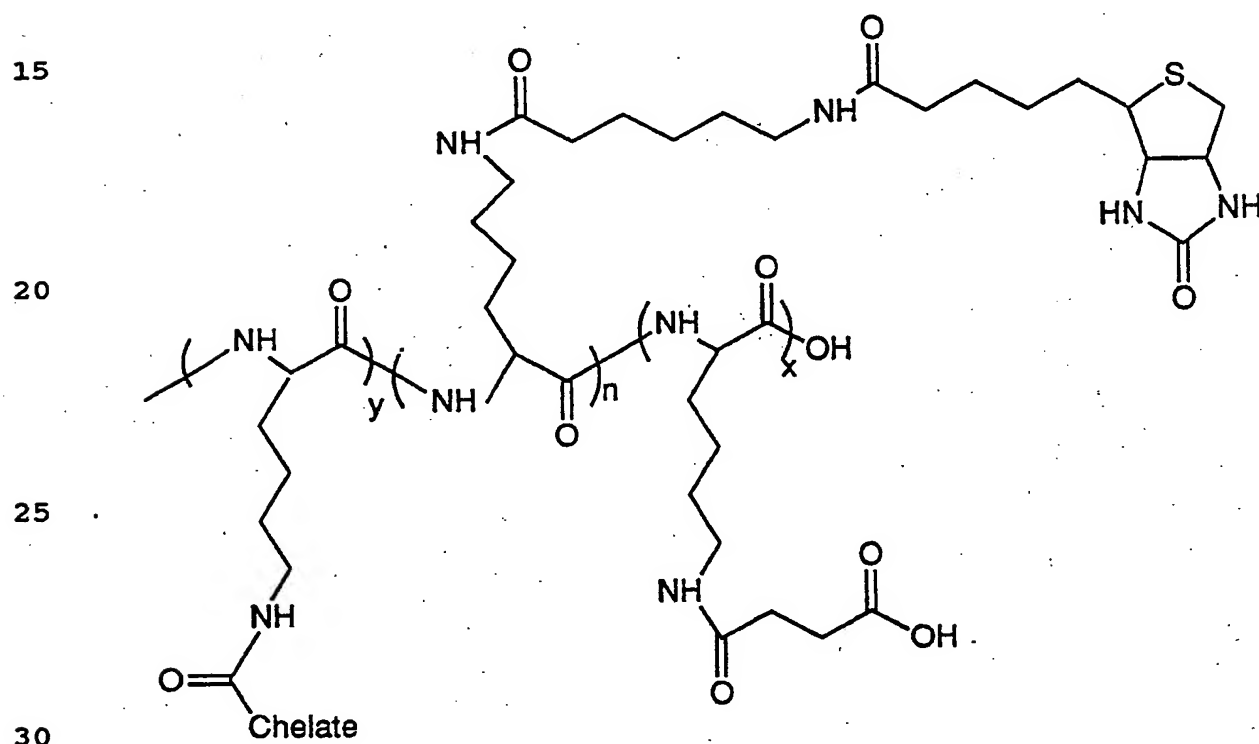
from 0 to about 25. Polylysine DBMs are capable of delivering multiple active agents to target cells. Moreover, the succinylation directs the biodistribution of the DBM and, therefore, the DBM-containing conjugates or DBM-containing conjugate metabolites to renal excretion. Exemplary succinylated polylysine conjugates of the present invention include chelate-succinylated polylysine-antibody molecules of the following structure:



where n ranges from about 16 to about 475; x ranges from about 0 to about 1; and y ranges from about 1 to about 10. These conjugates are prepared, for example, by reacting epsilon amino lysine moieties of the polymer with chelate active esters, such as N-hydroxysuccinimide

esters, tetrafluorophenyl esters or the like followed by reaction with succinic anhydride or the like. A carboxylate group (e.g., the carboxy terminus) of the succinylated polylysine is then reacted with an antibody amino group in EDCI. Alternatively, succinylated-polylysine conjugates of the present invention are preparable in accordance with Example XI hereof.

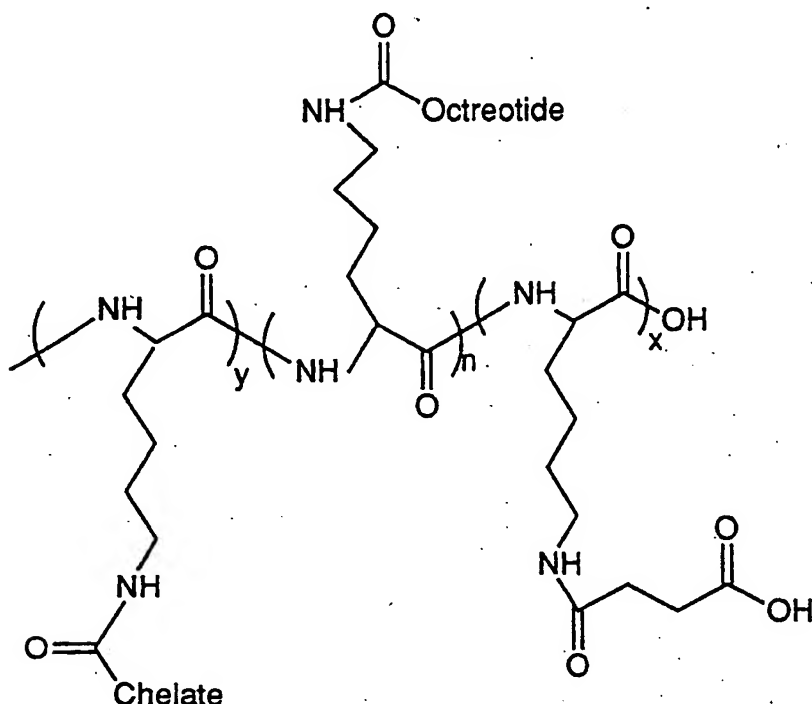
Other examples of succinylated polylysine DBMs of the present invention include DBM-ligand molecules, where, for example, biotin-N-hydroxysuccinimide ester is conjugated to the epsilon amino groups of polylysine to form a compound of the following structure:



where n ranges from about 1 to about 5; y ranges from about 5 to about 20; and x ranges from about 16 to about 475. Other active esters employable for this purpose

are tetrafluorophenyl esters and the like.

Peptide-succinylated polylysine conjugates are prepared, for example, by conjugating the carboxy terminus of the peptide with the polymer epsilon amino groups, resulting in a compound of the structure shown below.



where n ranges from about 1 to about 5; y ranges from about 5 to about 20; and x ranges from about 16 to about 475. To prepare chelate-succinylated polylysine-biotin and chelate-succinylated polylysine-protein conjugates, polylysine is acylated with a serial offering of biotin active ester derivative (or protein) followed by chelate active ester. Succinic anhydride activation is then conducted in accordance with known techniques therefor. Exemplary active esters for this purpose are N-hydroxysuccinimide or the like. The absolute and relative offering ratios of the biotin/peptide and chelate will impact the structure or structure distribution of the final product. Derivatization of the DBM adversely impacts the biodistribution directing properties thereof. Consequently, more heavily derivitized DBM is less likely to properly direct the

biodistribution of bound molecules in accordance with the present invention. Preferably, the molecular weight of the DBM is >> the molecular weight of the molecules bound thereto.

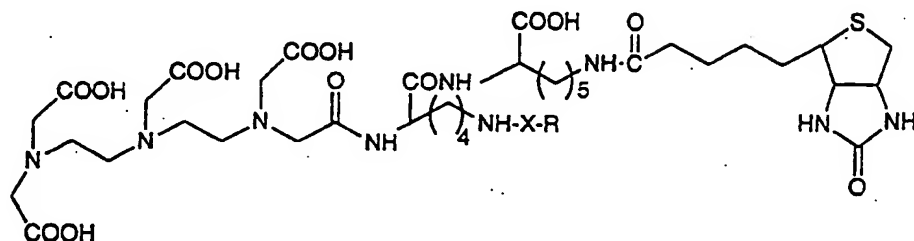
5 Another aspect of the present invention involves lower molecular weight DBMs (i.e., ranging from about 500 to about 2000 Dal.) of high polarity. Preferably, DBMs of this aspect of the present invention are highly charged, i.e., greater than about 2 ionic charges/1000
10 dalton molecular weight. An exemplary DBM of this aspect of the present invention is diethylene triamine pentaacetic acid (DTPA described by Hnatowich, IJARI, 33: 327-333, 1982); DTPA derivatives (e.g., carbon-backbone substituted DTPA derivatives, such as SCN-Bz-DTPA);
15 ethylenediamine tetraacetic acid (EDTA); aminotriacetic acid (NTA); ethylene glycoldiamino tetraacetic acid (EGTA); other polyimino acetic acids; 1,4,7,10-tetraazacyclododecane-N,N',N'',N'''-tetraacetic acid (DOTA) derivative or other polycarboxylic
20 acid molecule; and the like. DTPA and DOTA and derivatives thereof are preferred DBMs in the practice of this aspect of the present invention.

Imaging studies of tumor and infection by pretargeting with avidin generally involve biotin
25 labeled with In-111, with In-bis-biotin-DTPA being the most commonly employed moiety. In-bis-biotin-DTPA is water soluble and contains regions of positive and negative charge imparted by tertiary amines and carboxylates, respectively. DTPA has been conjugated to
30 octreotide for the purpose of labeling octreotide with In-111 (J. Nucl. Med., 33: 652- 658, 1992).

Very little study has been reported with respect to technetium labeling of biotin. Virzi et al., "The preparation and evaluation of 12 biotin derivatives
35 labeled with Tc-99m," J. Nucl. Med., 1192, 33:5, 920 (abstract) discuss aminothiols and polyaminocarboxylate chelating agents, resulting in conjugates of poor in

vivo stability.

The present invention provides stable biotin-DTPA conjugates, such as the following:



where R is a chelate and X is a spacer included in the structure for synthetic convenience. More specifically, X is preferably a carbonyl spacer, such as $-\text{CO}-$, $-\text{CO}-$, $(\text{CH}_2)-$, $-\text{COCH}_2\text{NHCO}-$ or the like. For rhenium radionuclides, R is preferably an N_2S_2 or an N_3S amidothiolate chelate. Also preferred in the practice of the present invention are iodobenzamide chelate-DBMs, where X is preferably a carbonyl spacer, $-\text{CO}-$, and R is preferably p-iodophenyl (PIP).

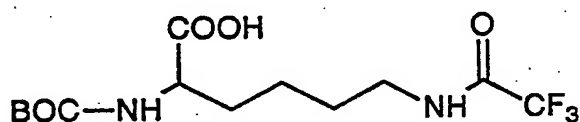
The above-identified conjugate employs a lysine amino acid group to link the ligand to the DBM and to link the chelate to the DBM. Other groups useful for this purpose include amino acids, such as aspartic acid, glutamic acid, serine, threonine, cysteine and the like; synthetic compounds containing appropriate functional groups, such as carboxylic acid, amine, alcohol or thiol; or the like. The structure shown above depicts the ligand attached to the amino terminus of lysine, with active agent attachment to the epsilon amino group thereof. Other options include the following:

- ligand to carboxy terminus/active agent to epsilon amino group;
- ligand to epsilon amino group/active agent to carboxy terminus;
- ligand to carboxy terminus/active agent to amino terminus; and
- ligand to amino terminus/active agent to carboxy

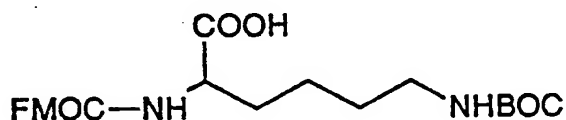
terminus.

The stable DTPA conjugates of the present invention direct the biodistribution of molecules associated therewith (particularly small molecules) to renal excretion. In this manner, small molecule (especially active agent) hepatobiliary excretion is decreased.

These embodiments of the DBMs of the present invention are prepared, for example, in accordance with the reaction mechanism set forth and described in Example VII herein. Briefly, a protected lysine-biotin conjugate (or other similar biotin derivative, such as glutamic acid biocytinamide or the like) is prepared; DTPA anhydride is opened with the lysine-biotin (or other derivative); the lysine (or other reactive moiety) is deprotected; the amine (or other functional group) is acylated with an active ester chelate, such as tetrafluorophenyl esters, N-hydroxysuccinimide esters and the like. A useful protecting group for this purpose is



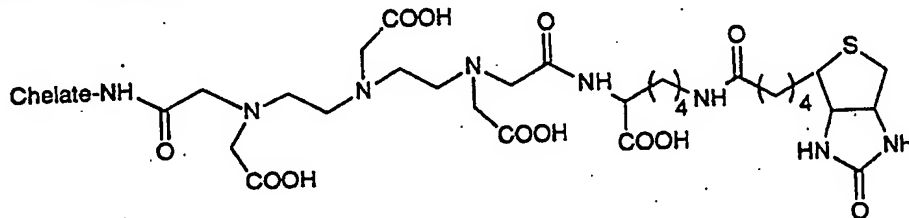
An alternative reaction scheme employing a different protection/deprotection strategy may be used in the practice of these embodiments of the present invention. In this alternative scheme, the following protecting group is employed.



As a result, the alternative reaction scheme proceeds as described above and in Example VII herein, with the most notable exception being that the deprotection of the epsilon lysine amino group involves TFA cleavage of BOC.

It is useful to note that DTPA is itself a chelate and has been used as such for radionuclides such as In-111 and Y-90. Consequently, precomplexation of the DTPA chelate may be necessary to form conjugates of the present invention involving a radionuclide chelate bound to a DTPA DBM. Preferred chelates used in the practice of the present invention (e.g., N_xS_y compounds and the like) may not require such precomplexation. The radionuclide-chelate complexes formed by the preferred chelates are generally thermodynamically more stable than the DTPA complexes thereof. Consequently, at the conditions employed for radiolabeling with, (e.g., Tc-99m, Re-186, Re-188 and the like), the radionuclide typically transchelates from the DTPA core to the preferred chelate core.

An example of another embodiment of the conjugates of the present invention includes has the formula set forth below:



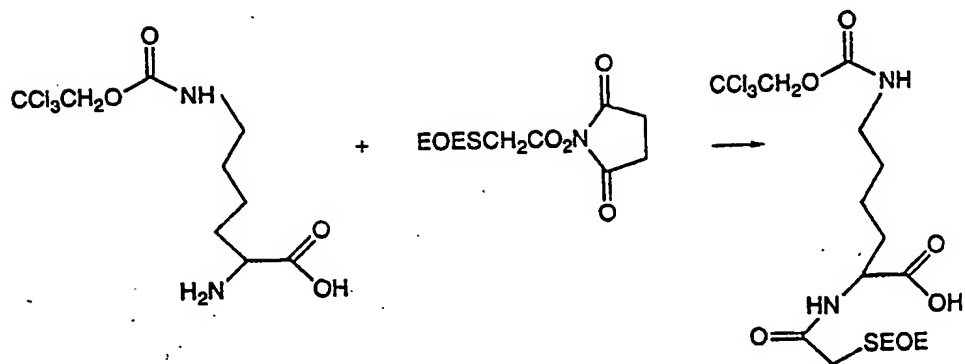
In this embodiment of the present invention one small molecule is attached to a carboxylate moiety of DTPA, while another small molecule is bound to a second carboxylate moiety thereof. These embodiments of the DBMs of the present invention are prepared, for example, in accordance with the reaction mechanism set forth and described in Example VII(C) herein.

These stable DTPA conjugate embodiments of the present invention direct the biodistribution of molecules associated therewith to renal excretion. In this manner, small molecule (especially active agent) hepatobiliary excretion is decreased.

Peptides may also be conjugated to DTPA derivatives in accordance with the present invention. The strategy for preparing, for example, a radiolabeled chelate-peptide-DTPA conjugate involves the preparation of a chelate-peptide conjugate which contains a free amino group for reacting with DTPA anhydride to form an amide linkage between a DTPA carboxylate group and a chelate or peptide amine group. For a peptide amenable to reaction with a chelate amine group, the following procedure may be employed to form a chelate-peptide conjugate. Briefly, a N_3S trifluorophenyl ester is reacted with ethylene diamine in CH_3CN to form an amide linkage and generate a chelate with a free amine. This chelate is reacted with BOP and DIEA (diethylamine) to form a chelate-peptide conjugate through reaction with an unprotected peptide carboxylate group (e.g., the acid moiety of an aspartic acid or of a glutamic acid amino acid). An unprotected amine group of the chelate-peptide conjugate (e.g., a lysine epsilon amino group) is reacted with a DTPA anhydride to form the product chelate-peptide-DBM conjugate. Preferably, the amine group employed in the ultimate conjugation is protected during the previous steps to avoid the generation of undesirable byproducts. TFA is exemplary of protecting groups that are employable for this purpose.

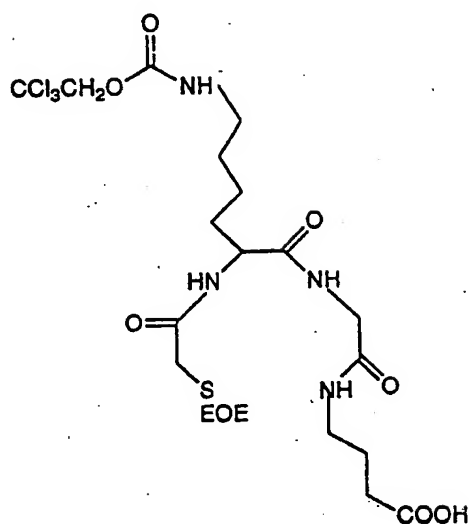
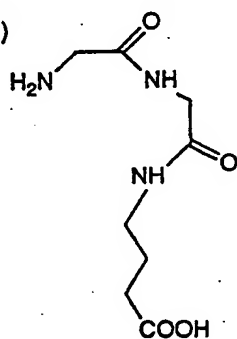
Alternatively, replacement of a glycine by a lysine in a N_3S chelate, i.e., mercaptoacetylglycylglycyl-gamma-amino butyric acid (MAGG-GABA) chelate modified as described above with ethylene diamine in CH_3CN , provides an amino group for reaction with an unprotected peptide carboxylate group (e.g., the acid moiety of an aspartic acid or a glutamic acid amino acid) to form a chelate-peptide conjugate using analogous chemistry to that employed in the reaction scheme discussed above. The derivitized chelate is prepared in accordance with the following reaction scheme.

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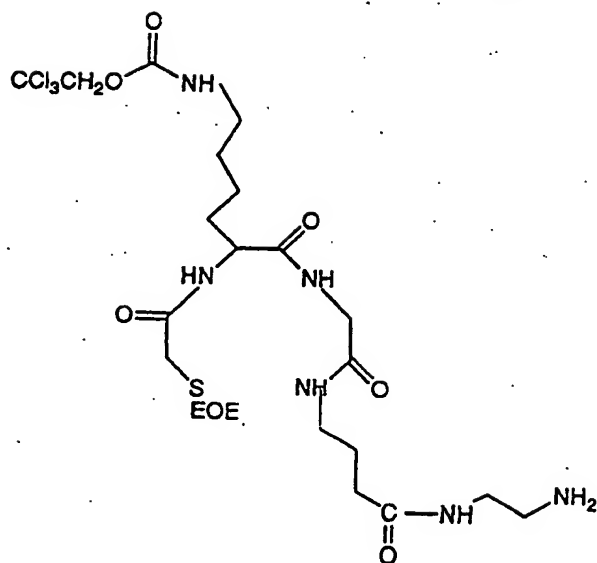
1) NHS
DCC

2)



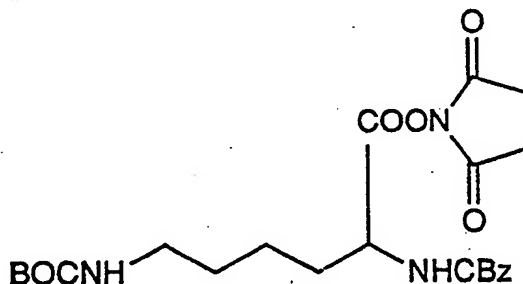
1) TFP, DCC

2)



The derivitized chelate also has an amino group available for subsequent reaction with DTPA anhydride to form the product DBM-chelate-peptide using analogous chemistry to the reaction scheme discussed above. Preferably, the amino group involved in the DBM conjugation is protected during the chelate-peptide conjugation. Any amino groups on the peptide are preferably protected until formation of a DBM-chelate-peptide molecule.

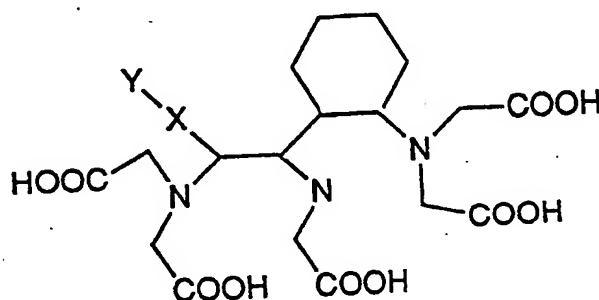
Another alternative involves derivitization of the peptide to provide an amino group for reaction with DTPA anhydride. A peptide lysine, for example, can be reacted with, for example, the following compound



to provide one protected amino group which can be deprotected with TFA and a second protected amino group which can be deprotected with hydrogen. The derivitized peptide is reacted with TFA, and the unprotected amine is then exposed to DTPA anhydride to form a peptide-DBM conjugate with a protected amine group. The conjugate is exposed to hydrogen to form a versatile intermediate that is reactive with a variety of chelate active esters to form a DBM-chelate-peptide conjugate. During this synthesis, any other amino groups on the peptide are preferably protected. TFA is an exemplary protecting group useful for this purpose.

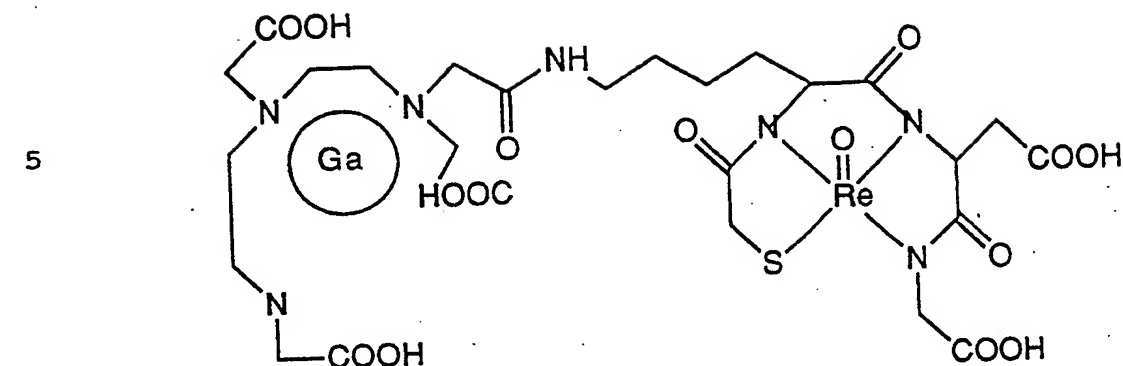
Antibodies or antibody fragments may also be conjugated to DTPA DBMS of the present invention. An exemplary procedure for this synthesis is discussed in Example VIII hereof.

Another embodiment of the present invention features cyclic DTPA DBMs of the following formula:



wherein X is a linking group and Y is a small molecule. Small molecules that are preferably used in this embodiment of the present invention are radiolabeled biotin, drugs, peptides, chelates and the like. Preferred linking groups useful in the cyclic DTPA DBM embodiments of the present invention are amides, esters, ethers, thioethers and the like. Conjugates of this aspect of the present invention are preparable using the techniques described herein for small molecule conjugation to DTPA. Cyclic DTPA synthesis is known in the art.

In addition, the present invention contemplates complexation of the DTPA core with a nonradioactive metal, such as iron, indium, gallium, aluminum or the like. An exemplary conjugate of the present invention incorporating such a DTPA core-complexed DBM is shown below.



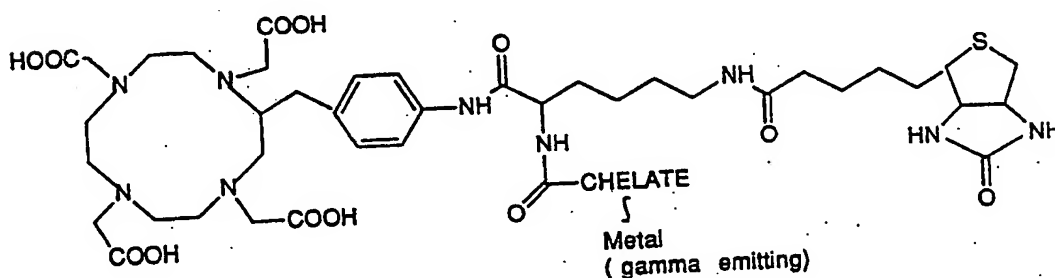
Other small molecules (e.g., peptides, ligands, anti-ligands, chelates or the like) may also be incorporated into these conjugates of the present invention.

Preparation of DTPA core-complexed DBM conjugates of the present invention which also include a radionuclide-chelate complex may be conducted in several ways. If the radionuclide preferably binds to the chelate rather than the DTPA core, the conjugate may be radiolabeled with the radionuclide with subsequent complexation with the nonradioactive metal. The nonradioactive metal complexation can be conducted first, provided that the nonradioactive metal preferentially binds to DTPA and the resultant metal-DTPA complex is stable with respect to radionuclide chelation conditions. Some rhenium labeling protocols, for example, require a powerful reducing agent (stannous chloride) and heat.

Alternatively, a procedure that does not require selective binding of a metal in the presence of two uncomplexed chelators can be employed. An exemplary procedure of this type involves chelating DTPA anhydride with a trivalent metal prior to conjugation thereof with the chelate. Subsequently, the metal-DTPA-chelate is radiolabeled with rhenium.

Another conjugate that is the subject of the present

invention is an improved 1,4,7,10-tetraazacyclododecane-
 N,N',N'',N'''-tetra acetic acid (DOTA) derivative or
 other polycarboxylic acid molecules. DOTA has been
 employed to deliver Y-90 to target cells and described
 by Renn and Meares for Y-90 in Bioconjugate Chem., 3:
 563-569, 1992. A problem with such delivery is an
 inability to follow the in vivo localization of the
 conjugates over time with gamma camera imaging, because
 Y-90 is not a gamma-emitter. In clinical trials, for
 example, it is very important to identify normal organ
 distribution of a conjugate incorporating a radionuclide
 to determine the maximum safe dose that can be
 administered to recipients. Procedures for obtaining
 biodistributions are also complicated by the lack of
 gamma emission, necessitating indirect measurement of
 bremsstrahlung from the beta-decay of Y-90. These
 problems are overcome in using a compound of the
 following formula:



These conjugates of the present invention are
 characterized by the attachment of a gamma-emitting
 radionuclide (e.g., In-111, Tc-99m, Re-186, Re-188 or
 the like) to the biotin side chain and the retention of
 Y-90 in the DOTA core. The chelate component of the
 conjugates of this embodiment of the present invention
 is selected based upon the radionuclide to be complexed
 therein. Exemplary chelates for use in this embodiment
 are N_xS_y chelates for rhenium and technetium, DTPA or

DOTA for In-111 and the like. The conjugates of this aspect of the present invention can be prepared in accordance with one of the two schemes shown below.

Scheme 1:

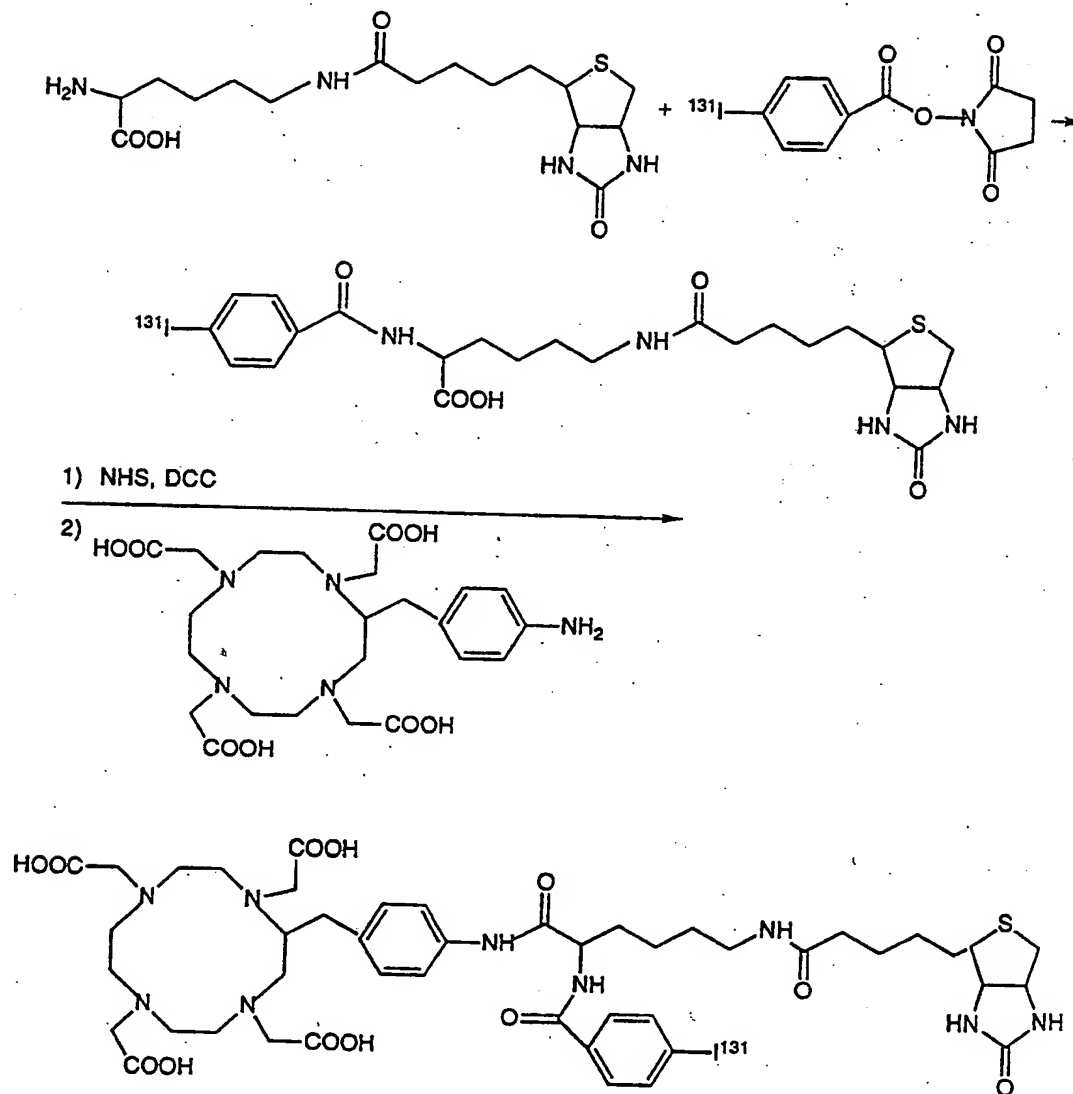
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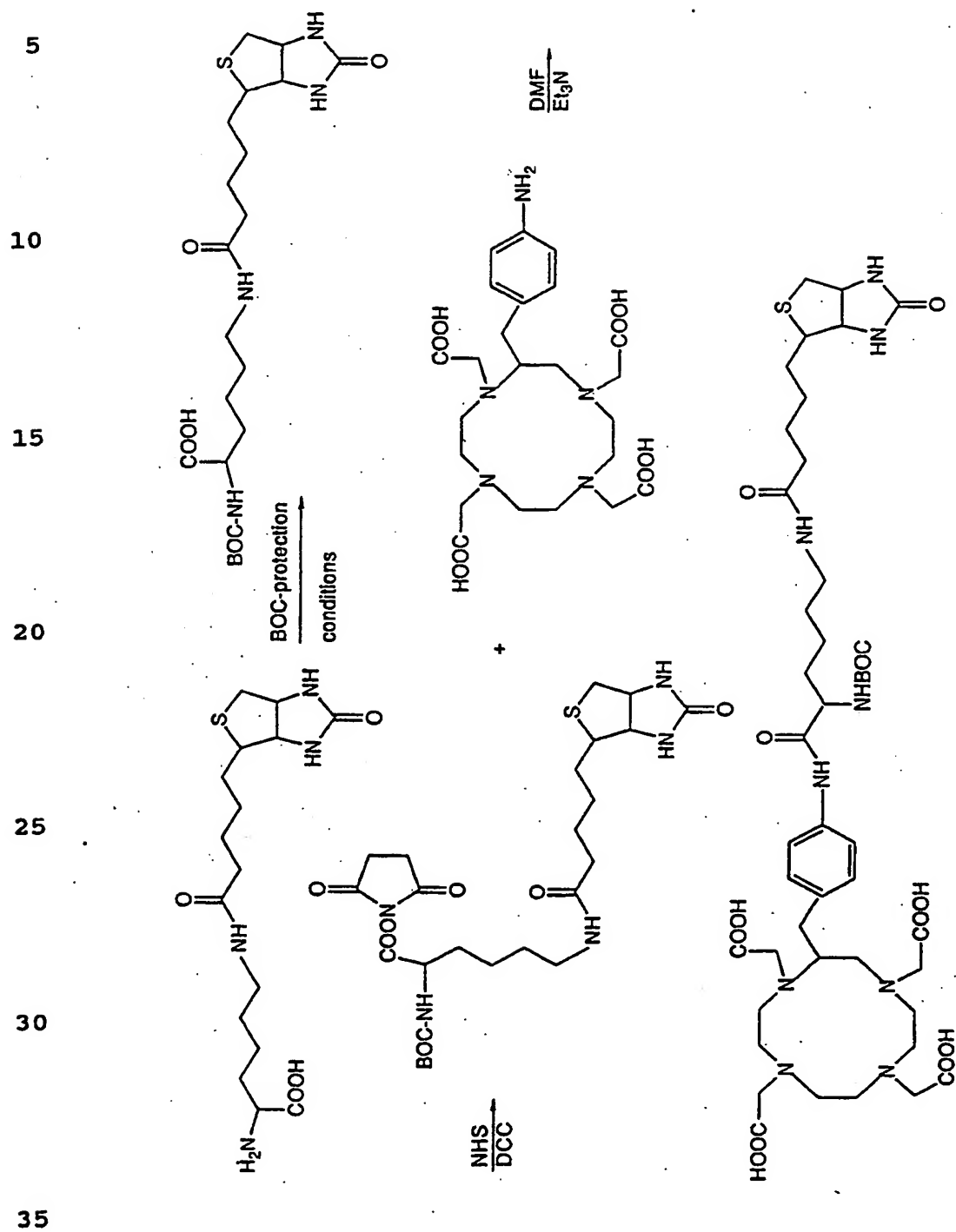
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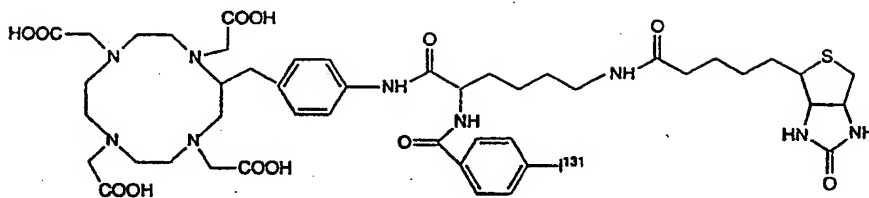
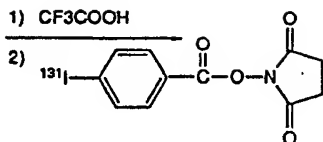
20

25



Scheme 2:





These conjugates of the present invention can be followed by gamma camera imaging and direct biodistribution data thereof can be obtained. In addition, the renal excretion properties of DOTA direct the distribution of the radionuclides bound to the biotin conjugate component, thereby serving as a DBM in the manner of the DTPA derivatives described above.

Another example of a DBM of the present invention is polyethylene glycol (PEG). Enzymes covalently attached to methoxy-PEG (5000 molecular weight), PEGylated enzymes, have been shown generally to be non-immunogenic. See, for example, Abuchowski et al., "Effect of Covalent Attachment of Polyethylene Glycol on Immunogenicity and Circulating Life of Bovine Liver Catalase," J. Biol. Chem., 252: 3581 (1977). Such PEGylated enzymes retained enzyme activity and exhibited an extended serum half-life.

Other proteinaceous moieties (e.g., proteinaceous active agents such as cytokines, proteinaceous ligands or anti-ligands, such as the anti-ligand streptavidin, antibodies, antibody fragments and other proteinaceous targeting moieties) may be PEGylated by substantially the same procedures by which enzymes are PEGylated. For the purpose of illustration, conjugates containing PEG,

biotin ligand, and a cytokine are discussed.

A problem with employing cytokine active agents, for example, is non-target site toxicity caused by liver and other non-target site uptake of such active agents when they are directly conjugated to targeting moieties such as monoclonal antibodies. For example, Reisfeld and Gilles et al. have presented data at the 1993 AACR Immunotherapy Meeting in Asheville, NC and the 1993 Antibody Engineering Conference in San Diego, CA indicating that anti-GD2 antibody in the form of ch14.18-IL-2 conjugates has shown low levels of tumor localization and high liver uptake.

PEGylated cytokines have been constructed and tested. Such PEGylated cytokines exhibit increased circulation times in the blood, indicating that liver uptake of the cytokines is blocked by the PEG modification. Exemplary cytokines include interleukins such as IL-2, IL-4, IL-6, IL-12, tumor necrosis factor (TNF), interferons such as gamma-interferon, growth factors, such as GM-CSF, M-CSF and the like, with IL-2 and TNF preferred. Preferably in pretargeting protocols of the present invention, PEGylated cytokine-ligand or PEGylated cytokine-anti-ligand conjugates are employed to direct localization of the active agent to pre-targeted anti-ligand or ligand, respectively, and to direct biodistribution of non-targeted conjugate to renal excretion.

Exemplary conjugates useful in two-step pretargeting protocols of the present invention which employ the biotin-avidin ligand-anti-ligand pair may be schematically represented as follows:

PEG----cytokine----biotin;

or

Biotin----PEG----cytokine;

or

Biotin----PEG----cleavable linker----cytokine;

or

Biotin----PEG----cleavable linker----cytokine----biotin,

wherein the number of biotins associated with each cytokine preferably ranges from 1 to about 3, and the number of PEGs associated with each cytokine preferably ranges from 1 to about 5, depending in part upon the size of the PEGs.

PEG----cytokine----biotin constructs are characterized by structural simplicity. One useful PEGylation procedure to generate such constructs employs previously derivatized cyanuric chloride-PEG (available from Sigma Chemical Co., St. Louis, Missouri) which reacts with protein amine groups and proceeds, for example, as follows:

- Protein or protein-bearing conjugate is buffer exchanged into 0.1M borate, pH 9.2;

- Cyanuric chloride-PEG is added at an appropriate stoichiometric level (excess of PEG in comparison to conjugate) and the mixture is incubated for one hour at room temperature; and

- the PEGylated product is purified by anion exchange chromatography, wherein the product passed through while underivatized protein or protein-bearing conjugate bound to the matrix.

Also, non-proteinaceous moieties having appropriate functional groups may also be PEGylated.

In general, the PEGylation procedure is as follows: a hydroxyl group of poly(ethyleneglycol) monomethylether is activated to form an active ester for reaction with an amine or other appropriate functional group on a proteinaceous or non-proteinaceous moiety. For example, poly(ethyleneglycol) monomethylether, commercially available from Sigma Chemical Company, St. Louis, Missouri, is reacted with succinic anhydride to form poly(ethyleneglycol) monomethylether mono succinic acid. The acid is reacted with DCC and N-hydroxy succinimide to form the product succinimide active ester.

Alternatively, other activating groups may be employed to produce a poly(ethyleneglycol) molecule

amenable to conjugation with proteins, peptides or other moieties having suitable functional groups. Exemplary of such activating groups are cyanuric chloride (described in Anal. Biochem., 165: 114 (1987) and J. Biol. Chem., 252: 3578 (1977)); carbonate active esters such as para-nitrophenylcarbonate, succinimidyl carbonate or the like (see, generally, Appl. Biochem. Biotech., 11: 141 (1985)); other activated carbonates such as imidazolyl carbonate (Anal. Biochem., 131: 25 (1983)).

The ratio of PEG to protein impacts the results, with increased PEG rendering the protein both non-antigenic (*i.e.*, unable to react with antibodies formed against the native protein) as well as non-immunogenic. For the cytokine IL-2, for example, the ratios of PEG:cytokine will range between from about 3:1 to about 5:1 for PEGs of approximately 3 kD and will be approximately 1:1 for 2 kD PEGs. The offering ratio necessary to obtain product conjugates with the ratios set forth above depends primarily upon the efficiency of the conjugation groups selected.

The ability of a PEGylated active agent to localize to target sites or to exert its activity at such target sites may be tested in accordance with known procedures therefor. A method for testing localization of a PEGylated molecule, such as a cytokine, is as follows: Tumor cells expressing antigen recognized by the targeting moiety that is conjugated with a ligand or an anti-ligand (*e.g.*, the anti-ligand streptavidin) are incubated in the presence of targeting moiety-streptavidin conjugate for 1 hour. Anti-ligand- or ligand-conjugated, PEGylated cytokine (*e.g.*, biotinylated PEGylated cytokine) is added for an additional hour.

The tumor cells are then mixed with cytotoxic effector cells that respond to the cytokine active agent. A ^{51}Cr release, ^3H thymidine release or dye

reduction assay is next employed to determine the degree of induced tumor cell killing mediated by the cytokine-activated effector cells. Control target cells (e.g., no cytokine bound to tumor cells) are also used to evaluate the cytotoxicity attributable to the presence of the cytokine. An additional control may be conducted by adding free (non-ligand or anti-ligand conjugated) cytokine at differing concentrations to the effector cell/target cell mixture. Examples of such a system involve targeted IL-2 bound to tumor cells incubated with natural killer cells isolated from peripheral blood of normal donors, and targeted TNF bound to tumor cells mixed with monocytes isolated from peripheral blood of donors.

In a preferred embodiment, pretargeting features localization of an active agent-ligand conjugate to a prelocalized targeting moiety-anti-ligand. Consequently, steric availability of the ligand to bind to the pretargeted anti-ligand is required. For this reason, constructs structured in this manner, biotin-PEG-cytokine, are preferred for use in pretargeting protocols exploiting the biotin-avidin or biotin-streptavidin ligand/anti-ligand pair. PEGylation of the cytokine may sterically hinder the ligand-anti-ligand interaction in constructs of the form ligand or anti-ligand-cytokine-PEG. Consequently, ligand or anti-ligand is preferably conjugated to PEG rather than the cytokine itself.

To produce ligand- or anti-ligand-PEG-cytokine constructs, a bifunctional PEG molecule is required. An example of such a derivative is 3-amino-2-hydroxypropyl-PEG, available in either 3350 dalton or 20 kD sizes from Sigma Chemical Co., St. Louis, MO. Using this bifunctional PEG molecule, biotin-PEG, for example, is formed by reacting an active ester form of biotin (prepared in accordance with the procedures described elsewhere herein) with the amino group of the

bifunctional PEG. The hydroxyl group of the bifunctional PEG can then be modified by reaction with succinic anhydride to provide a carboxylate functional group. This functional group is then converted to active ester form using known techniques, including those described elsewhere herein, for subsequent reaction with amino moieties of the cytokine (e.g., lysine residues).

The utility of ligand- or anti-ligand-PEG-cytokine and PEG-cytokine-ligand or -anti-ligand constructs is limited to those cytokines for which PEG derivatization does not substantially impair function. If PEGylation diminishes function, the linkage between the cytokine and PEG in the construct is designed to be cleavable at the target cell site, thereby facilitating cytokine efficacy upon target site association. Suitable cleavable linkers have a cleavage half-life under physiological conditions of from about 2 to about 15 hours to allow target site localization followed by efficient cytokine release. Exemplary linkers of this type are disulfide, hydrazone, hydrazide, ester, and the like. Other suitable cleavable linkers are those that are stable at a pH that is greater than 4 or less than 7-8. Such cleavable linkers are known in the art. For example, U.S. Patent Nos. 5,017,693; 5,141,648; 5,094,848; and 5,171,563 discuss cleavable linkers as does European Patent No. EP 0 318 943.

Useful procedures for the preparation of constructs with cleavable linkages (biotin----PEG----cleavable linker----cytokine-containing constructs) involve reagents having hydroxy or amino functional groups, for example. More specifically, the active ester of SPDP (3-(2-pyridyldithio)propionic acid N-hydroxy succinimide ester available from Sigma Chemical Co, St. Louis, MO) reacts with an amine group of PEG to provide a precursor for a cleavable disulfide linkage. Then the hydroxy group of PEG is converted into active ester form (via

reaction with succinic acid anhydride or through other suitable techniques therefor). An amino derivative of biotin is then reacted with the active ester, and the free SPDP functional group is reacted with an endogenous
5 sulfhydryl group of the cytokine generated by reaction of the cytokine with a reducing agent such as DTT or the like, or with a sulfhydryl group introduced into the cytokine structure by modification of a lysine residue of the cytokine by agents such as iminothiolane or
10 Traut's reagent.

Amino-biotin is prepared in accordance with known techniques. For example, reaction of a commercially available NHS ester derivative of biotin, such as N-hydroxysuccinimido biotin or biotinamidocaproate N-
15 hydroxysuccinimide ester, with an excess of ethylene diamine gives an amide-linked biotin moiety bearing a free amine group for conjugation.

Cleavable linkages between the active agent and the DBM may be useful whenever the active agent requires
20 release to be active once targeted. Exemplary of such active agents are certain drugs.

The molecular weight of the PEG polymer employed as a DBM in accordance with the present invention may be adjusted to favor the desired rate of renal excretion or to decrease the reduction in target site localization
25 caused by liver uptake of non-targeted conjugate. Cytokines of interest are generally about 20 kD in size. Alterations in cytokine size will impact non-target localization and excretion of the modified cytokine. For example, higher molecular weight cytokine-containing
30 conjugates (prepared with 20kD-PEG available from Sigma Chemical Co.) will be excreted more slowly than such conjugates prepared with 3350 dalton PEG (also available from Sigma). Also, combinations of PEGs of different
35 molecular weight may be employed by a practitioner in the art to optimize cytokine localization and excretion for specific applications. These principles with regard

to polymer molecular weight selection are applicable to other polymeric DBMs discussed herein.

Alternatively, PEG may be employed as a linking moiety between another polymer-active agent and a ligand or anti-ligand. An exemplary conjugate of this embodiment of the present invention useful in two-step pretargeting protocols of the present invention which employ the biotin-avidin ligand-anti-ligand pair may be schematically represented as follows:

Biotin----PEG----cleavable linker---cytokine, such as
Biotin----PEG----glu----glu----cytokine.

In addition, dextran polymers of various molecular weights may also be employed as a DBM for cytokines in accordance with the present invention. For example, conjugates useful in two-step pretargeting protocols of the present invention which employ the biotin-avidin ligand-anti-ligand pair may be schematically represented as follows:

Dextran----cytokine----biotin;

or

Biotin----dextran----cytokine;

or

Biotin----dextran----cleavable linker----cytokine;

or

Biotin---dextran---cleavable linker---cytokine---biotin, wherein the number of biotins associated with each cytokine preferably ranges from 1 to about 3, and the number of dextrans associated with each cytokine preferably ranges from 1 to about 5, depending in part upon the size of the dextrans. The chemistry for the formation of cytokine-dextran conjugates is analogous to that described herein for other dextran conjugations.

The invention is further described through presentation of the following examples. These examples are offered by way of illustration, and not by way of limitation.

Example I

**Synthesis of Chelate-Ligand Conjugates and
DBM-Chelate-Radionuclide Conjugate Preparation:**

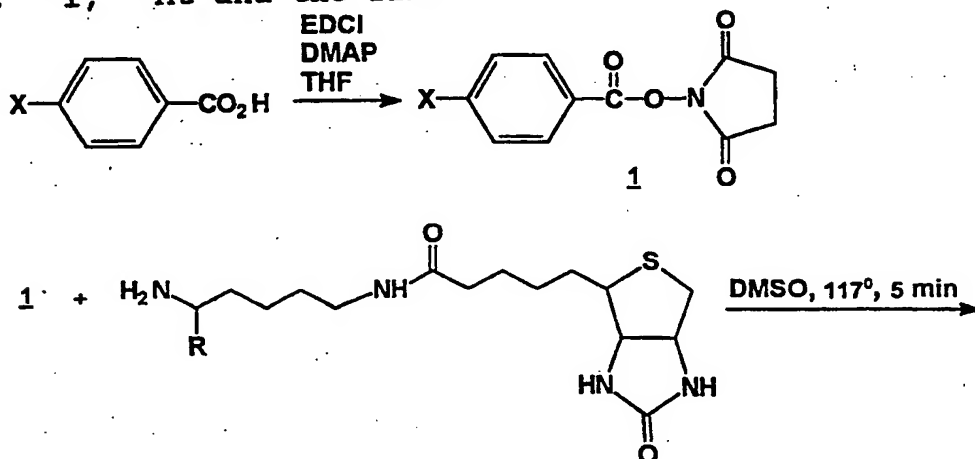
Tentacle Method

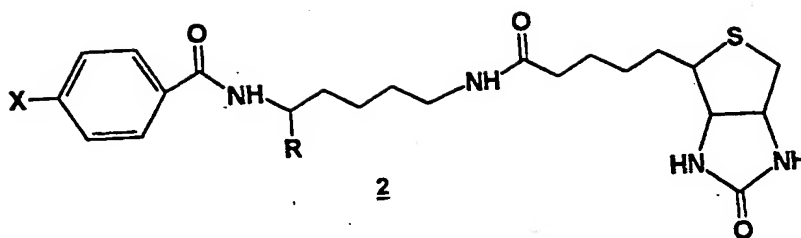
A. Preparation of Iodine Chelate-Ligand.

Radioiodinated biotin derivatives prepared by exposure of poly-L-lysine to excess NHS-LC-biotin and then to Bolton-Hunter N-hydroxysuccinimide esters in DMSO has been reported. After purification, this product was radiolabeled by the iodogen method (see, for instance, Del Rosario et al., J. Nucl. Med. 32:5, 1991, 993 (abstr.)). Because of the high molecular weight of the resultant radioiodinated biotin derivative, only limited characterization of product (i.e., radio-HPLC and binding to immobilized streptavidin) was possible.

Preparation of radioiodinated biotin according to the following method provides certain advantages. First, the radioiodobiotin derivative is a low molecular weight compound that is amenable to complete chemical characterization. Second, the disclosed methods for preparation involve a single step and eliminate the need for a purification step.

Briefly, iodobenzamide derivatives corresponding to biocytin (R = COOH) and biotinamidopentylamine (R = H) were prepared according to the following scheme. In this scheme, "X" may be any radiohalogen, including ^{125}I , ^{131}I , ^{123}I , ^{211}At and the like.





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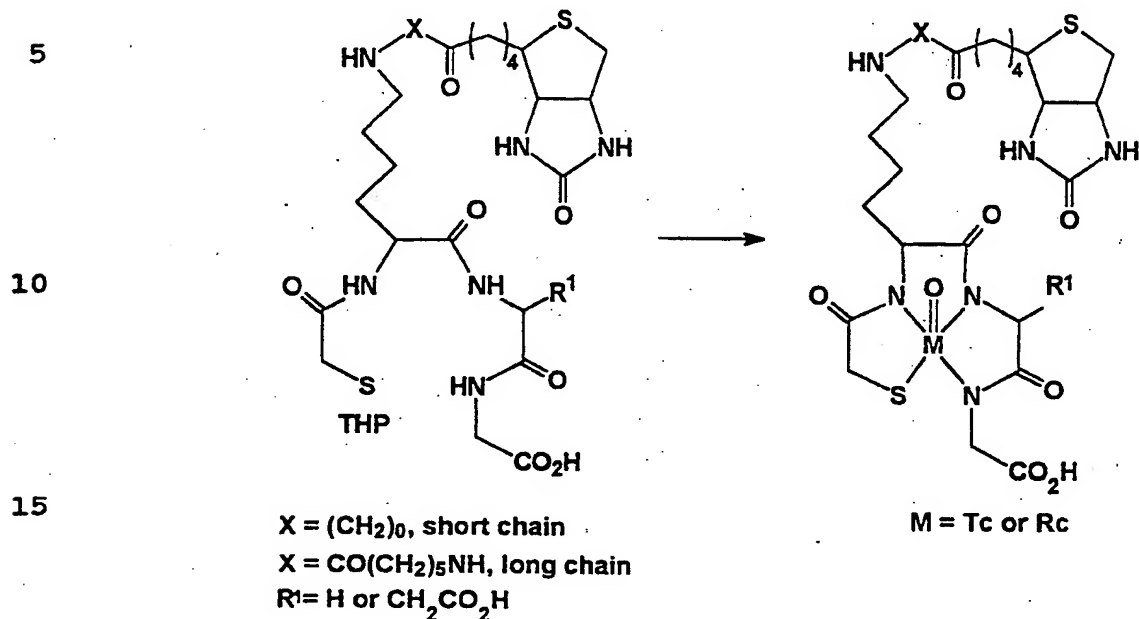
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Preparation of 1 was generally according to Wilbur et al., J. Nucl. Med. 30:216-26, 1989, using a tributyltin intermediate. Water soluble carbodiimide was used in the above-depicted reaction, since the NHS ester 1 formed intractable mixtures with DCU. The NHS ester was not compatible with chromatography; it was insoluble in organic and aqueous solvents and did not react with biocytin in DMF or in buffered aqueous acetonitrile. The reaction between 1 and biocytin or 5-(biotinamido) pentylamine was sensitive to base. When the reaction of 1 and biocytin or the pentylamine was performed in the presence of triethylamine in hot DMSO, formation of more than one biotinylated product resulted. In contrast, the reaction was extremely clean and complete when a suspension of 1 and biocytin (4 mg/ml) or the pentylamine (4 mg/ml) was heated in DMSO at 117°C for about 5 to about 10 min. The resultant ¹²⁵I-biotin derivatives were obtained in 94% radiochemical yield. Optionally, the radioiodinated products may be purified using C-18 HPLC and a reverse phase hydrophobic column. Hereinafter, the resultant radioiodinated products 2 are referred to as PIP-biocytin (R = COOH) and PIP-pentylamine (R = H).

B. Preparation of Technetium and Rhenium Chelate-Ligand.

A chelating compound that contains an N₃S chelating

core was attached via an amide linkage to biotin. Radiometal labeling of an exemplary chelate-biotin conjugate is illustrated below.



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The spacer group "X" permits the biotin portion of the conjugate to be sterically available for avidin binding. When "R¹" is a carboxylic acid substituent (for instance, CH_2COOH), the conjugate exhibits improved water solubility, and further directs *in vivo* excretion of the radiolabeled biotin conjugate toward renal rather than hepatobiliary clearance.

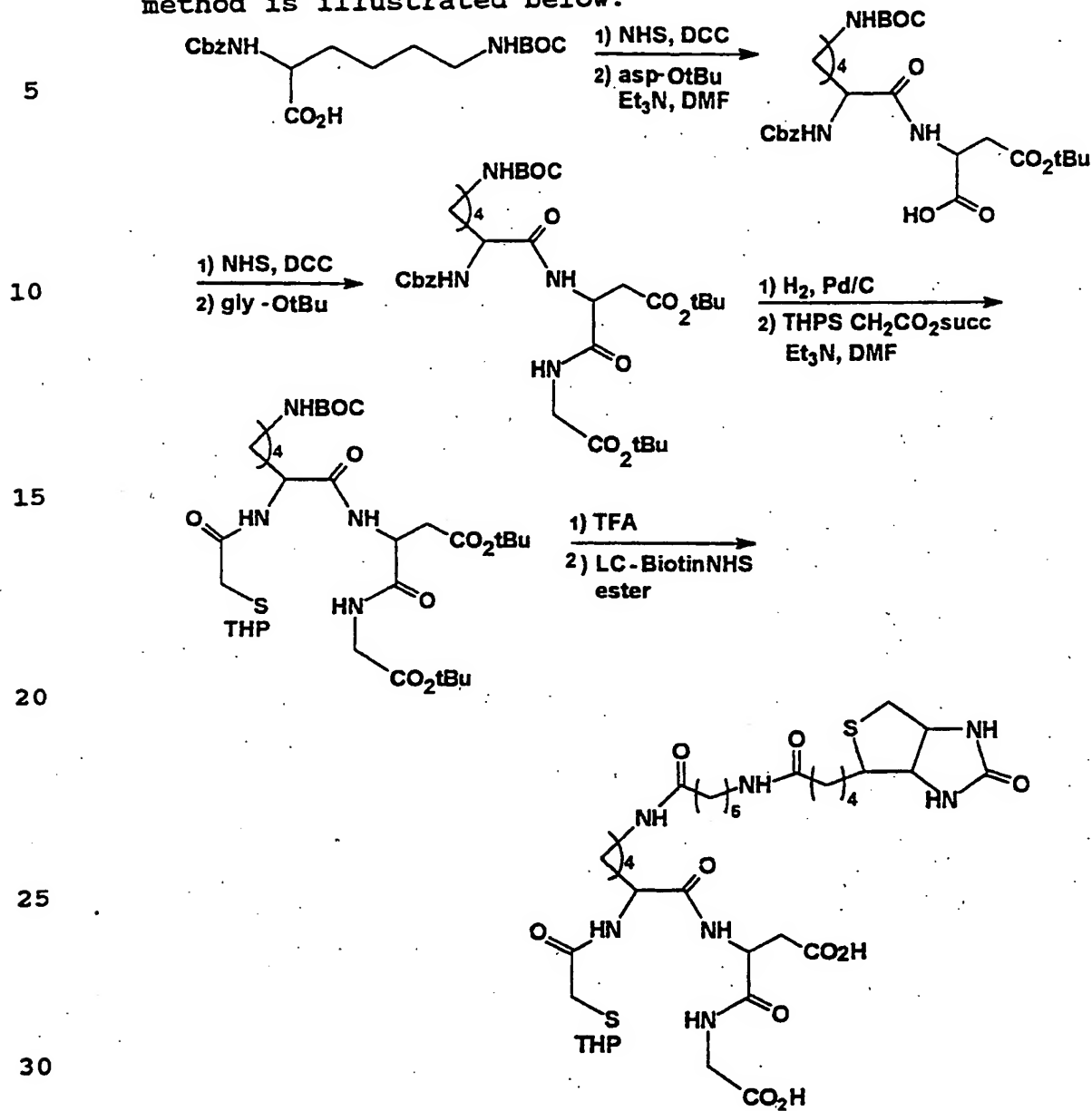
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Briefly, N- α -Cbz-N- ϵ -t-BOC protected lysine was converted to the succinimidyl ester with NHS and DCC, and then condensed with aspartic acid β -t-butyl ester. The resultant dipeptide was activated with NHS and DCC, and then condensed with glycine t-butyl ester. The Cbz group was removed by hydrogenolysis, and the amine was acylated using tetrahydropyranyl mercaptoacetic acid succinimidyl ester, yielding S-(tetrahydropyranyl)-mercaptoacetyl-lysine. Trifluoroacetic acid cleavage of the N-t-BOC group and t-butyl esters, followed by

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condensation with LC-biotin-NHS ester provided (Σ -caproylamide biotin)-aspartyl glycine. This synthetic method is illustrated below.



¹H NMR: (CD₃OD, 200 MHz Varian): 1.25-1.95 (m, 24H), 2.15-2.25 (broad t, 4H), 2.65-3.05 (m, 4H), 3.30-3.45 (dd, 2H), 3.50-3.65 (ddd, 2H), 3.95 (broad s, 2H), 4.00-4.15 (m, 1H), 4.25-4.35 (m, 1H), 4.45-4.55 (m, 1H), 4.7-5.05 (m overlapping with HOD).

Elemental Analysis: C, H, N for $C_{35}H_{57}N_7O_{11}S_2 \cdot H_2O$
calculated: 50.41, 7.13, 11.76
found: 50.13, 7.14, 11.40

5 The chelate-biotin conjugate was radiolabeled with
either ^{99m}Tc pertechnetate or ^{186}Re perrhenate. Briefly,
 ^{99m}Tc pertechnetate was reduced with stannous chloride in
the presence of sodium gluconate to form an intermediate
Tc-gluconate complex. The chelate-biotin conjugate was
10 added and heated to 100°C for 10 min at a pH of about
1.8 to about 3.3. The solution was neutralized to a pH
of about 6 to about 8, and yielded an N_3S -coordinated
 ^{99m}Tc -chelate-biotin conjugate. C-18 HPLC gradient
elution using 5-60% acetonitrile in 1% acetic acid
demonstrated two anomers at 97% or greater radiochemical
15 yield using δ (gamma ray) detection.

Alternatively, ^{186}Re perrhenate was spiked with cold
ammonium perrhenate, reduced with stannous chloride, and
complexed with citrate. The chelate-biotin conjugate
was added and heated to 90°C for 30 min at a pH of about
20 2 to 3. The solution was neutralized to a pH of about
6 to about 8, and yielded an N_3S -coordinated ^{186}Re -
chelate-biotin conjugate. C-18 HPLC gradient elution
using 5-60% acetonitrile in 1% acetic acid resulted in
radiochemical yields of 85-90%. Subsequent purification
25 over a C-18 reverse phase hydrophobic column yielded
material of 99% purity.

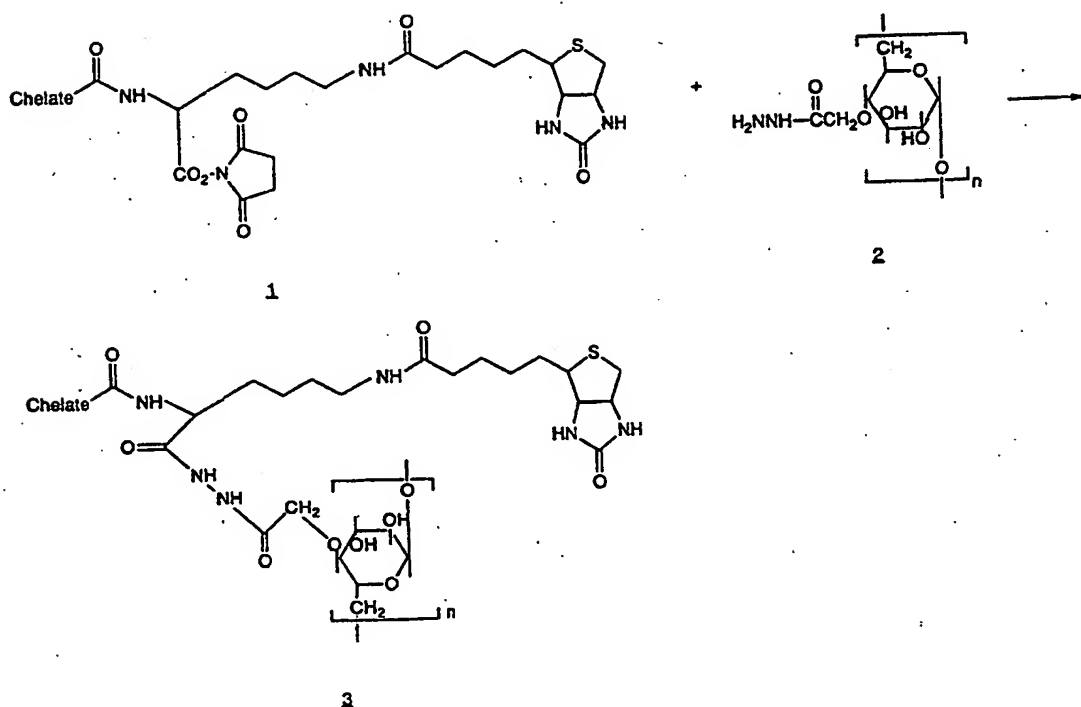
C. DBM-Chelate-Radionuclide Conjugate Preparation
Employing the Tentacle Chelate Method. Polylysine
30 (approximately 10,000 Dal. molecular weight, available
from Sigma Chemical Co., St. Louis, Missouri) and
dextran (lysine fixable, available from Sigma Chemical
Co.) were derivitized with SPDP and purified from
unreacted SPDP using size exclusion chromatography
35 (using a PD-10 column available from Pharmacia,
Piscataway, New Jersey). The resultant SPDP-derivitized
adducts were reduced with DTT in pH 4.7 0.2 M NaOAc

buffer to generate free reactive thiols. Reduced Tc, generated from stannous gluconate as described, for example, by Carlsson et al., *Biochem. J.*, **173**: 723-737, 1978, was added. A 90% incorporation of Tc was obtained for the polylysine adduct within 15 min, as measured by ITLC. 96% of the radioactivity coeluted with the dextran using size exclusion (PD-10) chromatography. These results are indicative of chelation.

Example II

Synthesis of Chelate-Ligand-Dextran-Conjugates

The reaction scheme for this synthesis is shown below.



where n ranges from about 50 (10,000 kD polymer) to about 500 (70,000 kD polymer). In this methodology, a chelate-ligand conjugate is prepared in accordance with any convenient procedure therefor (e.g., the procedures described in Examples I and IV hereof). Preferably the chelate-ligand conjugate structure includes a free carboxyl group. The free carboxy group is derivitized under conditions sufficient to generate an active ester (e.g., EDCI, DMAP and THF as described in Example I(A) hereof), with an exemplary active ester chelate-ligand starting material is shown as 1. Dextran hydrazide 2 is conjugated with 1 to give the product dextran-(chelate-ligand)_n 3.

Example III

Radiolabeling Biotin-Dextran, Lysine Fixable and Biodistribution of Radiolabeled Biotin-Dextran

A. Preparation Protocol. A solution of 3.0 mg biotin-dextran, lysine fixable (BDLF, available from Sigma Chemical Co., St. Louis, Missouri) in 0.3 ml PBS and 0.15 ml 1 M sodium carbonate, pH 9.25, was added to a dried residue (1.87 mCi) of N-succinimidyl p-I-125-iodobenzoate prepared in accordance with Wilbur, et al., J. Nucl. Med., 30: 216-226, 1989.

B. Analysis and Biodistribution. After 45 minutes at room temperature, the solution was passed through a G-25 gel filtration PD-10 column (Pharmacia) and 1 ml fractions were collected. Evaluation of fraction 4, containing 0.66 mCi (35%) of the total radioactivity, by HPLC (BioRad Biogel TSK 50-XL) showed 97.9% of the activity in a peak eluting at 11.47 minutes. By molecular weight standards, the elution time is consistent with a 10,000 molecular weight species. A single broad radiometric band at 13.4 minutes was observed by HPLC using a Sephacryl S-200 column.

Incubation of an aliquot of fraction 4 with immobilized streptavidin resulted in binding of 76% of the total radioactivity to the immobilized streptavidin. This result is indicative of the presence of intact biotin bound to the radioiodinated dextran.

Biodistribution studies were conducted on radiolabeled biotin-dextran and radiolabeled PIP-biocytin preparable in accordance with the procedure outlined in Example I. The results of those studies are shown in Figs. 1A (intestine), 1B (bladder) 1C (liver), 1D (kidney) and 1E (blood clearance). These results show that iodine labeled biotin-dextran exhibits a dramatic increase in urinary excretion in comparison to PIP-biocytin. In addition, the biotin-dextran molecule shows much lower hepatobiliary excretion than PIP-biocytin, indicating that the lipophilic biotin molecule is "masked" by the hydrophilic, polar dextran and is therefore subject to renal excretion.

C. Radiolabeling with Y-90. The free amino groups of biotin-dextran-lysine fixable (available from Sigma Chemical Co., St. Louis, MO) are succinylated with succinic anhydride in aqueous sodium bicarbonate. The carboxy groups of the resultant biotin-dextran-lysine-amido succinate are activated by conversion to NHS esters by using EDCI (1-(3-dimethyl-aminopropyl)-3-ethyl carbodiimide hydrochloride) and NHS. The resultant NHS esters are coupled to aminobenzyl-DOTA in pyridine and DMF. Aminobenzyl-DOTA is prepared substantially in accordance with the procedure set forth in Renn et al., Bioconj. Chem., 3:563-569, 1992, or any suitable alternative procedure therefore. The product, bis-DOTA-benzylamido-succinamido-lysyl-biotinyl-dextran, is purified by reverse phase C-18 preparative HPLC.

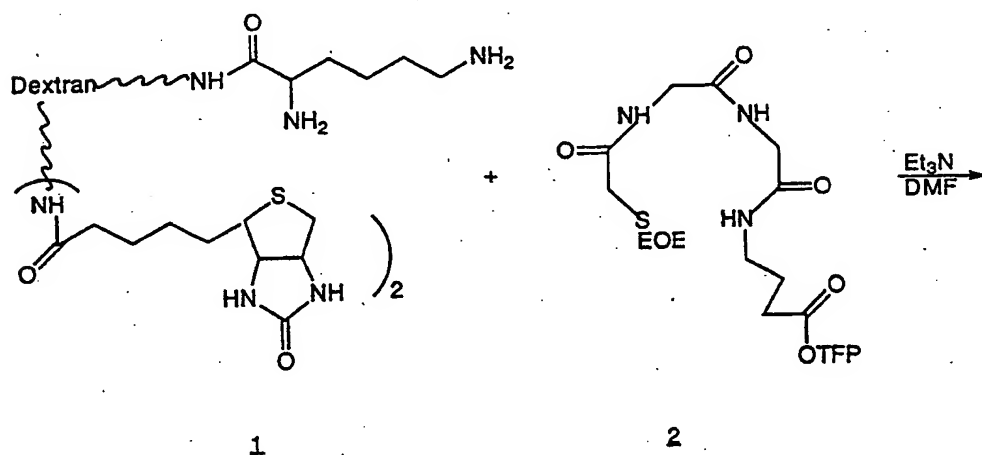
Carrier free $^{90}\text{YCl}_3$ (20-200 μL in 0.5N HCl) was obtained from DuPont (Wilmington, Delaware). The carrier free molecule was diluted with ammonium acetate buffer (0.5M, pH 5) to a total volume of 0.4 mL. Fifty

microliters (500 mg/mL) of ascorbic acid and 50-100 μ L (10 mg/mL) of the product DOTA-biotin conjugate prepared above are added to the buffered $^{90}\text{YCl}_3$ solution. Ascorbic acid is present as an antioxidant to prevent radiolysis of the preparation as discussed above. The mixture was incubated for one hour at 80°C. Upon completion of the incubation, 55 μ L of 100 mM DTPA (diethylenetriamine-pentaacetic acid) is added to the mixture to chelate any unbound Y-90. The final preparation was diluted to 10 mL with 0.9% NaCl.

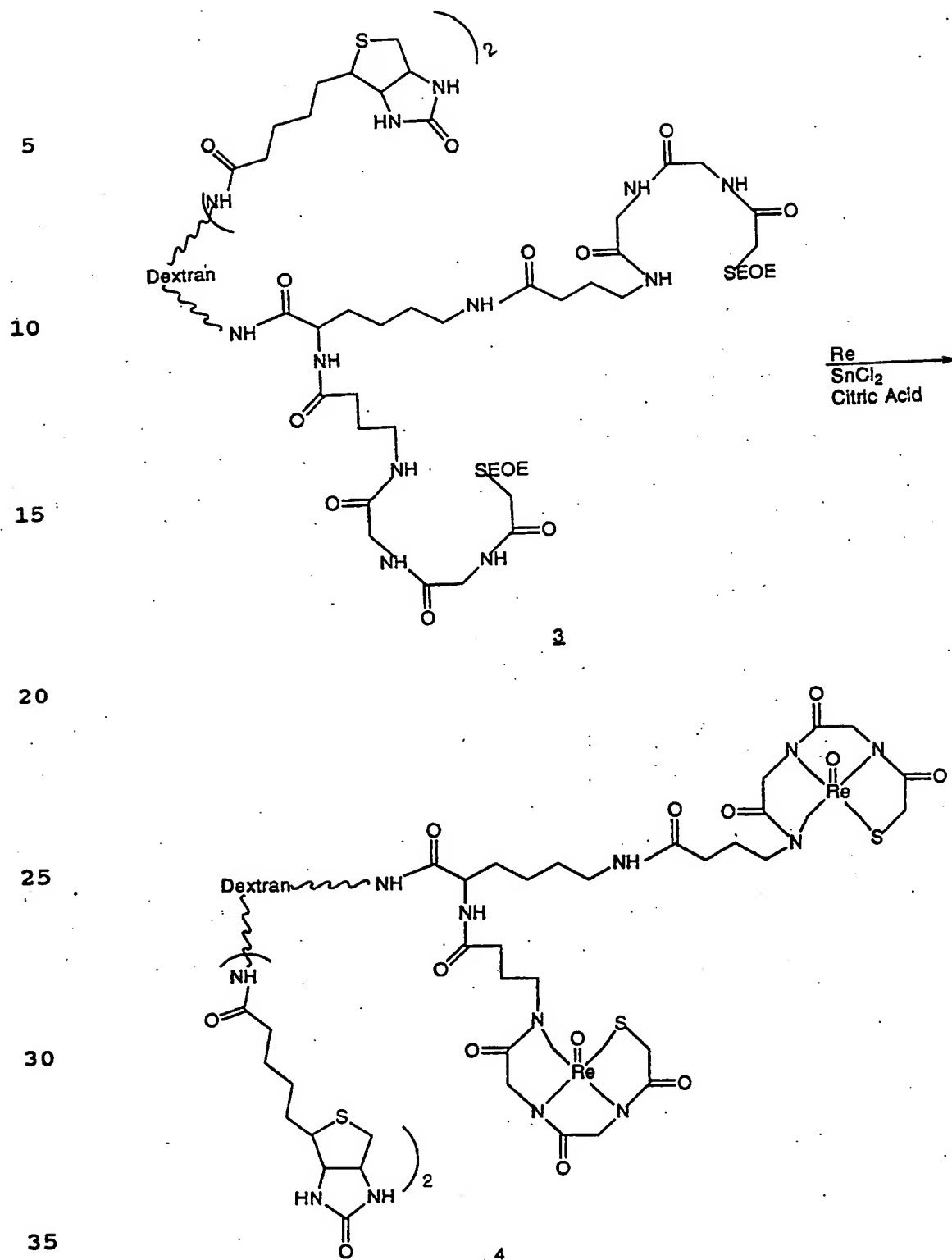
Example IV

Rhenium Radiolabeling of BDLF: The Pre-Formed and the Post-Formed Approaches; Technetium Labeling of BDLF

A. The Post-Formed Approach. The reaction scheme for this synthesis is shown below.

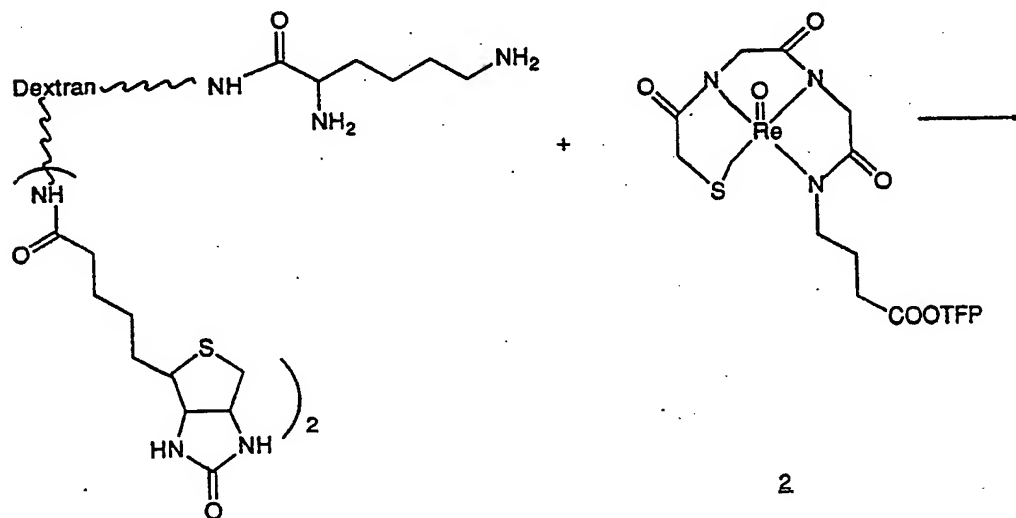


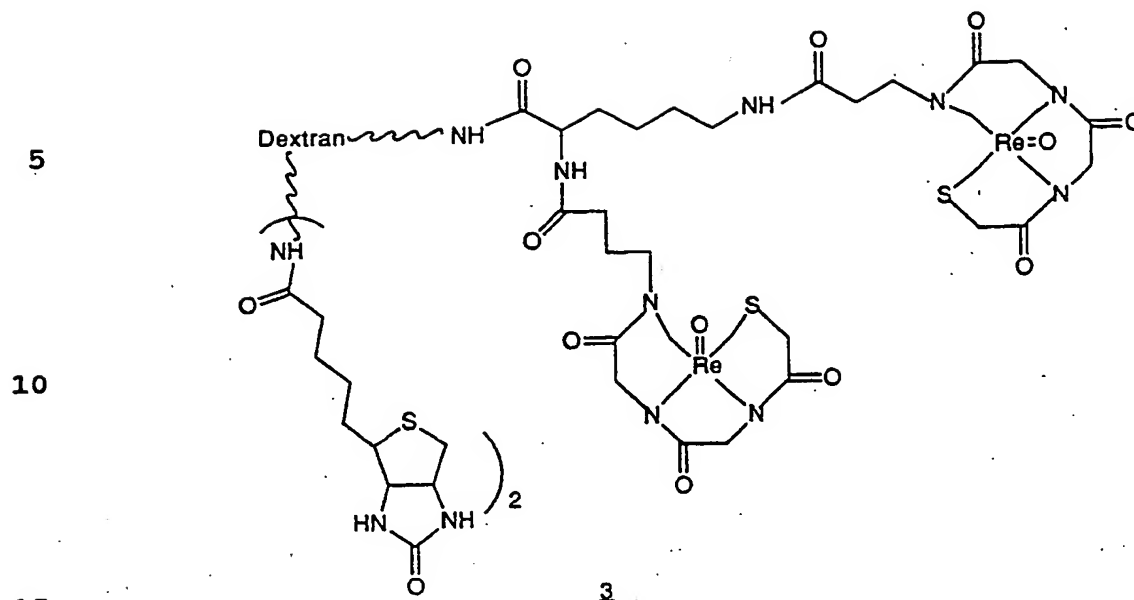
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BDLF 1 is condensed with an N_3S chelate tetrafluorophenyl ester 2 under basic conditions, e.g., Et_3N and DMF, Et_3N and DMSO or $NaHCO_3$ and water, to form a chelate-BDLF conjugate 3 through the formation of an amide bond, for example, at the lysine epsilon amino group of 1. Rhenium is complexed within the chelate component of 3 using known techniques therefor, e.g., reduction of perrhenate from aqueous solution conducted under elevated temperature in the presence of tin chloride and citric acid (or as otherwise described in U.S. Patent No. 4,965,392), to form the product radionuclide chelate-BDLF conjugate 4. Post-formed chelating methodologies are preferred for use with reactants and products that are stable to acidic conditions and heat that are required for the chelation step. Post-formed chelation is more convenient for kit formation, because it involves a single vial kit (i.e., radioactivity is added to a vial containing BDLF-chelated conjugate).

B. The Pre-Formed Approach. The reaction scheme for this synthesis is shown below.





BLDF 1 is reacted with a tetrafluorophenyl ester of a rhenium conjugated N₃S chelate 2 prepared, for example, in accordance with procedures described in U.S. Patent No. 4,965,392. This reaction preferably is conducted under basic conditions to facilitate the formation of the BDLF-radiolabeled chelate product 3, wherein the chelate is bound to a lysine epsilon amino group of BDLF. Exemplary conjugation conditions are also discussed in U.S. Patent No. 4,965,392. The pre-formed methodology has the following advantages: only a single step of the procedure for DBM-chelate conjugation involves the DBM and the DBM is not exposed to labeling conditions, which are potentially harmful to the DBM.

C. Technetium Labeling of BDLF. To a solution of 3.2 mg (0.32 μ mol) of BDLF (Sigma Chemical Co.) in 1.0 ml PBS and 0.1 ml 0.5 M borate buffer, pH 8.5, was added 50 μ l (2.5 mg, 8 μ mol) of a DMSO solution of N-succinimidyl 3-(2-pyridyldithio)propionate (SPDP). After 1 hour of mixing at room temperature, derivitized BDLF was isolated from unreacted SPDP by G-25 size

exclusion chromatography (PD-10, Pharmacia).

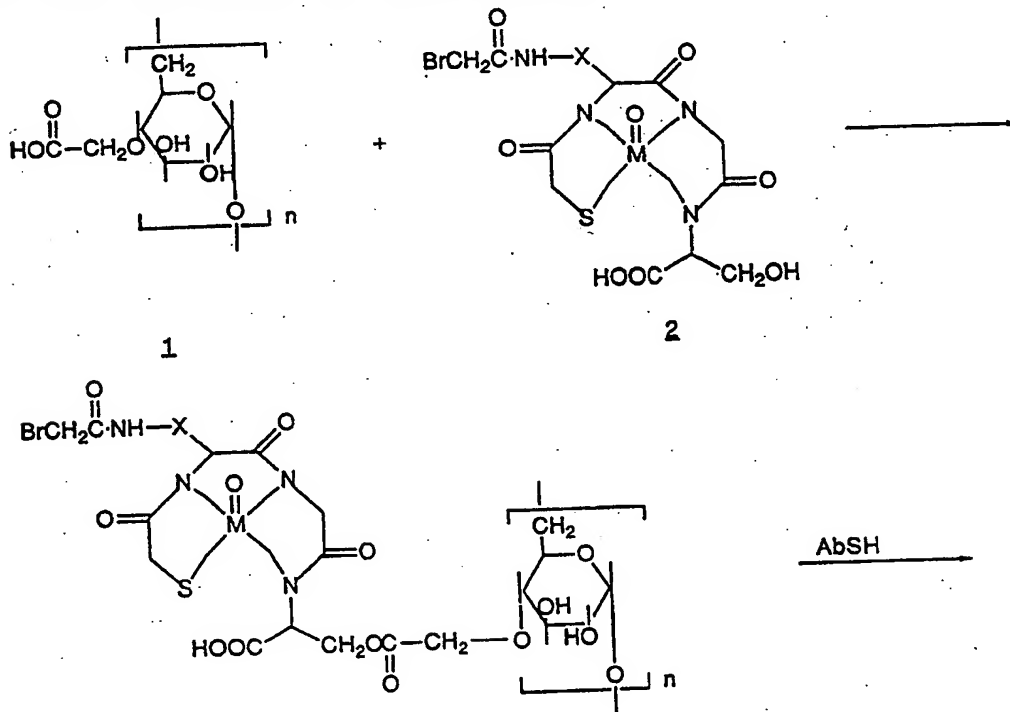
Approximately 1 mg of SPDP-BDLF (0.60 ml) was reduced by addition of 0.22 ml (2.25 mg) of dithiothreitol (DTT) in PBS. After mixing at room temperature for 0.5 hours, reduced SPDP-BDLF was isolated by G-25 (PD-10) size exclusion chromatography. A positive DTNB test on the isolated material was indicative of reactive thiols.

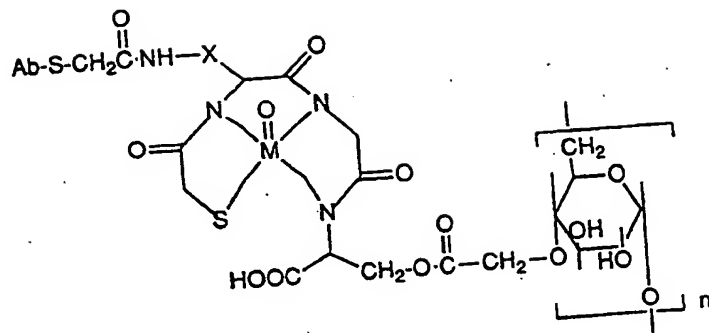
Tc-99m labeling was performed by addition of 2.1 mCi Tc-gluconate, which was prepared as discussed in Kasina et al., *J. Nucl. Med.*, 32:1445, 1991, to approximately 0.5 mg (1.0 ml) of reduced SPDP-BDLF. After 30 minutes at room temperature, the labeled adduct was isolated in 92% radiochemical yield by G-25 (PD-10) chromatography.

Example V

Radiolabeled Antibody-Dextran Conjugate Synthesis

A. From Carboxymethyl Dextran. The reaction scheme for this synthesis is shown below.



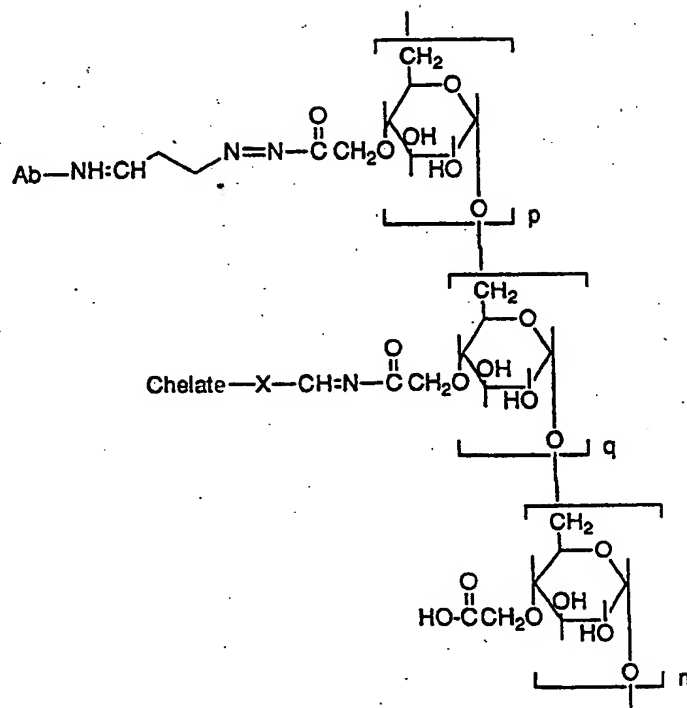
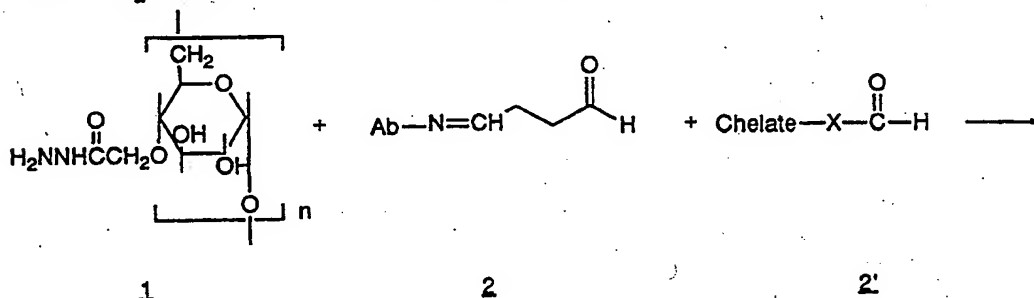


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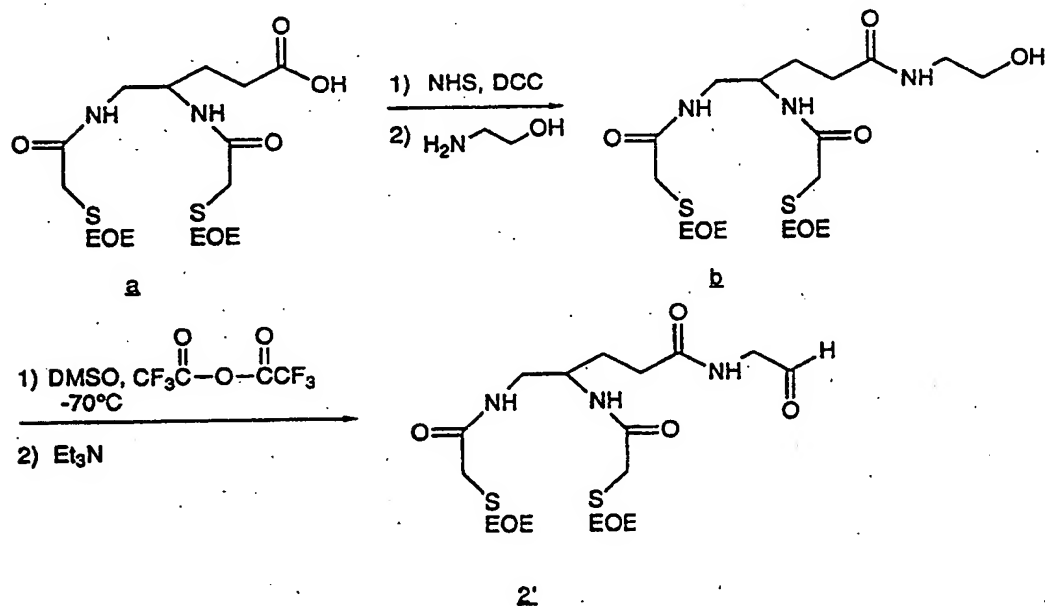
where X is selected from alkyl and amide, e.g., $-(CH_2)_4-$, m is selected from Tc and Re, and n ranges from about 50 to about 500. Carboxymethyl dextran 1 is reacted with a chelate compound 2 that is formed as described below. Alpha-t-BOC-lysine is condensed with bromo- (or iodo-) acetic acid N-hydroxy succinimidyl (NHS) ester to give epsilon-bromoacetamido-alpha-BOC lysine. Conversion of the epsilon-bromoacetamido-alpha-BOC lysine to an NHS ester by reaction with NHS and DCC. Condensation with glycine affords epsilon-bromoacetamido-alpha-BOC-lysyl-glycine. Conversion of the glycine compound to the NHS ester derivative (e.g., with NHS and DCC) followed by condensation with aminoethanol provides epsilon-bromoacetamido-alpha-BOC-lysyl-glycyl-amidoethanol. BOC cleavage with trifluoroacetic acid and condensation with S-ethoxy-ethyl mercapto-acetic acid NHS ester results in a S-ethoxylethyl-mercaptoacetyl-epsilon-bromoacetamido-lysyl-glycyl-amidoethanol chelate compound. The chelate is complexed with rhenium using standard labeling conditions described above (e.g., aqueous perrhenate

solution reduced with tin chloride in the presence of citric acid) to form 2. 1 and 2 are coupled using carbodiimide EDCI (1,3-dimethylaminopropyl)-3-ethyl carbodiimide hydrochloride) to form metal-chelate-dextran conjugate 3. 3 is conjugated to an antibody thiol group to displace the bromide on the chelate and give the thioether-antibody-radiolabeled chelate-dextran conjugate 4.

B. From Dextran Hydrazide. The reaction scheme for this synthesis is shown below.



where X is selected from $-(CH_2)_r-$, $-NHCO(CH_2)_r-$; r ranges from about 1 to about 10; n ranges from about 50 to about 500, p ranges from about 0.5 to about 1, and q ranges from about 49 to about 499. Dextran hydrazide 1 is conjugated to antibody aldehyde 2 and chelate-aldehyde 2' to form dextran hydrazone conjugate 3 substantially in accordance with the methodology employed by Hurwitz et al., *J. Med. Chem.*, 28:137, 1985, in the preparation of an antibody-dextran-5-FU conjugate. Antibody aldehydes 2 are formed by condensation of antibody amines with glutaraldehyde in accordance with known techniques therefor to give the antibody imine conjugate having a terminal aldehyde moiety. Chelate aldehyde 2' synthesis is shown below:



Chelate $C_5N_2S_2$ carboxylic acid a is condensed with NHS, DCC to give the chelate NHS ester. The NHS ester is condensed with aminoethanol to provide chelate-

amidoethanol b. Swern oxidation of b with DMSO and trifluoroacetic anhydride provides chelate aldehyde 2'.

Example VI

5 Somatostatin Derivative (Octreotide)-Dextran Synthesis

10 A. From Dextran Hydrazide. Briefly, suitably blocked octreotide, i.e., with the lysine amino (trifluoroacetamido-lysine for subsequent cleavage with piperidine) and both threonine hydroxy groups blocked, is acylated at the amino terminus by condensation with a chelate active ester, such as a tetrafluorophenyl ester in DMF and triethylamine. The resultant chelate-octreotide is activated with carbodiimide or converted to the N-hydroxysuccinimide ester and condensed with dextran hydrazide to give the acylhydrazide chelate-octreotide-dextran product.

20 B. From Carboxymethyl Dextran. A BOC-amino protected, trifluoroacetamide-lysine-octreotide carboxy succinimidyl ester is prepared in accordance with known procedures therefor. See, for example, UK Patent Application GB 2 225 579. The ester is condensed with chelate amine to provide chelate linked to the octreotide carboxy terminus by an amide linkage. Subsequent cleavage of the N-BOC protecting group with trifluoroacetic acid is undertaken. The resultant amine is condensed with carboxymethyl dextran using carbodiimide coupling chemistry (i.e., carboxymethyl dextran previously activated with EDCI) provides chelate-octreotide-carboxymethyl dextran with the octreotide amino terminus linked to the dextran carboxy moiety by an amide linkage.

35 C. From Oxidized Dextran. Dextran aldehyde is prepared by periodate oxidation of dextran. The octreotide amino terminus is linked by an amide linkage

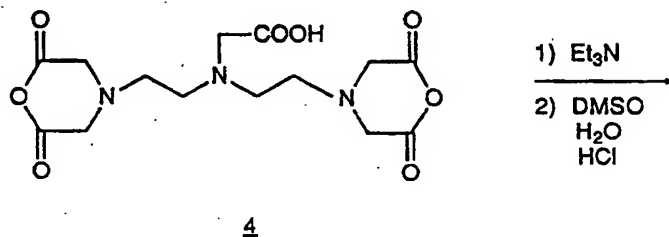
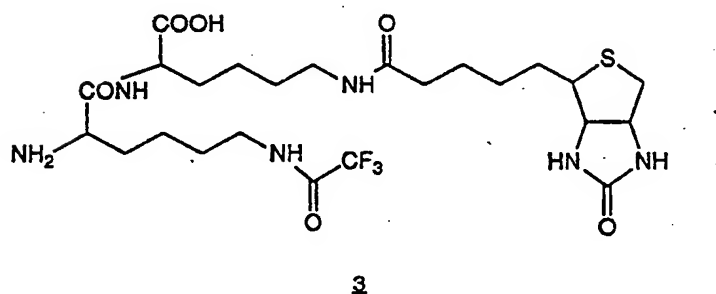
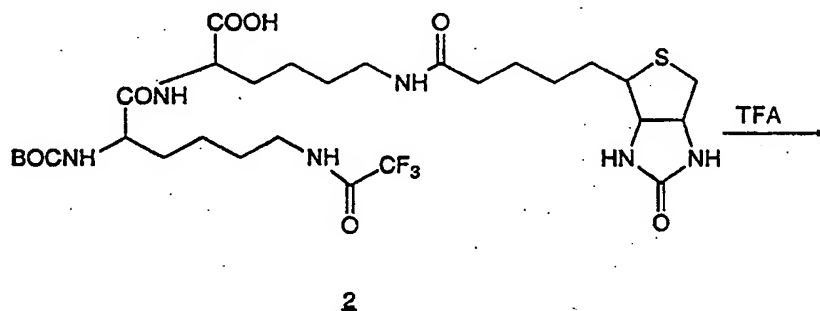
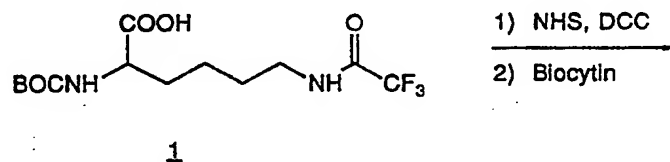
to a chelate carboxyl group by coupling suitably blocked octreotide to a chelate active ester, such as a trifluorophenyl ester. The carboxy terminus of octreotide is then converted to the methyl ester for conversion to a hydrazide upon reaction with hydrazine. It is worthy of note that the preparation of the chelate-octreotide-hydrazide can be conducted by first generating the carboxy terminus hydrazide and subsequently coupling the amino terminus to the chelate active ester. Conjugation of chelate-octreotide-hydrazide to the polyaldehyde dextran provides the acyl hydrazone product.

C. From Aminodextran. Methods of introducing amino groups on dextran are known. For example, Noguchi et al., Bioconj. Chem., 3: 132-137, 1992, suggests introducing a spacer through an ether linkage to a dextran hydroxyl group by reacting dextran with 6-bromohexanoic acid under basic conditions (4 M NaOH at 80°C for 3 hours). Next, ethylenediamine is reacted with the carboxyl group of the spacer arm of the dextran backbone using EDCI at pH 5. Another method for amino group introduction is discussed in Mann et al., Bioconj. Chem., 3: 154, 1992. In this method, a dextran hydroxy group is derivitized with acrylonitrile. The nitrile is subsequently reduced to form the amino dextran derivative.

The octreotide amino terminus is linked by an amide linkage to a chelate carboxyl group by coupling octreotide to a chelate active ester, such as a trifluorophenyl ester. The carboxy terminus of octreotide is then activated with EDCI and reacted with aminodextran to form an amide linkage between the dextran amine and the octreotide carboxy terminus and a chelate-octreotide-dextran product.

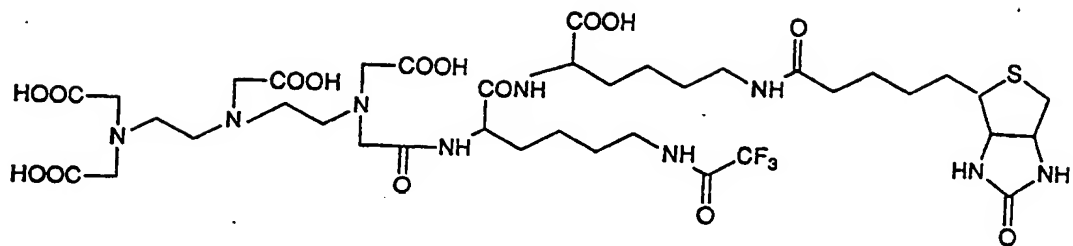
Example VII**Synthesis of DTPA/Biotin/Radionuclide Conjugate**

A. Synthesis-1 of DTPA-Biotin-Radionuclide. The synthesis of DTPA-biotin-radionuclide conjugate proceeds as follows:



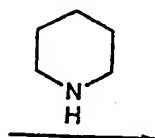
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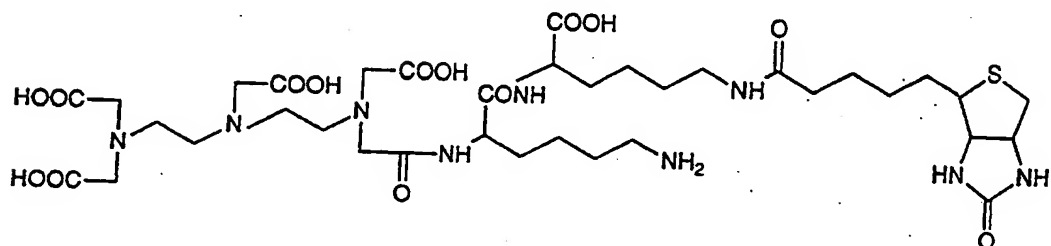


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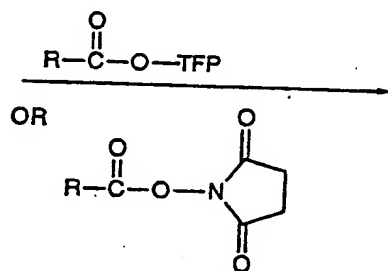


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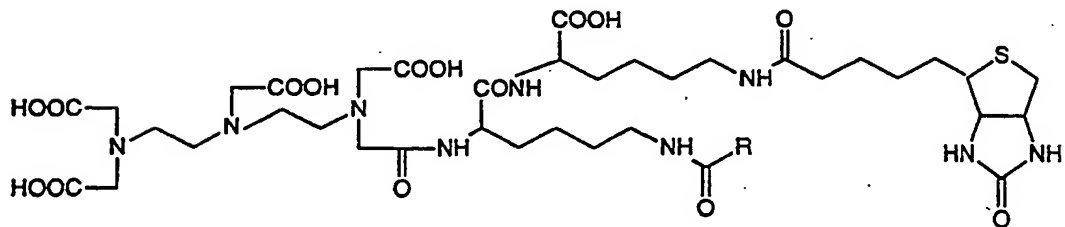
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N-alpha-BOC-N-epsilon-trifluoroacetamide (TFA)-lysine 1 was converted to a N-hydroxysuccinimidyl ester with NHS and DCC and then condensed with biocytin free base. The alpha amino group of the resultant lysylbiocytinamide derivative 2 was selectively deprotected with trifluoroacetic acid. A 5 molar excess of DTPA- dianhydride was added to the selectively deprotected product 3 to maximize the formation of the monoamide adduct 5. The epsilon-TFA group of the DTPA-lysylbiocytinamide 5 was cleaved with piperidine to form an amine 6, which is acylated with N-hydroxysuccinimidyl p-iodobenzoate to give the iodinated final product 7 (R is p-iodophenyl). Alternatively, Re-chelate-DTPA conjugate 7 is formed by acylation of the amino with Re-complexed chelate, Re-oxomercaptoacetylglycylglycyl-gamma-amino butyric acid tetrafluorophenyl ester.

(1) Synthesis of alpha-N-BOC-epsilon-N-trifluoroacetamido-lysine-biocytinamide 2. N-hydroxysuccinimide (486 mg, 4.22 mmol) and DCC (870 mg, 4.22 mmol) were added to an ice cold solution of N-alpha-BOC-N-epsilon-TFA-lysine (Bachem, Torrance, CA) (1.156g, 3.38 mmol) in acetonitrile (7.0 ml). The reaction was warmed to 23°C and stirred for 1 hour. Glacial acetic acid (10 drops) was added to quench excess DCC. The reaction mixture was filtered. The filtrate was evaporated to give the N-alpha-BOC-N-epsilon-TFA-lysine succinimidyl ester as a white solid (1.43g, 100% yield). The product was used without purification. The product and biocytin (1.38g, 3.71 mmol) were suspended in DMSO (15 ml). The suspension was heated at 118° for 15 minutes until all the solids dissolved. The solution was applied to a 45 mm column (Kontes) filled to a height of eight inches with C-18 reverse phase silica (VWR). The column was eluted with one bed volume each of 80%, 40% and 50% methanol/water. The product was obtained in 56% yield.

¹H NMR (DMSO): 1.20-1.80 (m, 21H, alkyl CH₂), 2.05 (t, 2H, CH₂CO), 2.75 (dd, 2H, SCH₂), 3.05 (dd, 2H, CH₂NHCO), 3.15 (dd, 2H, CH₂NHTFA), 3.90 (broad s, 1H, CHNHBOC), 4.15 (m, 2H, 2 x CHN), 4.32 (m, 1H, CHN), 6.35 (s, 1H, NHCONH), 6.42 (s, 1H, NHCONH), 6.85 (d, 1H, NHBOC), 7.75 (t, 1H, HNCO), 7.95 (d, 1H, NHCO), 9.40 (t, 1H, NHTFA). Proton assignment was confirmed by decoupling experiments.

(2) Synthesis of epsilon-trifluoroacetamido-biocytinamido lysine 3. 10 ml of a 50% methylene chloride: trifluoroacetic acid solution was added to 0.94g of 2. The solution was stirred at 23°C for 1 hour, then evaporated. The residue was purified by reverse phase C-18 flash chromatography to give 3 as a fluffy white solid, quantitative yield.

¹H NMR (DMSO): 1.20-1.80 (m, 18H, alkyl CH₂), 2.05 (t, 2H, CH₂CO), 2.70 (dd, 2H, SCH₂), 3.3-3.85 (m, integration obscured by water in DMSO), 4.05-4.35 (m, 4H, 4 x CHN), 6.40 (broad d, 2H, NHCONH), 7.80 (t, 1H, HNCO), 8.15 (m, 3H, NH₂), 8.65 (d, 1H, NHCO), 9.40 (t, 1H, NHTFA).

(3) Synthesis of biocytinamido-epsilon-trifluoroacetamido-N-DTPA-lysine amide 5. Triethylamine (0.44 ml, 3.19 mmol) was added to a solution of 3 (54 mg, 0.79 mmol) in DMSO (3.0 ml). Diethylene triamine pentaacetic acid dianhydride (DTPA) (1.03g, 3.19 mmol) was added all at once to the reaction solution. The reaction was stirred at 23°C for 90 minutes and then quenched by the addition of water (5.0 ml). Stirring was continued for 45 minutes. The mixture was filtered.

The solid was washed with 50% methanol/water and then methanol (50 ml of each). The filtrate was evaporated, and the residue was purified by reverse phase C-18 flash chromatography to give 5 as a white solid in 43% yield.

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¹H NMR (DMSO): 1.10-1.80 (m, 18H, alkyl CH₂), 2.80-4.0 (m, integration not calculated because of overlapping solvent peaks), 4.15 (m, 1H, CHN), 4.25 (m, 1H, CHN), 6.35 and 6.45 (2 x s, 2 x 1H, NHCONH), 7.85 (t, 1H, HNCO), 8.25 (d, 1H, NHCO), 8.40 (d, 1H, NHCO), 9.70 (t, 1H, NHTFA). Single peak on reverse phase HPLC, retention time 27.6 min with a gradient t=0, 30% A, 70% B; t=60 min, 100%A where A = 60% CH₃CN/H₂O 0.1% TFA and B = 99.9% H₂O, 0.1% TFA.

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(4) Synthesis of biocytinamido-N-alpha-DTPA-lysine amide 7. A solution of 1 M piperidine in THF/H₂O, 1:1 (5.0 ml) was added to solid 5 (100mg). The reaction solution was stirred at 23°C for 60 hours, acidified by the addition of glacial acetic acid and concentrated. The residue was purified by reverse phase flash chromatography twice. The produce (32mg) was obtained as a white solid in 35% yield.

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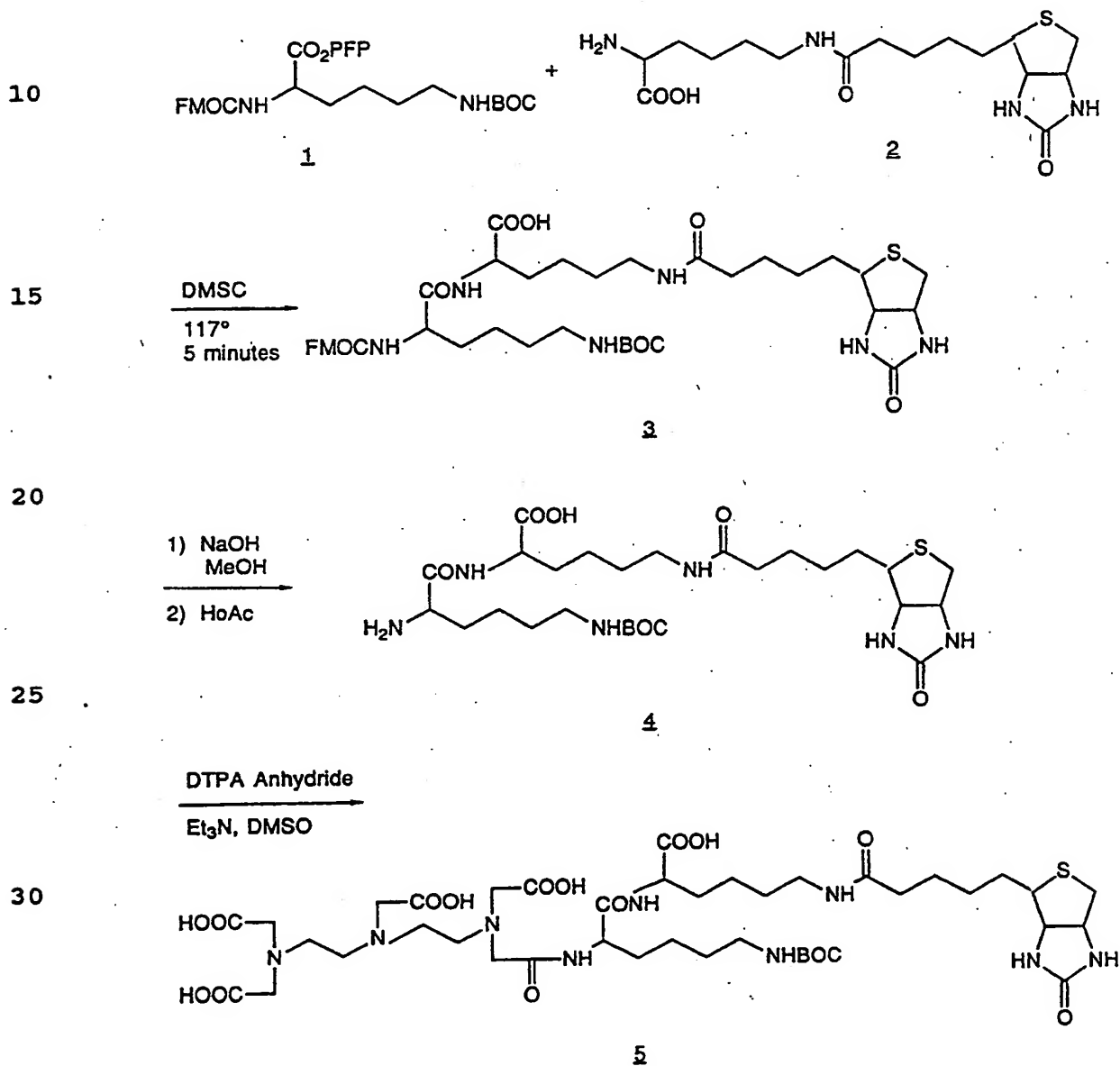
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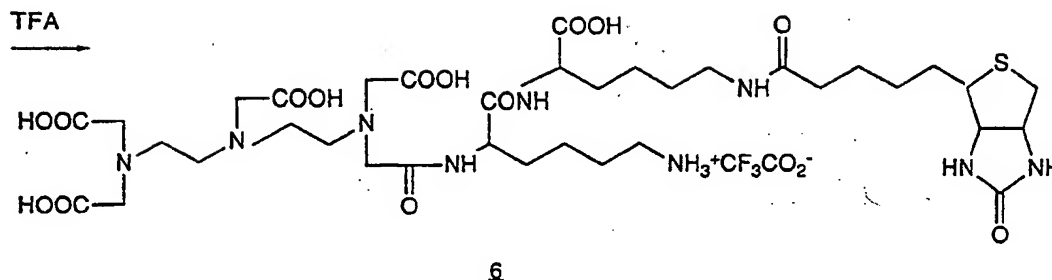
¹H NMR (DMSO): 1.20-1.80 (m), 2.05 (t, CH₂CO), 2.80-4.15 (broad multiplet obscured by water in DMSO), 4.15 (m, CHN), 4.35 (m, 1H, CHN), 6.40 and 6.55 (2 singlets, NHCONH), 8.00 (broad s, HNCO), 8.15 (broad s, NHCO), 9.05 (broad s, NHCO). Single peak on reverse phase HPLC, retention time 7.94 min with a gradient t=0, 30% A, 70% B; t=60 min, 60%A, 40%B where A

= 60% CH₃CN/H₂O 0.1% TFA and B = 99.9%
H₂O, 0.1% TFA.

5 B. Synthesis-2 of DTPA-Biotin-Radionuclide.
Alternatively, the synthesis of DTPA-biotin-radionuclide
conjugate proceeds as follows:



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A solution of N-alpha-FMOC-N-epsilon-BOC-lysine pentafluorophenyl ester 1 and biocytin 2 in DMSO was heated at 117° for 5 minutes to give N-alpha-FMOC-N-epsilon-BOC lysylbiocytinamide 3. The FMOC group was cleaved by dissolving 3 in a 1:1 solution of 2N NaOH:MeOH and stirring at room temperature for 20 minutes. The reaction solution was washed with toluene to remove FMOC reagent byproduct. The aqueous layer was evaporated to give the amine 4 which was used in the next reaction step without purification. A solution of 4 and a 5 molar excess of DTPA-dianhydride with triethylamine in DMSO is stirred at room temperature to give the monoamide 5. Deprotection of the epsilon-BOC amine with trifluoroacetic acid affords the final product 6.

25

(1) Synthesis of alpha-N-FMOC-epsilon-BOC-lysyl biocytinamide 3. A mixture of alpha-N-FMOC-epsilon-BOC-lysine pentafluorophenyl ester (1.153g, 1.82 mmol) (Sigma Chemical Company) and biocytin (0.75g, 2.00 mmol) (Sigma Chemical Company) in DMSO (8.0 ml) was heated at 117°C for 5 minutes until all of the solids dissolved. The product was purified by reverse phase C-18 flash chromatography (45 mm column packed with 8 inches of C-18 silica, eluted with one bed volume each of 30%, 50% and 70% methanol/water) to give 3 as a white solid (0.889g) in 59% yield.

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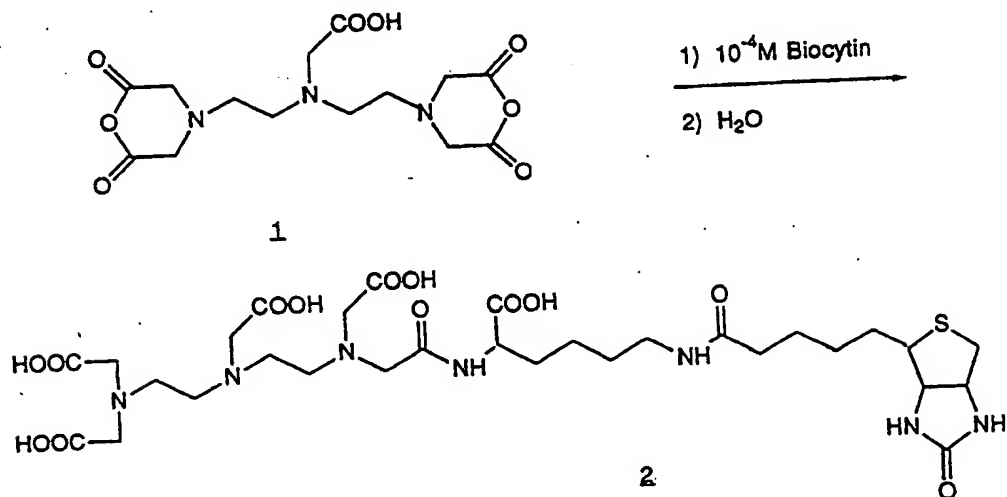
¹H NMR (DMSO): 1.15-1.80 (m, 27H, OC(CH₃)₃ and alkyl

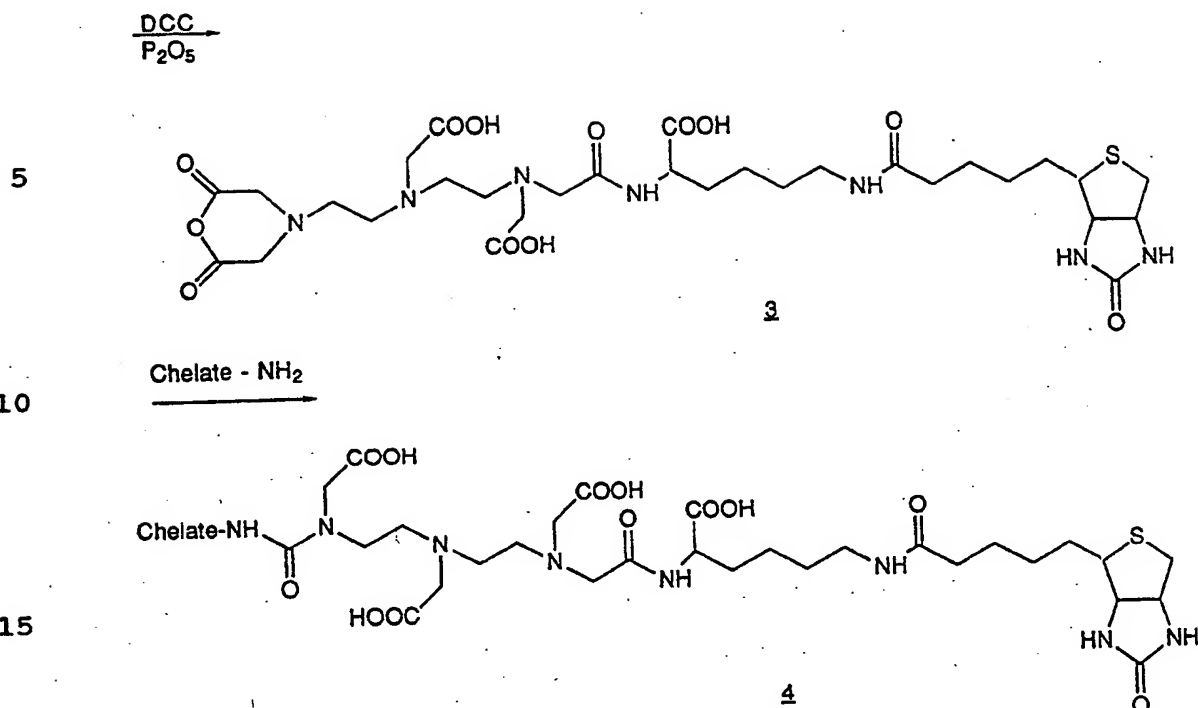
CH₂ groups), 2.05 (t, 2H, CH₂CO), 2.75 (dd, 2H, SCH₂), 2.85-3.15 (m, 4H, CH₂NCO x 2), 3.98-4.47 (m, 6H, CHN x 4, FmocCH₂), 6.35 (s, 1H, NHCO), 6.45 (s, 1H, NHCO), 6.80 (t, 1H, HNCO), 7.30-7.55 (m, 5H), 7.70-8.25 (m, 6H).

(2) Synthesis of epsilon-BOC-lysyl biocytinamide 4. A 1:1 solution of 2 N NaOH and methanol (12.0 ml) was added to 3 (231mg, 0.28 mmol). The suspension was stirred at 23°C for 30 minutes. The stirred suspension was acidified by the addition of glacial acetic acid and concentrated. The residue was partitioned between toluene and water. The aqueous layer was evaporated to give 4 containing NaCl.

¹H NMR (D₂O): 1.30-2.00 (m, 27H, OC(CH₃)₃ and alkyl CH₂ groups), 2.35 (t, 2H, CH₂CO), 3.05 (dd, 2H, SCH₂), 3.10-3.25 (m, 4H, NCH₂ x 2), 3.35 (dd, 1H, CHS), 4.05 (dd, 1H, CHN), 4.55 (dd, 1H, CHN), 4.65 (dd, 1H, CHN).

C. Synthesis of Radionuclide-DTPA-Biotin. The preparation of Radionuclide-DTPA-Biotin conjugates proceeds as shown below:





attached to the carboxyl group of lysine. One lysine amino group is attached to a DTPA carboxylate. The remaining lysine amino group is acylated with bromoacetic acid NHS ester for subsequent attachment of antibody thiols. Thus, displacement of the bromide provides a thioether bond to the antibody.

N-alpha-FMOC (fluorenomethyl)-lysine (Bachem, Torrance, CA) is condensed with bromoacetic acid N-hydroxysuccinimidyl ester to give epsilon-bromoacetamido-N-alpha-TFA-lysine. Conversion to the N-hydroxysuccinimidyl ester is accomplished by reacting the compound with NHS and DCC. Condensation with S-ethoxyethylmercaptoacetyl-glycylglycyl-gamma-aminobutyryl ethylene diamine (formed by reacting a tetrafluorophenyl ester of S-ethoxyethylmercaptoacetyl-glycylglycyl-gamma-aminobutyric acid with ethylene diamine in CH_3CN) provides N_3S chelate-lysine bromoacetamide. Deprotection of the amine by reaction with piperidine provides a free amine for reaction with DTPA dianhydride. This reaction opens the DTPA dianhydride and provides a DBM-chelate conjugate having a DTPA carboxylate attached by an amide linkage to a lysine amino group associated with the chelate. The N_3S core is radiolabeled with rhenium under the post-formed reaction conditions described above. The rhenium-chelate-DTPA conjugate is conjugated to an antibody thiol group by displacement of the bromide to form a thioether bond, also as described above.

Example IX

Radiolabeled Polyglutamyl-Antibody Conjugate Synthesis

A. Antibody-DBM Stable Linkage. The N-terminus of the polyglutamyl molecule is acylated with N-t-BOC glycine-N-hydroxy-succinimidyl ester to give polyglutamic acid monofunctionalized with a protected amine. The protected amine permits selective terminal

monofunctionalization to a reactive isothiocyanato group for attachment to a single antibody. A polymer molecule bearing a uniquely reactive terminus for univalent attachment to biomolecules overcomes the problem of crosslinking antibodies and resultant aggregation as discussed in Mann et al., Bioconjugate Chem., 3:154-159, 1992.

The BOC-glycineamidopolyglutamic acid derivative is then coupled to a chelate-amine, e.g., S-ethoxyethylmercaptoacetyl-glycylglycyl-gamma-aminobutylamidoethylenediamine as described above, using EDCI as a coupling agent to give a polyglutamic acid derivative with n carboxylates attached to chelate amino groups and p free carboxylates. The chelate is labeled with rhenium under standard conditions therefor as described above. The BOC protecting group of the DBM-chelate conjugate is cleaved with formic acid, with the resultant free amino group being converted to an isothiocyanate with thiophosgene in bicarbonate. An antibody amino group is coupled to the isothiocyanate under basic conditions to give the thiourea-linked antibody-DBM-chelate conjugate. Such conjugates have a stable antibody-DBM linkage.

B. Antibody-DBM Acid Labile Linkage. BOC-polyglutamic acid is coupled to a chelate amino group using EDCI to give N-BOC-polyglutamate-chelate conjugate. This conjugate contains n carboxylates attached to chelate amino groups and p free carboxylates. Formic acid deprotection of the BOC group and condensation with N-BOC(hydroxylamino)acetic acid NHS ester gives the BOC-protected terminal alkoxy amino polyglutamate-chelate conjugate. The chelate is labeled with rhenium under standard conditions therefor as described above. BOC-deprotection with formic acid and condensation with antibody aldehyde gives a alkoximine-linked antibody-DBM-chelate conjugate.

Alternatively, Cbz (carbobenzyloxy) groups may be employed as protecting groups in the synthesis of alkoximine linked antibody-DBM-containing conjugates. When Cbz groups are employed, trimethylsilyliodide is used in the deprotection reaction.

The alkoximine linker is susceptible to hydrolysis in the liver as well as in the acidic environment produced by lysosomes inside target cells (e.g., tumor cells). Provided the antibody-DBM-chelate conjugates exhibit good serum stability, improved targeting is expected with alkoximine-linked conjugates.

C. Antibody-DBM Enzyme Cleavable Linkage. Mono-t-butylsuccinate is coupled to Cbz-aminoethanol with isopropenyl chlorocarbonate (Et_3N , DMAP) to give a t-butyl aminoethyl succinate ester. Trifluoroacetic acid cleavage of t-butyl ester and activation with NHS and DCC provides a 2-Cbz-aminoethyl-mono-NHS succinate ester. The NHS ester is coupled to polyglutamic acid to form an N-terminal functionalized polyglutamate derivative. The carboxyl groups of polyglutamate are activated with EDCI and coupled to chelate amine groups. The chelate is labeled with rhenium under standard conditions therefor as described above. Trimethylsilyliodide cleavage of the Cbz protecting group provides a free amino group which can either be directly coupled to antibody aldehyde groups or converted to an isothiocyanate with thiophosgene. The latter protocol is completed by formation of a thiourea conjugate by reacting an antibody amino group with the DBM isothiocyanate moiety.

This conjugate of the present invention exhibits monofunctionalization of the N-terminus of the DBM, the advantage of which (prevention of the formation of cross-linked conjugates) having been discussed above. The aminoethyl ester linker is expected to be susceptible to esterases in the liver and kidney.

Cleavage of this linker releases the radioactivity on the DBM (*i.e.*, decouples the radioactivity from the antibody targeting moiety).

Example X

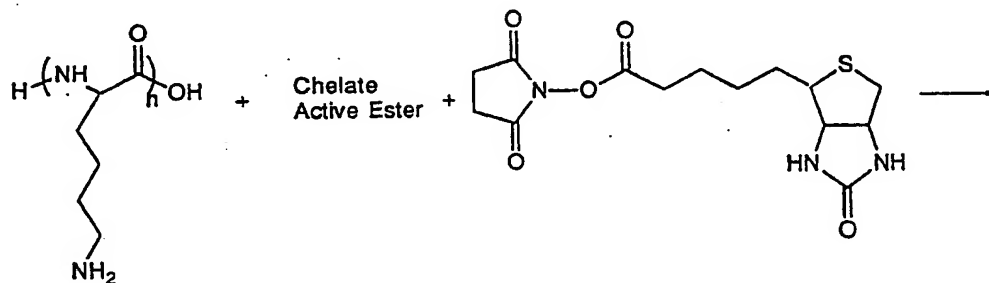
Radiolabeled Polyglutamyl-Biotin Conjugate Synthesis

A polyglutamate amine group is acylated with biotin-NHS ester which is prepared in accordance with conventional chemistry therefor or obtained from Sigma Chemical Company. This acylation results in an amide-linked DBM-ligand conjugate. Polyglutamate carboxylates are then activated with EDCI and coupled to chelate amine groups to form the product chelate-DBM-ligand conjugate. Optionally, additional biotin molecules can be linked to the DBM by coupling EDCI-activated polyglutamyl carboxylates to biocytin. The chelate is labeled with radionuclide under standard conditions therefor.

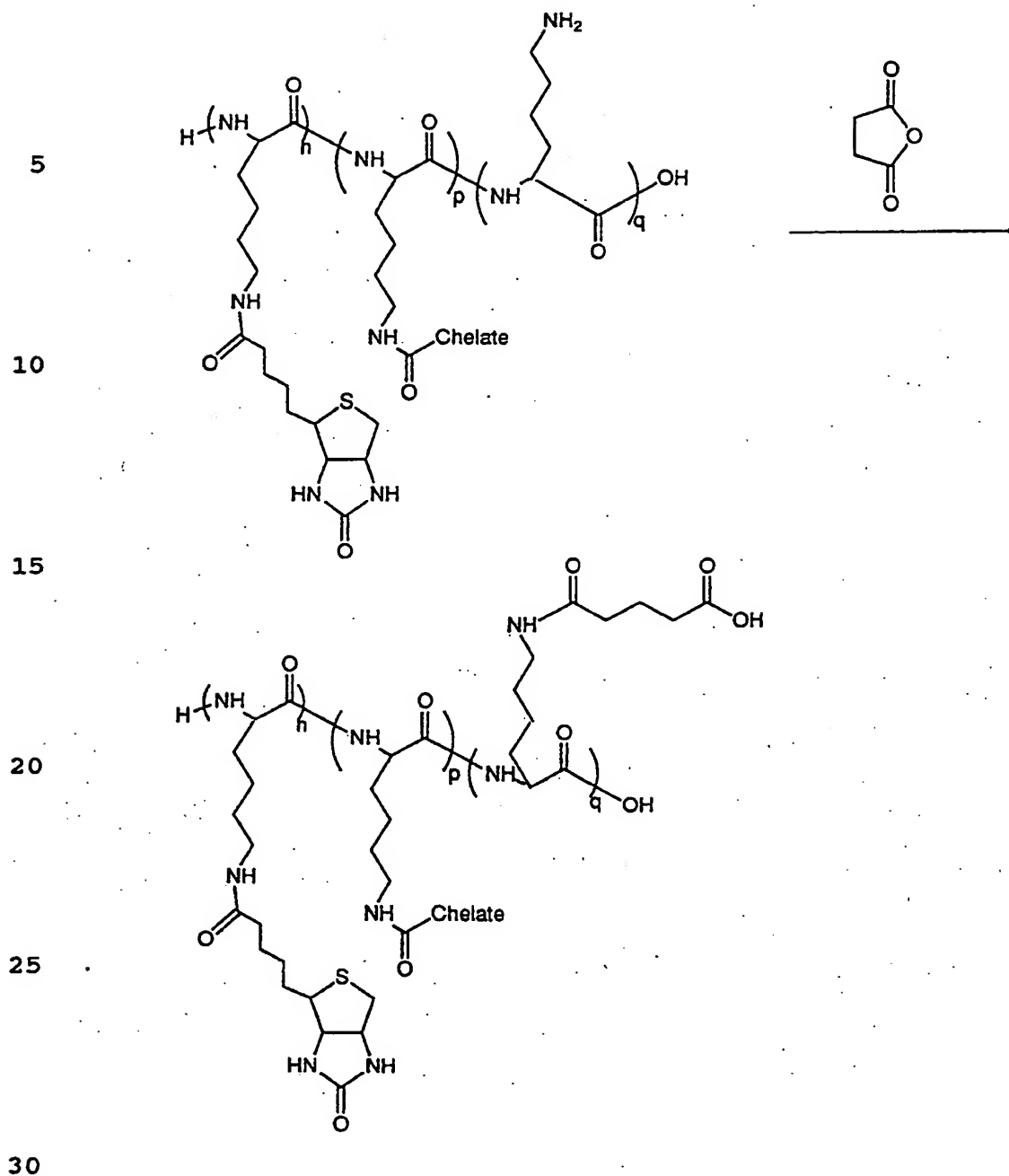
Example XI

Radiolabeled Succinylated Polylysine-Biotin Conjugate Synthesis

Chelate-succinylated polylysine-biotin conjugates are preparable in accordance with the reaction scheme shown below:



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Polylysine is acylated with p moles of chelate active ester to form a DBM (polylysine)-chelate conjugate. The DBM-chelate conjugate is then acylated with n moles of biotin NHS ester to form a biotin-DBM (polylysine)-chelate conjugate. The remaining amino groups of polylysine are capped with succinic anhydride to form a biotin-DBM (succinylated polylysine)-chelate conjugate.

Example XII**Targeted, Direct Labeled Protocol**

5 A patient is suspected of having lung cancer. A NR-
LU-10 monoclonal antibody, specific for a pancarcinoma
antigen, is conjugated to carboxymethyl dextran. The
antibody-DBM conjugate is further derivitized with 3-5
chelates, which have been previously radiolabeled with
a diagnostic radionuclide, to form an antibody-DBM-
10 (radionuclide chelate)_{3,5} conjugate. The final conjugate
is administered to the patient in a diagnostically
effective dose and in a pharmaceutically acceptable
diluent. The antibody directs the conjugate to the
target site, while the DBM directs the biodistribution
of non-targeted conjugate and metabolites thereof.

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Example XIII**Three-Step Pretargeting**

20 A patient has ovarian cancer. A monoclonal antibody
directed to an ovarian cancer cell antigen is conjugated
to biotin to form a MAB-biotin conjugate. The MAB-biotin
conjugate is administered to the patient in an amount in
excess of the maximum tolerated dose of conjugate
administerable in a targeted, direct label protocol and
is permitted to localize to target cancer cells for 24-
25 48 hours. Next, an amount of avidin sufficient to clear
non-targeted MAB-biotin conjugate and bind to the
targeted biotin is administered. A biotin-dextran-
radionuclide chelate conjugate of the type discussed in
Examples II-IV above is dispersed in a pharmaceutically
30 acceptable diluent and administered to the patient in a
therapeutically effective dose. The biotin-dextran-
radionuclide chelate conjugate localizes to the targeted
MAB-biotin-avidin moiety or is removed from the patient
via the renal pathway.

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Example XIV**Two-Step Pretargeting**

5 A patient has colon cancer. A monoclonal antibody directed to a colon cancer cell antigen is conjugated to streptavidin to form a MAb-streptavidin conjugate. The MAb-streptavidin conjugate is administered to the patient in an amount in excess of the maximum tolerated dose of conjugate administerable in a targeted, direct label protocol and is permitted to localize to target cancer cells for 24-48 hours. A biotin-DTPA-radionuclide chelate conjugate of the type discussed in Example VII above is dispersed in a pharmaceutically acceptable diluent and administered to the patient in a therapeutically effective dose. The biotin-DTPA-radionuclide chelate conjugate localizes to the targeted MAb-streptavidin moiety or is removed from the patient via the renal pathway.

20 While in the foregoing specification this invention has been described in relation to certain preferred embodiments thereof, and many details have been set forth for purposes of illustration, it will be apparent to those skilled in the art that the invention is susceptible to additional embodiments and that certain of the details described herein may be varied considerably without departing from the basic principles of the invention.

WHAT IS CLAIMED IS:

1. A method of directing the biodistribution of a small molecule in a diagnostic or therapeutic protocol for the treatment of a mammalian recipient, which method comprises administering to the recipient a conjugate having an improved biodistribution profile, the conjugate comprising:

a directed biodistribution molecule ranging between about 5 kD and about 70 kD and having a polarity facilitating excretion from the recipient via a renal pathway; and

a small molecule conjugated or complexed to the directed biodistribution molecule, the small molecule being of a structure that is generally not specifically excreted via the renal pathway and the small molecule is selected from the group comprising a chelate or a ligand, wherein the conjugate is excreted from the recipient by the renal pathway.

2. A method according to claim 1 wherein the directed biodistribution molecule comprises a polysaccharide selected from the group comprising a dextran, an inulin, a polyglutamate polymer, a hydroxypropylmethacrylamide polymer, or a succinylated polylysine polymer.

3. A method according to claim 2 wherein the small molecule comprises a diagnostically or therapeutically active agent.

4. A method according to claim 2 wherein the small molecule comprises biotin and an active agent, such that the biotin and the active agent are together associated with a single functional group of the directed biodistribution molecule or are separately associated with functional groups of the directed biodistribution molecule.

5. A method according to claim 2, further comprising

a targeting moiety conjugated to or complexed with the directed biodistribution molecule.

5 6. A method according to claim 5 wherein the targeting moiety is an antibody, an antibody fragment, a peptide, an oligonucleotide or analogs or derivatives thereof.

10 7. A method according to claim 1 wherein a plurality of small molecules are conjugated or complexed to the directed biodistribution molecule, the small molecules comprising one chelate and one ligand.

15 8. A method according to claim 7 wherein the small molecules conjugated or complexed to the directed biodistribution molecule comprise from about 2 to about 5 ligand molecules and from about 3 to about 20 chelate molecules.

9. A conjugate according to claim 1 wherein the directed biodistribution molecule has a molecular weight ranging from about 5 kD to about 15 kD.

20 10. A method of directing the biodistribution of a small molecule in a diagnostic or therapeutic protocol for the treatment of a mammalian recipient, which method comprises administering to the recipient a conjugate having an improved biodistribution profile, the conjugate comprising:

25 a directed biodistribution molecule having a molecular weight less than about 2 kD and having a polarity facilitating excretion from the recipient via a renal pathway; and

30 a small molecule conjugated or complexed to the directed biodistribution molecule, the small molecule being of a structure that is generally not specifically excreted via the renal pathway, wherein the conjugate is excreted from the recipient by the renal pathway.

35 11. A method according to claim 10 wherein the directed biodistribution molecule comprises a polyimino acetic acid selected from the group comprising

diethylene triamine-pentaacetic acid (DTPA), higher homologs and derivatives thereof.

12. A method according to claim 10 wherein the directed biodistribution molecule comprises a cyclic DTPA derivative or a 1,4,7,10-tetraazacyclododecane-N,N',N'',N'''-tetra acetic acid (DOTA) derivative, the DOTA derivative directed biodistribution molecule being bound to a chelate complexed with a gamma emitting radionuclide and, optionally, being complexed with a therapeutic radionuclide selected from the group comprising Y-90, Sm-153, Eu-169 or Lu-177.

13. A method according to claim 12 wherein the small molecule comprises a diagnostically or therapeutically active agent.

14. A method according to claim 12 wherein the small molecule comprises biotin and an active agent, such that the biotin and the active agent are together associated with a single functional group of the directed biodistribution molecule or are separately associated with functional groups of the directed biodistribution molecule.

15. A method according to claim 12, further comprising a targeting moiety conjugated to or complexed with the directed biodistribution molecule.

16. A method according to claim 15 wherein the targeting moiety is an antibody, an antibody fragment, a peptide, an oligonucleotide or analogs or derivatives thereof.

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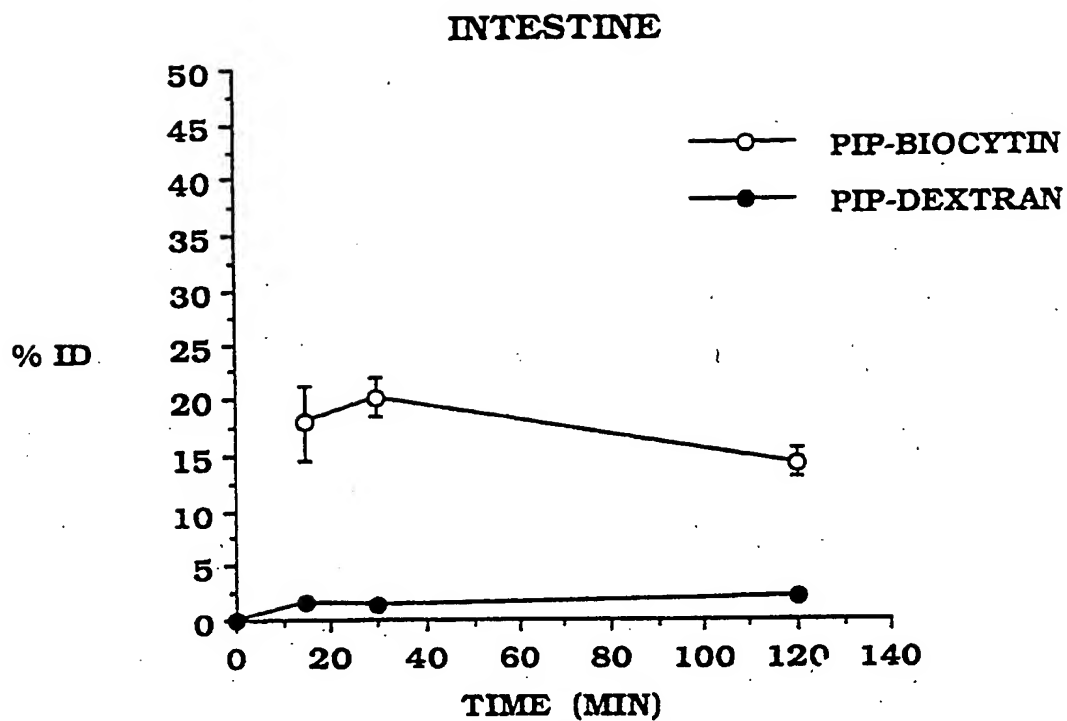


FIG. 1A

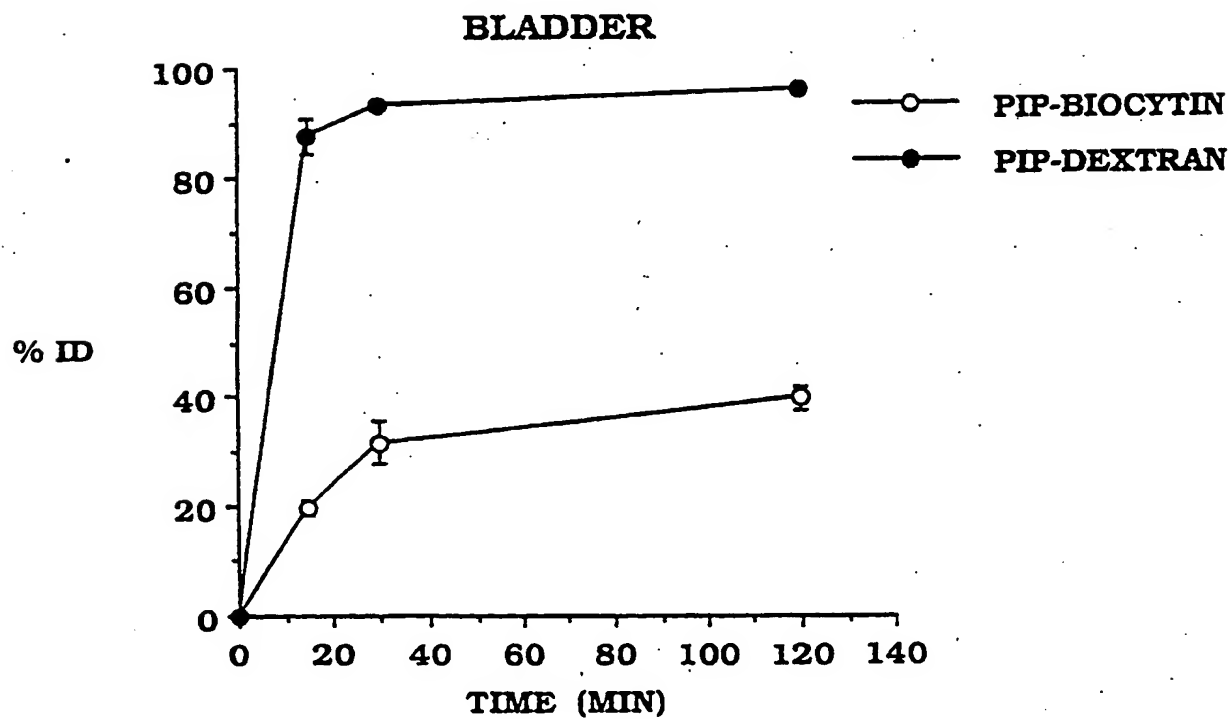


FIG. 1B

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LIVER

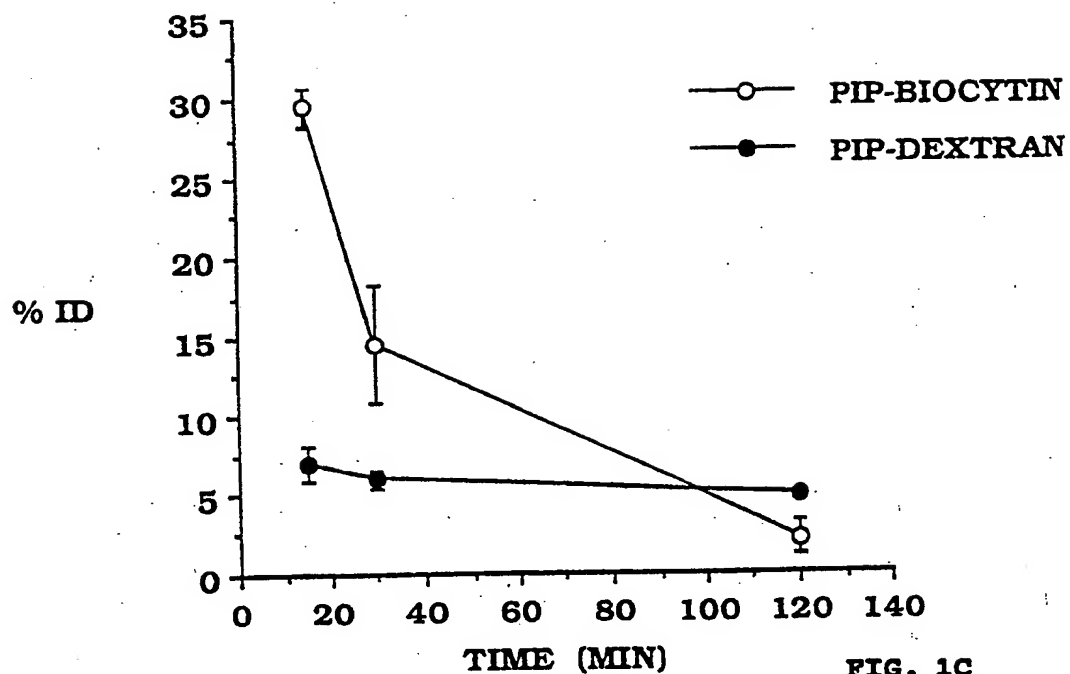


FIG. 1C

KIDNEY

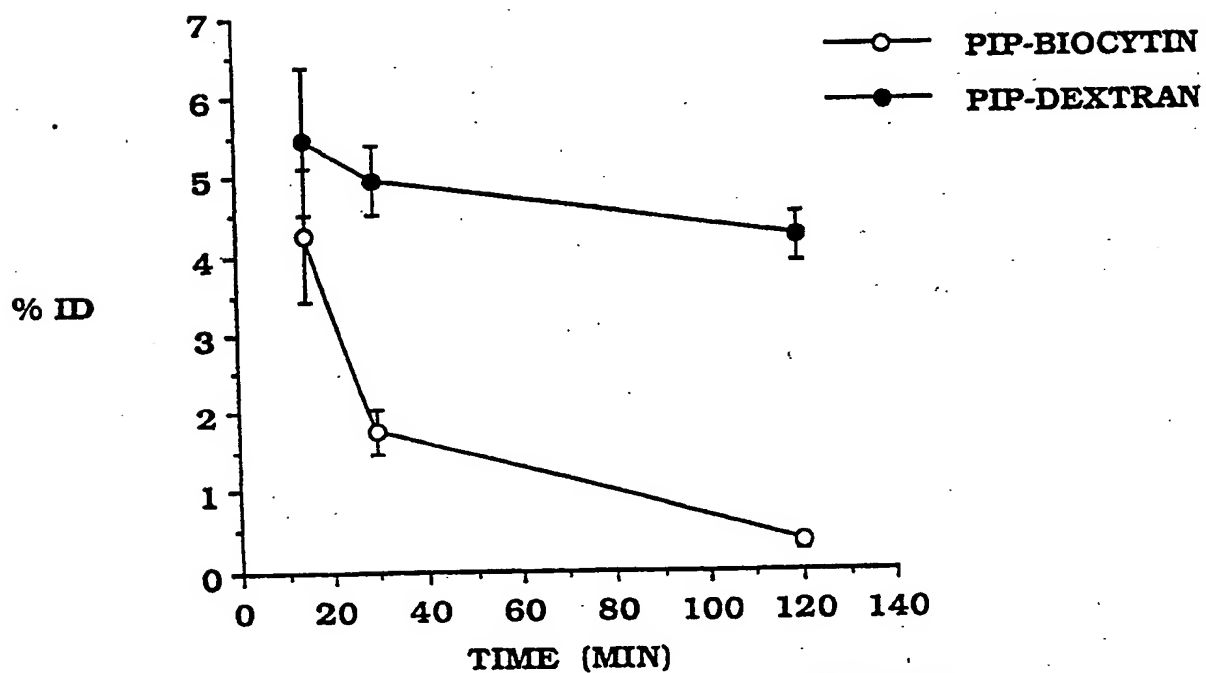


FIG. 1D

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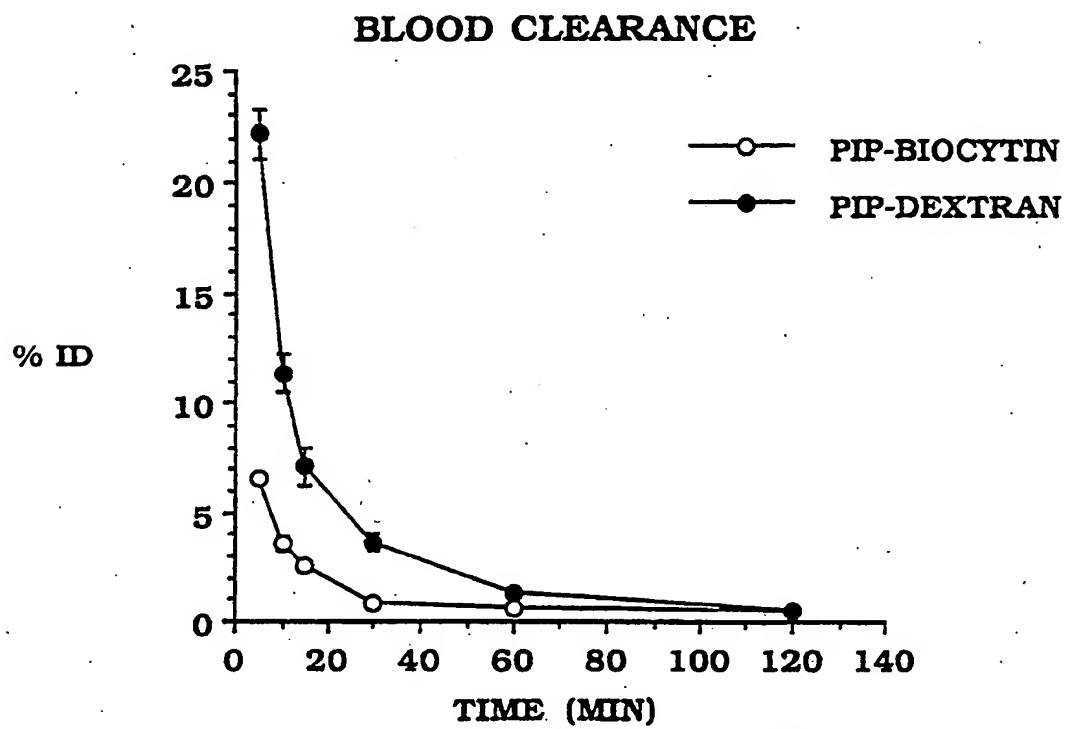


FIG. 1E

INTERNATIONAL SEARCH REPORT

Int. application No.
PC 4/01036

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) A61K 43/00, 49/00

US CL : 424/1.49, 1.53, 1.73; 514/59, 387; 530/391.3, 391.5, 391.7, 391.9

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 424/1.49, 1.53, 1.73; 514/59, 387; 530/391.3, 391.5, 391.7, 391.9

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS - "biotin", "dextran", "ab?", "radio?"

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US, A, 5,162,505 (DEAN ET AL) 10 NOVEMBER 1992 SEE COLUMNS 3-6	1-3, 5-13, 15-16
X	US, A, 5,057,313 (SHIH ET AL) 15 OCTOBER 1991 SEE ENTIRE DOCUMENT, ESP. COL. 4, LINES 22-33	1-3, 5-13, 15-16
X	US, A, 5,053,503 (DEAN ET AL) 01 OCTOBER 1991 SEE SUMMARY	10-13, 15-16
X	US, A, 4,985,233 (KLAVENESS ET AL) 15 JANUARY 1991 SEE ENTIRE DOCUMENT	1-3, 5-11

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be part of particular relevance	X*	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
I earlier document published on or after the international filing date	Y*	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
I document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)		
O document referring to an oral disclosure, use, exhibition or other means		
P document published prior to the international filing date but later than the priority date claimed	*G*	document member of the same patent family

Date of the actual completion of the international search
27 MAY 1994

Date of mailing of the international search report

17 JUN 1994

Name and mailing address of the ISA/US
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INTERNATIONAL SEARCH REPORT

International application No.
PCT/ISA/210 01036

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US, A, 4,855,353 (KURAMI ET AL) 08 AUGUST 1989 SEE COLUMNS 3-5	1-3, 5-13 15-16
X -- Y	US, A, 4,863,713 (GOODWIN ET AL) 05 SEPTEMBER 1989 SEE SUMMARY AND COLUMN 5	1-10 ----- 11-16
X -- Y	US, A, 4,849,208 (STAVRIANOPOULOS) 18 JULY 1989 SEE SUMMARY	10 ----- 1-9,11-16
X -- Y	US, A, 4,732,974 (NICLOTTI ET AL) 22 MARCH 1988	10 ----- 1-9,11-16
X -- Y	US, A, 4,385,046 (MILBRATH ET AL) 24 MAY 1983	1-3,7-10 ----- 4-6,11-16

